



ORIGINAL ARTICLE

Antitumor effect of guava leaves on lung cancer: A network pharmacology study



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Abstract Guava is known for its hypoglycemic, antiviral, antibacterial, anti-inflammatory, antioxidant, and antitumor properties. In this study, triterpenoids, sesquiterpenes, and flavonoids were examined as potential targets of constituents of guava leaves. Our study was aimed to reveal the antitumor mechanism and construct the network pharmacology network of guava leaf constituents and lung cancer. The potential targets of guava leaf constituents were searched in target databases, while the disease genes were searched in the GeneCards database. The common targets of drugs and diseases were screened out. A network map was constructed by the Cytoscape software, and the GO and KEGG pathways were analyzed. The existing cases were studied by SystemsDock molecular docking and cBioPortal tumor database study. Among the 66 chemical constituents of guava leaves, 153 of their targets were the lung cancer genes involved in many signaling pathways, such as the PI3K-Akt signaling pathway, in small cell lung cancer and non-small cell lung cancer. There was a binding activity between ligand compounds and receptor proteins. Guava leaves inhibited tumor through a gene regulatory network, and may play an important role in gene-targeting therapy. Through network pharmacology, we found that guava leaves had potential targets that interacted with various tumors, regulating the signaling pathways of cancers. This study preliminarily verified the pharmacological basis and the mechanism of the antitumor effect of guava leaves, providing a foundation for further research.

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1. Introduction

Guava (*Psidium guajava*), a member of the *Myrtaceae* family, is an evergreen shrub or small arbor with a wide range of habitats. Guava is found in countries in tropical or subtropical areas such as South America, Africa, and Southern Asia (Gutierrez et al., 2008; Feng et al., 2015). Guava leaves, also known as *Folium Psidii Guajavae*, are the dry leaves and leafy shoots of guava. The substances in guava leaves are triter-

penoids (Shao et al., 2012a), flavonoids, tannins (Seo et al., 2014), sesquiterpenes, miscellaneous quinones, volatile oils, and benzophenone glycosides. Guava is known for its hypoglycemic, antiviral, antibacterial, anti-inflammatory, antioxidant, and antitumor properties (Seo et al., 2014). These substances in guava leaves are of great research value. In a previous study, researchers identified novel types of aldehyde terpenes with their spectral characteristics and summarized the chemical structures of 17 heterodialdehyde compounds (Ouyang et al., 2015). Moreover, Psiguadial C and Psiguadial D showed significant biological activities, including the inhibitions of protein tyrosine phosphatase 1B (PTP1B) and human hepatoma cells (HepG2) (Shao et al., 2012b). Triterpenoids are known to exert antitumor effects (Song and Zhu, 2011; Lu et al., 2016). Flavonoids are one of the main functional components in guava leaves and have various pharmacological effects (Alnaqeb et al., 2019; Luo et al., 2019). Researchers extracted flavonoids from guava leaves and obtained approximately 9.89 mg/g of total flavonoids (Wang et al., 2016b). Flavonoids in plants are also known for their antitumor activity, which mainly involves regulation of immune function, repression of tumor cell adhesion and signal transmission, and inhibition of cellular proliferation and tumor angiogenesis (Kandaswami et al., 2005). In this study, triterpenoids, sesquiterpenes, and flavonoids were examined as potential targets of guava leaves.

Previous studies have investigated antitumor substances in guava leaves. It was found that guava leaf extracts exhibit potent antitumor activity (Ashraf et al., 2016) and play an inhibitory role in HeLa and Ec109 cells (Lee and Park, 2010).

Lung cancer has been the leading cause of cancer deaths among men since the early 1950s. A total of 1,824,701 lung cancer cases were estimated worldwide in 2012, accounting for nearly 32% for women and 68% for men (Rafieanesh et al., 2016). In contrast, a total of 1,589,925 lung cancer deaths were estimated in 2012, of which 31% were women and 69% were men (Rafieanesh et al., 2016). The number of new lung cancer cases has risen to 7,328,000, and 5,807,000 deaths occurred in China in 2013 (Chen et al., 2017). Lung cancer is the most common malignant tumor in China with high morbidity and mortality rates (Xing et al., 2019). Therefore, finding ways to treat lung cancer is of vital importance.

The concept of network pharmacology is based on multidisciplinary theories such as systems biology and multidirectional pharmacology (Boezio et al., 2017). Utilizing various techniques, such as omics, high-throughput screening, network visualization, and network analysis can help us better understand the molecular mechanism of diseases and the pharmacological mechanism of drugs from a multidimensional perspective (Wu and Wu, 2015; Danhof, 2016; Boezio et al., 2017). The method of network pharmacology and the databases available for research also tend to be diverse (Hu et al., 2014; Lee, 2015; Wang et al., 2019). It is straightforward in visual analysis of the results through target prediction, pharmacological mechanism research, active component research, and construction of network graphs (Boezio et al., 2017).

Herein, we aimed to construct the network pharmacology of constituents from guava leaves and investigate the potential of these constituents on lung cancer.

2. Materials and methods

2.1. Materials

The databases used in this study included TCMSP, PubChem, PharmMapper, STRING, UniProt, GeneCards, Venny 2.1.0, KOBAS 3.0, SystemsDock, DisGeNET, and CbioPortal. The software used included ChemDraw Office 2010 (PerkinElmer), Cytoscape 3.7.1 (Cytoscape), and FunRich 3.1.3 (<http://www.funrich.org/download>).

2.2. Methods

2.2.1. Collection of active constituents and chemical structures

Ouyang et al. (2015) systematically sorted out the compounds found in guava leaves through literature research; thus, these two documents were used as standards. The 2D or 3D structure of the target compound was searched in the PubChem (Kim et al., 2016) database, but for some of the compounds that were not included in the database, the chemical structure was drawn using the ChemBioDraw Ultra 12.0 software (PerkinElmer).

2.2.2. Screening of potential targets and acquisition of disease genes

Screening of potential targets: the TCMSP (Ru et al., 2014) database was used for the screening of potential targets, using “Chemical name” as the key word and the English name of the target compound as the potential target. PharmMapper (Liu et al., 2010) was used to screen potential targets. The 2D or 3D structures of the compound were used as input to screen potential targets. All potential targets of guava leaf constituents were converted to gene names using the STRING database and the UniProt database, and the species selected was *Homo Sapiens*.

2.2.3. Acquisition of disease genes

Through the human gene database GeneCards, using “Lung cancer” as a key word, disease target genes with greater correlation with lung cancer were extracted, and the first 500 genes (based on the descending order of the correlation score) were selected as disease genes.

The searched potential target genes were compared with the disease genes in the Venny database, and the common genes were selected to obtain potential targets for the treatment of lung cancer with guava leaf constituents.

2.2.4. Topological analysis of target protein networks and gene assignment

The STRING database was used for gene regulatory network construction based on potential targets of guava leaf constituents and lung cancer genes. The species was set to “*Homo sapiens*” and the minimum interaction threshold was set to 0.97. PPI network interaction maps of potential targets of guava leaf constituents derived from the database was downloaded. Network topology analysis was performed using the Cytoscape (Kohl et al., 2011) database. The gene type was assigned to the gene through the DisGeNET (Bauer-Mehren et al., 2010) database, and the protein/gene was sequentially input for retrieval of the related gene and the target type

(protein class) information. Interactions between compounds and target proteins were analyzed by constructing a network map of “Guava leaf - constituent category - active constituent - gene”.

2.2.5. GO and KEGG enrichment analysis

GO analysis was performed using the ClueGO plug-in in the Cytoscape software, and the gene symbols of targets of guava leaf constituent for lung cancer were input into ClueGO for gene ontology (GO) enrichment analysis (Biology Process, Molecular Function and Cellular Component). The gene symbols of targets of guava leaf constituent for lung cancer were converted into Entrez ID by the Funrich software, and KEGG pathway enrichment analysis was performed using KOBAS 3.0.

2.2.6. Molecular docking

The 10 genes that were relatively strong in the network diagram of “Guava leaf - constituent category - active constituent - gene” were molecularly docked with five compounds by systemsDock (Hsin et al., 2013; Hsin et al., 2016).

2.2.7. cBioportal analysis

The gene expression of existing case samples in the database was analyzed by cBioportal. cBioportal was financially funded by the Memorial Sloan-Kettering Cancer Center. It mainly addresses a large number of data problems obtained from large sample tumor genomic studies so that the results can be easily explored and directly applied to oncology (Wu et al., 2019).

2.2.8. Gene expression analysis and pathway activity analysis

The mRNA expression and the pathway activity of genes of interest were analyzed in the GSCALite database (<http://bioinfo.life.hust.edu.cn/web/GSCALite/>) following the instructions provided in this platform.

3. Results

3.1. Collection of active ingredients and their chemical structures

The 2D or 3D structures of all compounds were searched in the PubChem database, and the files in the sdf format were saved. Compounds not included in the database were identified by chemical structure, which were then drawn using the ChemDraw Ultra 12.0 software, and the sdf file was saved (Fig. 1). The structures of the compounds were searched by PubChem, and their molecular formulas and accession numbers in the database (PubChem CID) were retrieved. The relevant information of the active constituents of Guava leaves is listed in Table 1. In total, 66 active components of guava leaves, including 17 triterpenoids such as ursane and oleanane pentacyclic triterpenes, 19 sesquiterpenoids, and 30 flavonoids.

3.2. Screening of potential targets and acquisition of disease genes

A total of 115 potential targets for four triterpenoids and 303 potential targets for the 19 flavonoids were obtained from the TCMSP database (Table 2). Molecular information on guava

leaves, including Lipinski's “five-law” parameters, namely relative molecular mass (MV), octanol–water partition coefficient (AlogP), possible hydrogen bond donor number (Hdon), possible hydrogen, number of bond receptors (Hacc), number of bonds allowed to rotate freely (RBN), oral bioavailability (OB), and drug-like degree (DL) were obtained. The targets of the active ingredients of guava leaves were obtained from the PharmMapper database, and targets with z' -score ≥ 1 were screened as potential targets of guava leaf active ingredients. From the definition in the database website, “Fit Score” and “ z' -score” are scores generated by the metric's Fit score, which is a pre-calculated library score matrix, and a large positive z' -score represents the target-to-query combination. A total of 198 target genes for triterpenoids, 215 target genes for sesquiterpenoids, and 302 target genes for flavonoids were retrieved from the PharmMapper database. A total of 246 potential targets for triterpenoids, 215 potential targets for sesquiterpenoids, and 535 potential targets for flavonoids were obtained from TCMSP and PharmMapper. The names of all potential targets were imported into the STRING database and Uniprot database, and converted into Gene Symbols.

In the Genecards database, with “Lung Cancer” as the keyword, a total of 20,649 results were associated with lung cancer, and the top 500 targets with “Score” values ranged in descending order.

The genes associated with the three types of compounds were compared with the lung cancer genes in the Venny 2.1.0 database, and a chart of gene interaction was obtained (Fig. 2A). A total of 153 genes were obtained from lung cancer, and the target genes of guava leaf constituents against lung cancer were obtained. The total number of genes was 16.4%. Among them, there were 4 (0.4%) cross-reactive genes in three sputum and lung cancer; 68 (7.3%) cross-genes between flavonoids and lung cancer; 3 (0.3%) genes between triterpenoids, sesquiterpenes, and lung cancer; 9 (1%) cross-linking genes for scorpion, flavonoids, and lung cancer; 27 (2.9%) cross-linking genes for triterpenoids, flavonoids, and lung cancer; and 42 (4.5%) cross-linking genes for triterpenoids, sesquiterpenes, flavonoids, and lung cancer.

3.3. Topological analysis of target protein networks and gene assignment

The 153 targets related to lung cancer and identified as targets for guava leaf constituents were introduced into the STRING database to obtain a protein interaction network. Topological analysis of the target protein network was performed in Cytoscape. The topological analysis results are shown in Fig. 2B. The color depth and size of the nodes represent the strength of interaction between the gene and other genes. The interaction network had a total of 153 number of nodes. The number of edges associated with the target protein was 443, and the average node degree was 5.79, with p value (PPI enrichment p -value) $< 1.0e-16$. The target genes with a degree greater than or equal to 10 in the guava leaf constituent and lung cancer cross-linking network topology analysis were sequentially introduced into the DisGeNET database to obtain the protein type corresponding to the target. The results showed that the types of these proteins were nucleic acid binding, transcription factor, calcium-binding protein, kinase, transferase, signaling molecule, enzyme modulator, receptor, hydrolase, protease,

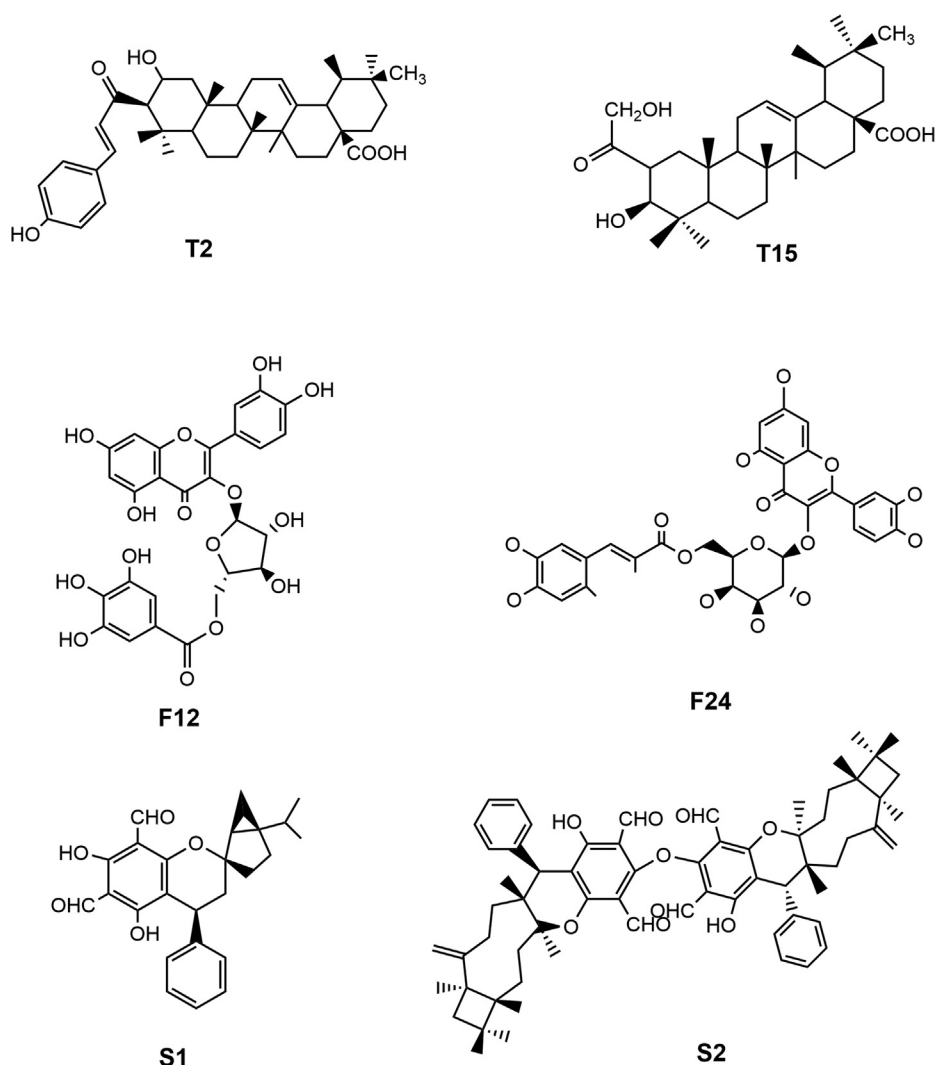


Fig. 1 The structural formula of six compounds, which were found but not included in the PubChem database. T2: 3 β -O-trans-p-coumaroylmaslinic acid; T15: psidiumoic acid; F12: guavinoside C; F24: quercetin-3-O-(6''-feruloyl); S1: Diguajadiol; S2: Guadiol A.

and transfer/carrier protein (Table 3). The network topological analysis of genes associated with lung cancer in guava leaves genes with a score of 10 or higher and their genes are also shown in Table 3.

3.4. Cytoscape network interaction analysis

There is a genetic interaction between drugs and diseases. This is a characteristic of multi-component and multi-target network pharmacology analysis. Through Cytoscape network visualization analysis, a “Guava leaf-compound class-active ingredient-gene” interaction network was obtained. The compound and target information on nodes ≥ 20 are listed in Table 4, where the Average Shortest Path Length is the average shortest path, Closeness Centrality is the center proximity, and Radiality is the radial degree.

As shown in Fig. 3 and Table 4, the three compounds in guava leaves that interacted strongly with lung cancer were quercetin, genistein, and apigenin. Moreover, the top ten targets were vitamin D3 receptor (VDR), cyclin-dependent kinase 2 (CDK2), dual specificity mitogen-activated protein kinase 1

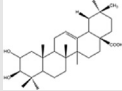
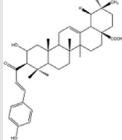
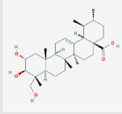
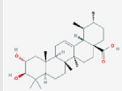
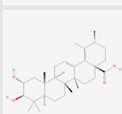
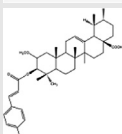
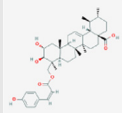
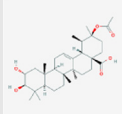
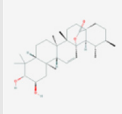
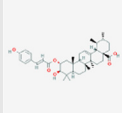
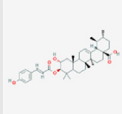
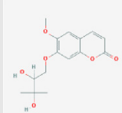
(MAP2K1), cyclin-dependent kinase 6 (CDK6), hepatocyte growth factor receptor (MET), tyrosine-protein kinase ABL1 (ABL1), peroxisome proliferator-activated receptor gamma (PPARG), interleukin-2 (IL2), epidermal growth factor receptor (EGFR), and progesterone receptor (PGR).

The network diagram of “Guava leaf-compound class-active constituents-gene” generated by Cytoscape is shown in Fig. 3, in which the green rectangle shape represents the traditional Chinese medicine guava leaf, and the three pale blue diamond shapes represent sesquiterpene, flavonoids, and triterpenoids; pink ovals represent compounds and yellow hexagons represent genes for the treatment of lung cancer with guava leaves. In the interaction network, the target information corresponding to the compound having a degree value of > 20 and the corresponding gene are listed in Table 5.

3.5. GO and KEGG enrichment analysis

Through ClueGO analysis, a total of 1430 GO biological processes were obtained, with 7536 interactions between the biological processes; 35 molecular functions, with 28

Table 1 Molecular information of guava leaf constituents. The first column is the compound name, the second column is the molecular type of the compound, the third column is the number of the constituent, with triterpenoids numbered from T1 to T17, flavonoids from F1 to F30, and sesquiterpenoids from S1 to S19. The molecular formula, the molecular structure, the PubChem ID for compound retrieved from the database and the Pharmmapper JOB ID (the search number of targets in the Pharmmapper database) were listed in the columns four to seven.

| Compound Name | Molecular type | Number | Molecular Formula | Molecular structure | PubChem CID | Pharmmapper JOB ID |
|---|----------------|--------|-------------------|--|-------------|--------------------|
| 2 α -hydroxyoleanolic acid | Triterpenoid | T1 | C30H48O4 |  | 15,560,128 | 190,417,034,302 |
| 3 β -O-trans-p-coumaroylmaslinic acid | Triterpenoid | T2 | C39H54O5 |  | / | 190,417,034,505 |
| asiatic acid | Triterpenoid | T3 | C30H48O5 |  | 119,034 | 190,417,034,630 |
| corosolic acid | Triterpenoid | T4 | C30H48O4 |  | 6,918,774 | 190,417,034,747 |
| goreishic acid I | Triterpenoid | T5 | C30H46O4 |  | 3,081,756 | 190,318,032,131 |
| guajanoic acid | Triterpenoid | T6 | C32H50O6 |  | 101,211,343 | 190,417,035,053 |
| guavacoumaric acid | Triterpenoid | T7 | C39H54O7 |  | 101,211,344 | 190,417,035,342 |
| guavanoic acid | Triterpenoid | T8 | C32H50O6 |  | 101,211,343 | 190,417,035,431 |
| ilelatifol D | Triterpenoid | T9 | C30H46O4 |  | 102,572,108 | 190,318,031,159 |
| isoneriucoumaric acid | Triterpenoid | T10 | C39H54O6 |  | 10,100,394 | 190,417,035,723 |
| jacoumaric acid | Triterpenoid | T11 | C39H54O6 |  | 11,700,083 | 190,417,035,813 |
| obtusinin | Triterpenoid | T12 | C15H18O6 |  | 3,604,942 | 190,318,030,756 |

(continued on next page)

Table 1 (continued)

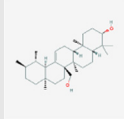
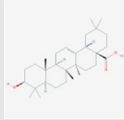
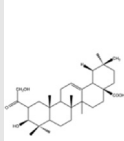
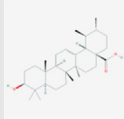
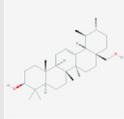
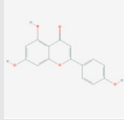
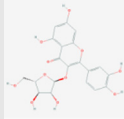
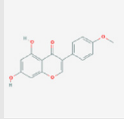
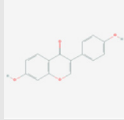
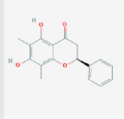
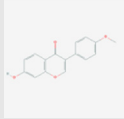
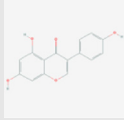
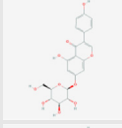
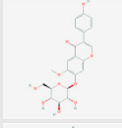
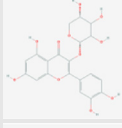
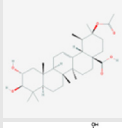
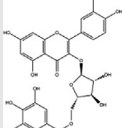
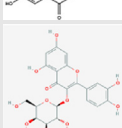
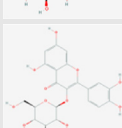
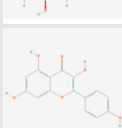
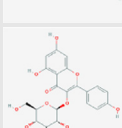
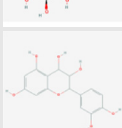
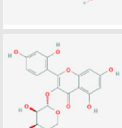

| Compound Name | Molecular type | Number | Molecular Formula | Molecular structure | PubChem CID | Pharmmapper JOB ID |
|---|----------------|--------|---|--|-------------|--------------------|
| obtusol (3 β , 27-dihydroxy-urs-12-ene) | Triterpenoid | T13 | C ₃₀ H ₅₀ O ₂ |  | 15,895,316 | 190,417,040,026 |
| oleanolic acid | Triterpenoid | T14 | C ₃₀ H ₄₈ O ₃ |  | 10,494 | 190,417,040,126 |
| psidiumoic acid | Triterpenoid | T15 | C ₃₂ H ₅₀ O ₅ |  | / | 190,417,040,228 |
| ursolic acid | Triterpenoid | T16 | C ₃₀ H ₄₈ O ₃ |  | 64,945 | 190,417,040,330 |
| uvoal | Triterpenoid | T17 | C ₃₀ H ₅₀ O ₂ |  | 92,802 | 190,417,040,432 |
| Apigenin | Flavonoid | F1 | C ₁₅ H ₁₀ O ₅ |  | 5,280,443 | 190,414,093,403 |
| Avicularin | Flavonoid | F2 | C ₂₀ H ₁₈ O ₁₁ |  | 5,490,064 | 190,414,093,603 |
| Biochanin | Flavonoid | F3 | C ₁₆ H ₁₂ O ₅ |  | 5,280,373 | 190,403,023,437 |
| Daidzein | Flavonoid | F4 | C ₁₅ H ₁₀ O ₄ |  | 5,281,708 | 190,414,094,000 |
| demthoxymatteucinol | Flavonoid | F5 | C ₁₇ H ₁₆ O ₄ |  | 180,550 | 190,414,094,140 |
| formononetin | Flavonoid | F6 | C ₁₆ H ₁₂ O ₄ |  | 5,280,378 | 190,414,094,323 |
| Genistein | Flavonoid | F7 | C ₁₅ H ₁₀ O ₅ |  | 5,280,961 | 190,414,094,458 |

Table 1 (continued)

| Compound Name | Molecular type | Number | Molecular Formula | Molecular structure | PubChem CID | Pharmmapper JOB ID |
|---------------------------------------|----------------|--------|---|--|-------------|--------------------|
| Genistin | Flavonoid | F8 | C ₂₁ H ₂₀ O ₁₀ |  | 5,281,377 | 190,414,094,600 |
| Glycitin/daidzin | Flavonoid | F9 | C ₂₂ H ₂₂ O ₁₀ |  | 187,808 | 190,414,094,811 |
| guaijaverin | Flavonoid | F10 | C ₂₀ H ₁₈ O ₁₁ |  | 5,481,224 | 190,414,095,056 |
| guavaric A | Flavonoid | F11 | C ₃₂ H ₅₀ O ₆ |  | 101,211,343 | 190,414,095,209 |
| guavinoside C | Flavonoid | F12 | C ₂₇ H ₂₂ O ₁₅ |  | / | 190,414,095,324 |
| hyperin | Flavonoid | F13 | C ₂₁ H ₂₀ O ₁₂ |  | 133,568,467 | 190,417,041,614 |
| isoquercetin(Isoquercitrin) | Flavonoid | F14 | C ₂₁ H ₂₀ O ₁₂ |  | 5,280,804 | 190,414,095,613 |
| kaempferol | Flavonoid | F15 | C ₁₅ H ₁₀ O ₆ |  | 5,280,863 | 190,414,095,757 |
| kaempferol-3-glucoside | Flavonoid | F16 | C ₂₁ H ₂₀ O ₁₁ |  | 5,282,102 | 190,414,095,928 |
| Leucocyanidin | Flavonoid | F17 | C ₁₅ H ₁₄ O ₇ |  | 71,629 | 190,414,100,124 |
| morin-3-O- α -L-lyxopyranoside | Flavonoid | F18 | C ₂₀ H ₁₈ O ₁₁ |  | 10,455,578 | 190,414,100,422 |
| myricetin | Flavonoid | F19 | C ₁₅ H ₁₀ O ₈ |  | 5,281,672 | 190,414,100,546 |

(continued on next page)

Table 1 (continued)

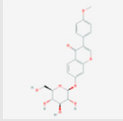
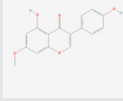
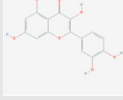
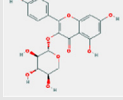
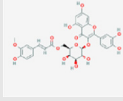
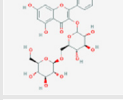
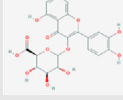
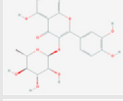
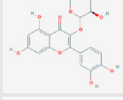
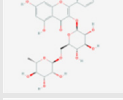
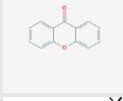
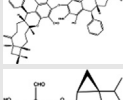
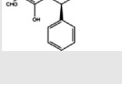
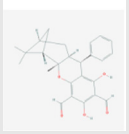
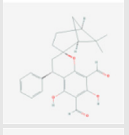
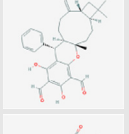
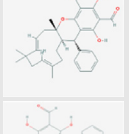
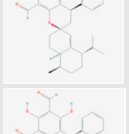
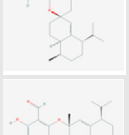
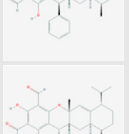
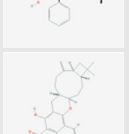
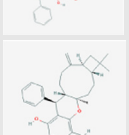
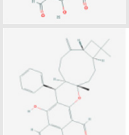

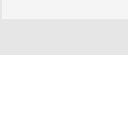
| Compound Name | Molecular type | Number | Molecular Formula | Molecular structure | PubChem CID | Pharmmapper JOB ID |
|--|------------------|--------|---|--|-------------|--------------------|
| Ononin | Flavonoid | F20 | C ₂₂ H ₂₂ O ₉ |  | 442,813 | 190,414,100,718 |
| prunetin | Flavonoid | F21 | C ₁₆ H ₁₂ O ₅ |  | 5,281,804 | 190,414,100,847 |
| quercetin | Flavonoid | F22 | C ₁₅ H ₁₀ O ₇ |  | 5,280,343 | 190,414,101,024 |
| quercetin-3-O-β-D-xylopyranoside | Flavonoid | F23 | C ₂₀ H ₁₈ O ₁₁ |  | 5,320,861 | 190,414,101,158 |
| quercetin-3-O-(6''-feruloyl)-β-D-galactopyranoside | Flavonoid | F24 | C ₃₁ H ₂₈ O ₁₅ |  | / | 190,414,101,321 |
| quercetin-3-O-gentiobioside | Flavonoid | F25 | C ₂₇ H ₃₀ O ₁₇ |  | 13,915,963 | 190,414,101,504 |
| quercetin-3-O-β-D-glucuronide | Flavonoid | F26 | C ₂₁ H ₁₈ O ₁₃ |  | 13,258,914 | 190,414,101,626 |
| quercitrin | Flavonoid | F27 | C ₂₁ H ₂₀ O ₁₁ |  | 5,280,459 | 190,414,101,724 |
| reynoutrin | Flavonoid | F28 | C ₂₀ H ₁₈ O ₁₁ |  | 5,320,863 | 190,414,101,824 |
| rutin | Flavonoid | F29 | C ₂₇ H ₃₀ O ₁₆ |  | 5,280,805 | 190,414,101,941 |
| xanthone | Flavonoid | F30 | C ₁₃ H ₈ O ₂ |  | 7020 | 190,414,102,035 |
| Diguajadial | Sesquiterpenoids | S1 | C ₆₀ H ₆₆ O ₉ |  | / | 190,321,031,034 |
| Guadial A | Sesquiterpenoids | S2 | C ₂₅ H ₂₆ O ₅ |  | / | 190,321,032,624 |

Table 1 (continued)

| Compound Name | Molecular type | Number | Molecular Formula | Molecular structure | PubChem CID | Pharmmapper JOB ID |
|---------------|------------------|--------|--|--|-------------|--------------------|
| Guadial B | Sesquiterpenoids | S3 | C ₂₅ H ₂₆ O ₅ |  | 122,377,745 | 190,319,051,501 |
| Guadial C | Sesquiterpenoids | S4 | C ₂₅ H ₂₆ O ₅ |  | 122,377,746 | 190,319,051,610 |
| Guajadial | Sesquiterpenoids | S5 | C ₃₀ H ₃₄ O ₅ |  | 101,447,677 | 190,318,025,632 |
| Guajadial B | Sesquiterpenoids | S6 | C ₃₀ H ₃₄ O ₅ |  | 137,346,032 | 190,319,050,244 |
| Guajadial C | Sesquiterpenoids | S7 | C ₃₀ H ₃₄ O ₅ |  | 134,714,902 | 190,319,050,403 |
| Guajadial D | Sesquiterpenoids | S8 | C ₃₀ H ₃₄ O ₅ |  | 134,714,901 | 190,319,050,732 |
| Guajadial E | Sesquiterpenoids | S9 | C ₃₀ H ₃₄ O ₅ |  | 134,714,904 | 190,319,050,851 |
| Guajadial F | Sesquiterpenoids | S10 | C ₃₀ H ₃₄ O ₅ |  | 134,714,903 | 190,319,050,938 |
| Guapsidial A | Sesquiterpenoids | S11 | C ₂₉ H ₃₂ O ₅ |  | 122,377,744 | 190,319,051,859 |
| Guajadial | Sesquiterpenoids | S12 | C ₃₀ H ₃₄ O ₅ |  | 46,197,930 | 190,320,031,551 |
| Psidial A | Sesquiterpenoids | S13 | C ₃₀ H ₃₆ O ₆ |  | 45,104,960 | 190,318,025,129 |
| Psidial B | Sesquiterpenoids | S14 | C ₃₀ H ₃₆ O ₆ |  | 45,104,961 | 190,318,025,249 |

(continued on next page)

Table 1 (continued)

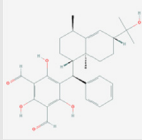
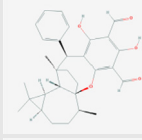
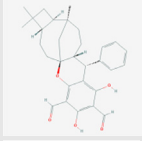
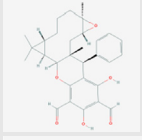
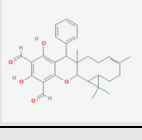
| Compound Name | Molecular type | Number | Molecular Formula | Molecular structure | PubChem CID | PharmMapper JOB ID |
|---------------|------------------|--------|--|--|-------------|--------------------|
| Psidial C | Sesquiterpenoids | S15 | C ₃₀ H ₃₄ O ₅ |  | 49,844,493 | 190,318,025,408 |
| Psiguadial A | Sesquiterpenoids | S16 | C ₃₀ H ₃₄ O ₅ |  | 49,844,493 | 190,318,025,837 |
| Psiguadial B | Sesquiterpenoids | S17 | C ₃₀ H ₃₄ O ₅ |  | 102,052,649 | 190,318,025,954 |
| Psiguadial C | Sesquiterpenoids | S18 | C ₃₀ H ₃₄ O ₆ |  | 122,224,646 | 190,319,051,306 |
| Psiguadial D | Sesquiterpenoids | S19 | C ₃₀ H ₃₄ O ₅ |  | 77,984,632 | 190,321,030,934 |

Table 2 TCMSP parameter information.

| Compound Name | Mol ID | MW | AlogP | Hdon | Hacc | RBN | OB (%) | DL |
|-------------------------------|-----------|--------|-------|------|------|-----|--------|------|
| Apigenin | MOL000008 | 270.25 | 2.33 | 3 | 5 | 1 | 23.06 | 0.21 |
| Avicularin | MOL007979 | 434.38 | -0.08 | 7 | 11 | 4 | 2.06 | 0.7 |
| Daidzein | MOL000390 | 254.25 | 2.33 | 2 | 4 | 1 | 19.44 | 0.19 |
| formononetin | MOL000392 | 268.28 | 2.58 | 1 | 4 | 2 | 69.67 | 0.21 |
| Genistein | MOL000481 | 270.25 | 2.07 | 3 | 5 | 1 | 17.93 | 0.21 |
| Genistin | MOL000480 | 432.41 | 0.16 | 6 | 10 | 4 | 13.35 | 0.75 |
| daidzin | MOL009720 | 416.41 | 0.43 | 5 | 9 | 4 | 14.32 | 0.73 |
| guaijaverin | MOL000702 | 434.38 | -0.08 | 7 | 11 | 3 | 29.65 | 0.7 |
| hyperin | MOL004368 | 464.41 | -0.59 | 8 | 12 | 4 | 6.94 | 0.77 |
| isoquercetin | MOL000437 | 302.25 | 0.34 | 5 | 7 | 1 | 5.92 | 0.28 |
| kaempferol | MOL000422 | 286.25 | 1.77 | 4 | 6 | 1 | 41.88 | 0.24 |
| kaempferol-3-O-β-D-glucoside | MOL001415 | 448.41 | -0.32 | 7 | 11 | 4 | 2.77 | 0.74 |
| Leucocyanidin | MOL007214 | 306.29 | 1.09 | 6 | 7 | 1 | 37.61 | 0.27 |
| myricetin | MOL002008 | 318.25 | 1.24 | 6 | 8 | 1 | 13.75 | 0.31 |
| Ononin | MOL000391 | 430.44 | 0.68 | 4 | 9 | 5 | 11.52 | 0.78 |
| prunetin | MOL000486 | 284.28 | 2.32 | 2 | 5 | 2 | 5.41 | 0.24 |
| quercetin | MOL000098 | 302.25 | 1.5 | 5 | 7 | 1 | 46.43 | 0.28 |
| quercetin-3-O-β-D-glucuronide | MOL001001 | 450.38 | -0.42 | 8 | 12 | 3 | 30.66 | 0.74 |
| quercitrin | MOL000701 | 448.41 | 0.3 | 7 | 11 | 3 | 4.04 | 0.74 |
| rutin | MOL000415 | 610.57 | -1.45 | 10 | 16 | 6 | 3.2 | 0.68 |
| 2α-hydroxyoleanolic acid | MOL012969 | 471.77 | 4.78 | 2 | 4 | 1 | 17.38 | 0.74 |
| asiatic acid | MOL006861 | 488.78 | 4.41 | 4 | 5 | 2 | 16.69 | 0.72 |
| oleanolic acid | MOL000263 | 456.78 | 6.42 | 2 | 3 | 1 | 29.02 | 0.76 |
| ursolic acid | MOL000511 | 456.78 | 6.47 | 2 | 3 | 1 | 16.77 | 0.75 |
| olmelin | MOL000510 | 284.28 | 2.32 | 2 | 5 | 2 | 25.21 | 0.24 |

interactions between the molecular functions; and 203 GO cell components, with 470 interactions between the cell components. KEGG analysis was performed by KOBAS 3.0, and

217 pathway enrichment results were obtained. GO analysis results of the interacting genes between guava leaves and lung cancer are shown in Table 6.

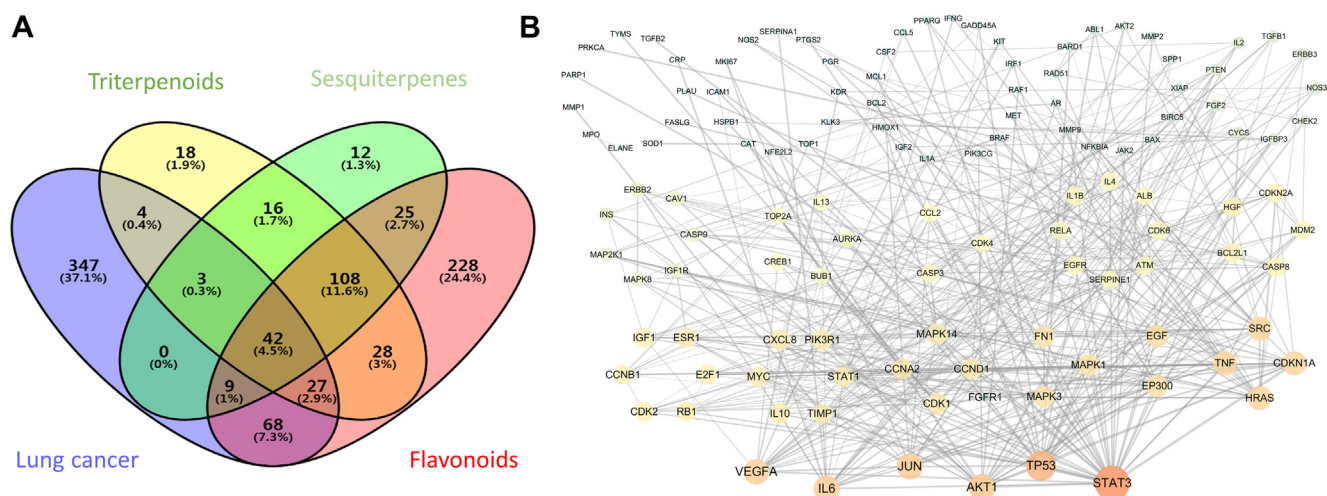


Fig. 2 Drug-disease interaction network analysis (A) Screening of guava leaf-lung cancer common gene. (B) Topological analysis of drug-disease interactive gene network.

Table 3 Topology analysis data of drug-disease interaction gene (Degree ≥ 10).

| Gene | Full Name | Degree | Uniprot ID | Protein class |
|--------|--|--------|------------|---|
| STAT3 | signal transducer and activator of transcription 3 | 35 | P40763 | nucleic acid binding; transcription factor |
| TP53 | tumor protein p53 | 27 | P04637 | transcription factor |
| AKT1 | AKT serine/threonine kinase 1 | 22 | P31749 | calcium-binding protein; kinase; transferase; transfer/carrierprotein |
| IL6 | interleukin 6 | 21 | P05231 | None |
| JUN | Transcription factor AP-1 | 21 | P05412 | nucleic acid binding; transcription factor |
| VEGFA | vascular endothelial growth factor A | 20 | P15692 | signaling molecule |
| SRC | non-receptor tyrosine kinase | 18 | P12931 | None |
| TNF | tumor necrosis factor | 18 | P01375 | signaling molecule |
| HRAS | HRas proto-oncogene, GTPase | 18 | P01112 | enzyme modulator |
| CDKN1A | cyclin dependent kinase inhibitor 1A | 18 | P38936 | None |
| EGF | epidermal growth factor | 15 | P01133 | None |
| EP300 | E1A binding protein p300 | 15 | Q09472 | nucleic acid binding; transcription factor; transferase |
| FN1 | fibronectin 1 | 14 | P02751 | signaling molecule |
| MAPK3 | mitogen-activated protein kinase 3 | 14 | P27361 | kinase; transferase |
| MAPK1 | mitogen-activated protein kinase 1 | 14 | P28482 | kinase; transferase |
| MAPK14 | mitogen-activated protein kinase 14 | 13 | Q16539 | kinase; transferase |
| CCNA2 | cyclin A2 | 13 | P20248 | enzyme modulator |
| CDK1 | cyclin dependent kinase 1 | 13 | P06493 | kinase; transferase |
| CCND1 | cyclin D1 | 13 | P24385 | enzyme modulator |
| PIK3R1 | phosphoinositide-3-kinase regulatory subunit 1 | 12 | P27986 | enzyme modulator |
| CXCL8 | C-X-C motif chemokine ligand 8 | 12 | P10145 | signaling molecule |
| MYC | MYC proto-oncogene | 12 | P01106 | nucleic acid binding; transcription factor |
| IL10 | interleukin 10 | 12 | P22301 | None |
| TIMP1 | TIMP metalloproteinase inhibitor 1 | 12 | P01033 | enzyme modulator |
| STAT1 | signal transducer and activator of transcription 1 | 12 | P42224 | nucleic acid binding; transcription factor |
| ESR1 | estrogen receptor 1 | 11 | P03372 | nucleic acid binding; receptor; transcription factor |
| IGF1 | insulin like growth factor 1 | 11 | P05019 | None |
| CCNB1 | cyclin B1 | 11 | P14635 | enzyme modulator |
| CDK2 | cyclin dependent kinase 2 | 11 | P24941 | kinase; transferase |
| RB1 | RB transcriptional corepressor 1 | 11 | P06400 | nucleic acid binding; transcription factor |
| E2F1 | E2F transcription factor 1 | 11 | Q01094 | nucleic acid binding; transcription factor |
| CDKN2A | cyclin dependent kinase inhibitor 2A | 10 | P42771 | None |
| HGF | hepatocyte growth factor | 10 | P14210 | hydrolase; protease |
| BCL2L1 | BCL2 like 1 | 10 | Q07817 | signaling molecule |
| CASP8 | caspase 8 | 10 | Q14790 | enzyme modulator; hydrolase; protease |
| MDM2 | MDM2 proto-oncogene | 10 | Q00987 | nucleic acid binding |

Table 4 Interaction network data of “guava leaf – compound class – active component – gene”(Degree ≥ 20).

| Node Name | Degree | Average Shortest Path Length | Closeness Centrality | Radiality |
|---------------------|--------|------------------------------|----------------------|------------|
| quercetin | 85 | 1.96396396 | 0.50917431 | 0.80720721 |
| genistein | 62 | 2.15315315 | 0.46443515 | 0.76936937 |
| apigenin | 53 | 2.22522523 | 0.44939271 | 0.75495495 |
| ursolic acid | 43 | 2.31531532 | 0.43190661 | 0.73693694 |
| daidzein | 38 | 2.37837838 | 0.42045455 | 0.72432432 |
| kaempferol | 33 | 2.45045045 | 0.40808824 | 0.70990991 |
| myricetin | 30 | 2.45045045 | 0.40808824 | 0.70990991 |
| rutin | 22 | 2.6036036 | 0.38408304 | 0.67927928 |
| formononetin | 21 | 2.53153153 | 0.39501779 | 0.69369369 |
| demthoxymatteucinol | 21 | 2.51351351 | 0.39784946 | 0.6972973 |
| Guadial B | 20 | 2.56756757 | 0.38947368 | 0.68648649 |
| VDR | 36 | 2.44594595 | 0.40883978 | 0.71081081 |
| CDK2 | 29 | 2.31981982 | 0.43106796 | 0.73603604 |
| MAP2K1 | 27 | 2.6981982 | 0.3706177 | 0.66036036 |
| CDK6 | 26 | 2.18468468 | 0.45773196 | 0.76306306 |
| MET | 25 | 2.90540541 | 0.34418605 | 0.61891892 |
| ABL1 | 24 | 2.42792793 | 0.41187384 | 0.71441441 |
| PPARG | 22 | 2.28378378 | 0.43786982 | 0.74324324 |
| IL2 | 22 | 2.37387387 | 0.42125237 | 0.72522523 |
| EGFR | 22 | 2.3018018 | 0.43444227 | 0.73963964 |
| PGR | 21 | 2.40990991 | 0.41495327 | 0.71801802 |
| MAPK14 | 21 | 2.58108108 | 0.38743455 | 0.68378378 |
| KIT | 21 | 2.77927928 | 0.35980551 | 0.64414414 |
| RARB | 20 | 2.77927928 | 0.35980551 | 0.64414414 |
| NOS2 | 20 | 2.53603604 | 0.39431616 | 0.69279279 |

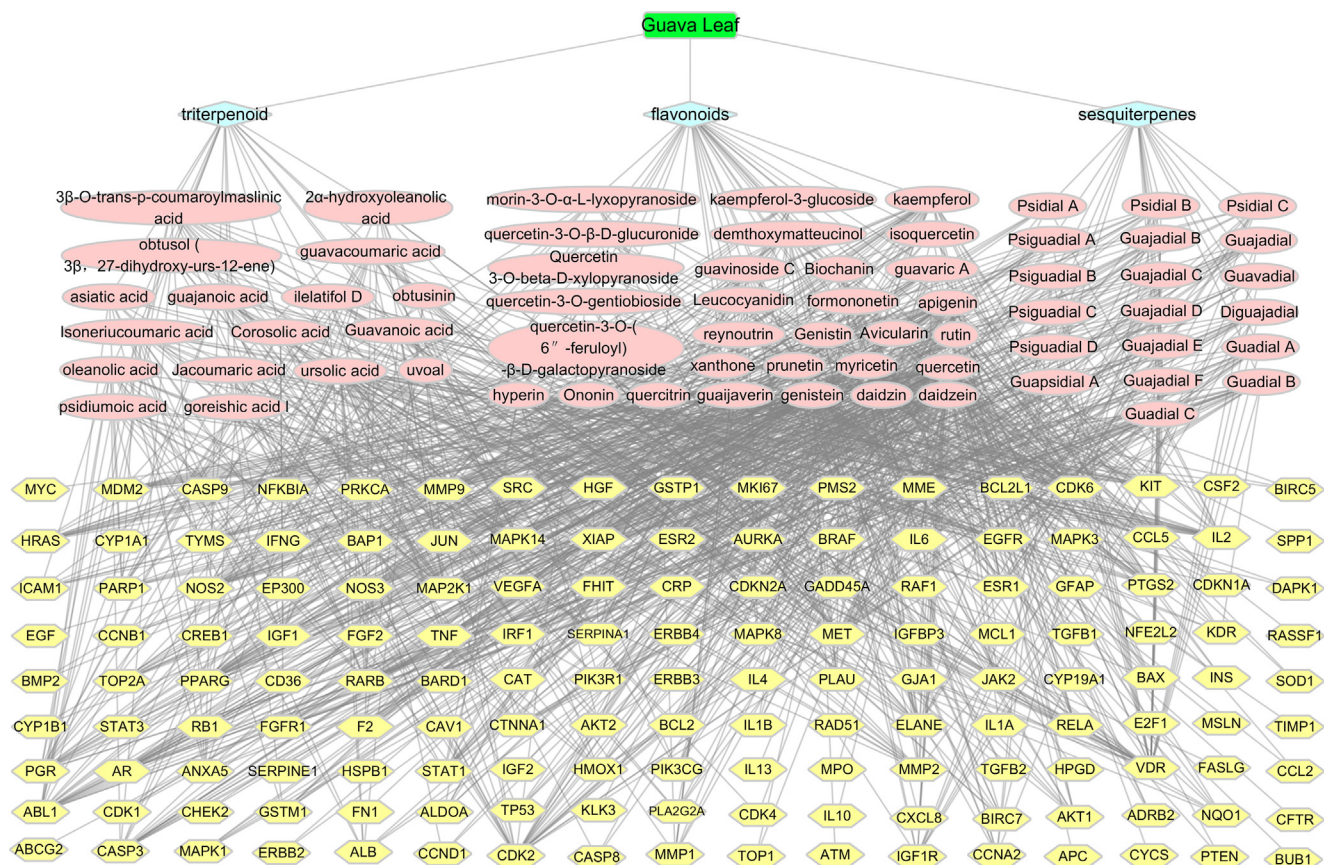
**Fig. 3** Interactive network of ‘guava leaf - compound class – active compound – gene’.

Table 5 Target and genetic information of compounds with interaction network Degree ≥ 20 .

| Compound | Targets | Genes |
|----------|---|----------|
| apigenin | RAC-alpha serine/threonine-protein kinase | AKT1 |
| apigenin | Adenomatous polyposis coli protein | APC |
| apigenin | Androgen receptor | AR |
| apigenin | Apoptosis regulator BAX | BAX |
| apigenin | Apoptosis regulator Bcl-2 | BCL2 |
| apigenin | Bcl-2-like protein 1 | BCL2L1 |
| apigenin | Caspase-3 | CASP3 |
| apigenin | Caspase-9 | CASP9 |
| apigenin | G2/mitotic-specific cyclin-B1 | CCNB1 |
| apigenin | G1/S-specific cyclin-D1 | CCND1 |
| apigenin | Cell division control protein 2 homolog | CDK1 |
| apigenin | Cell division protein kinase 2 | CDK2 |
| apigenin | Cell division protein kinase 4 | CDK4 |
| apigenin | Cell division protein kinase 6 | CDK6 |
| apigenin | Cyclin-dependent kinase inhibitor 1 | CDKN1A |
| apigenin | Cyclin-dependent kinase inhibitor 2 | CDKN2A |
| apigenin | Cytochrome <i>c</i> | CYCS |
| apigenin | Cytochrome P450 19A1 | CYP19A1 |
| apigenin | Estrogen receptor | ESR1 |
| apigenin | Estrogen receptor beta | ESR2 |
| apigenin | Prothrombin | F2 |
| apigenin | Basic fibroblast growth factor receptor 1 | FGFR1 |
| apigenin | Heme oxygenase 1 | HMOX1 |
| apigenin | Intercellular adhesion molecule 1 | ICAM1 |
| apigenin | Interferon gamma | IFNG |
| apigenin | Insulin-like growth factor 1 receptor | IGF1R |
| apigenin | Interleukin-13 | IL13 |
| apigenin | Interleukin-2 | IL2 |
| apigenin | Interleukin-4 | IL4 |
| apigenin | Insulin | INS |
| apigenin | Transcription factor AP-1 | JUN |
| apigenin | Vascular endothelial growth factor receptor 2 | KDR |
| apigenin | Induced myeloid leukemia cell differentiation protein Mcl-1 | MCL1 |
| apigenin | E3 ubiquitin-protein ligase Mdm2 | MDM2 |
| apigenin | Interstitial collagenase | MMP1 |
| apigenin | Matrix metalloproteinase-9 | MMP9 |
| apigenin | NF-kappa-B inhibitor alpha | NFKBIA |
| apigenin | Nitric oxide synthase, endothelial | NOS3 |
| apigenin | Phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit, gamma isoform | PIK3CG |
| apigenin | Phospholipase A2, membrane associated | PLA2G2A |
| apigenin | Urokinase-type plasminogen activator | PLAU |
| apigenin | Prostaglandin G/H synthase 2 | PTGS2 |
| apigenin | Retinoblastoma-associated protein | RB1 |
| apigenin | Transcription factor p65 | RELA |
| apigenin | Alpha-1-antitrypsin | SERPINE1 |
| apigenin | Proto-oncogene tyrosine-protein kinase Src | SRC |
| apigenin | Tumor necrosis factor | TNF |
| apigenin | DNA topoisomerase II | TOP2A |
| apigenin | Cellular tumor antigen p53 | TP53 |
| apigenin | Vitamin D3 receptor | VDR |
| apigenin | Vascular endothelial growth factor A | VEGFA |
| apigenin | Baculoviral IAP repeat-containing protein 4 | XIAP |
| daidzein | Beta-2 adrenergic receptor | ADRB2 |
| daidzein | Ubiquitin carboxyl-terminal hydrolase BAP1 | BAP1 |
| daidzein | BRCA1-associated RING domain protein 1 | BARD1 |
| daidzein | Apoptosis regulator BAX | BAX |
| daidzein | Caspase-3 | CASP3 |
| daidzein | Catalase | CAT |
| daidzein | Caveolin-1 | CAV1 |
| daidzein | Cell division protein kinase 2 | CDK2 |
| daidzein | Cell division protein kinase 6 | CDK6 |
| daidzein | Cyclin-dependent kinase inhibitor 1 | CDKN1A |
| daidzein | Histone acetyltransferase p300 | EP300 |

(continued on next page)

Table 5 (continued)

| Compound | Targets | Genes |
|---------------------|---|---------|
| daidzein | Estrogen receptor | ESR1 |
| daidzein | Estrogen receptor beta | ESR2 |
| daidzein | Prothrombin | F2 |
| daidzein | Growth arrest and DNA damage-inducible protein GADD45 alpha | GADD45A |
| daidzein | Intercellular adhesion molecule 1 | ICAM1 |
| daidzein | Insulin-like growth factor 1 receptor | IGF1R |
| daidzein | Interleukin-4 | IL4 |
| daidzein | Interleukin-6 | IL6 |
| daidzein | Transcription factor AP-1 | JUN |
| daidzein | Mitogen-activated protein kinase 14 | MAPK14 |
| daidzein | Mitogen-activated protein kinase 8 | MAPK8 |
| daidzein | Antigen KI-67 | MKI67 |
| daidzein | Neprilysin | MME |
| daidzein | Nitric oxide synthase, inducible | NOS2 |
| daidzein | Nitric oxide synthase, endothelial | NOS3 |
| daidzein | Progesterone receptor | PGR |
| daidzein | Phosphatidylinositol 3-kinase regulatory subunit alpha | PIK3R1 |
| daidzein | Peroxisome proliferator activated receptor gamma | PPARG |
| daidzein | Peroxisome proliferator activated receptor gamma | PPARG |
| daidzein | Prostaglandin G/H synthase 2 | PTGS2 |
| daidzein | DNA repair protein RAD51 homolog 1 | RAD51 |
| daidzein | Transcription factor p65 | RELA |
| daidzein | Signal transducer and activator of transcription 1-alpha/beta | STAT1 |
| daidzein | Tumor necrosis factor | TNF |
| daidzein | Cellular tumor antigen p53 | TP53 |
| daidzein | Vascular endothelial growth factor A | VEGFA |
| demthoxymatteucinol | Proto-oncogene tyrosine-protein kinase ABL1 | ABL1 |
| demthoxymatteucinol | Serine/threonine-protein kinase 6 | AURKA |
| demthoxymatteucinol | Bone morphogenetic protein 2 | BMP2 |
| demthoxymatteucinol | Cell division protein kinase 6 | CDK6 |
| demthoxymatteucinol | Catenin alpha-1 | CTNNA1 |
| demthoxymatteucinol | Leukocyte elastase | ELANE |
| demthoxymatteucinol | Basic fibroblast growth factor receptor 1 | FGFR1 |
| demthoxymatteucinol | Insulin-like growth factor 1 receptor | IGF1R |
| demthoxymatteucinol | Tyrosine-protein kinase JAK2 | JAK2 |
| demthoxymatteucinol | Vascular endothelial growth factor receptor 2 | KDR |
| demthoxymatteucinol | Mast/stem cell growth factor receptor | KIT |
| demthoxymatteucinol | Dual specificity mitogen-activated protein kinase kinase 1 | MAP2K1 |
| demthoxymatteucinol | Nitric oxide synthase, endothelial | NOS3 |
| demthoxymatteucinol | NAD(P)H dehydrogenase [quinone] 1 | NQO1 |
| demthoxymatteucinol | Progesterone receptor | PGR |
| demthoxymatteucinol | Phospholipase A2, membrane associated | PLA2G2A |
| demthoxymatteucinol | Urokinase-type plasminogen activator | PLAU |
| demthoxymatteucinol | Peroxisome proliferator activated receptor gamma | PPARG |
| demthoxymatteucinol | Thymidylate synthase | TYMS |
| demthoxymatteucinol | Vitamin D3 receptor | VDR |
| formononetin | Beta-2 adrenergic receptor | ADRB2 |
| formononetin | Androgen receptor | AR |
| formononetin | C-C motif chemokine 5 | CCL5 |
| formononetin | Cyclin-A2 | CCNA2 |
| formononetin | Cell division protein kinase 2 | CDK2 |
| formononetin | Cell division protein kinase 6 | CDK6 |
| formononetin | Estrogen receptor | ESR1 |
| formononetin | Estrogen receptor beta | ESR2 |
| formononetin | Prothrombin | F2 |
| formononetin | Interleukin-4 | IL4 |
| formononetin | Transcription factor AP-1 | JUN |
| formononetin | Mitogen-activated protein kinase 14 | MAPK14 |
| formononetin | Mitogen-activated protein kinase 8 | MAPK8 |
| formononetin | Neprilysin | MME |
| formononetin | Nitric oxide synthase, inducible | NOS2 |
| formononetin | Nitric oxide synthase, endothelial | NOS3 |
| formononetin | Progesterone receptor | PGR |

Table 5 (continued)

| Compound | Targets | Genes |
|--------------|--|--------|
| formononetin | Peroxisome proliferator activated receptor gamma | PPARG |
| formononetin | Peroxisome proliferator activated receptor gamma | PPARG |
| formononetin | Prostaglandin G/H synthase 2 | PTGS2 |
| genistein | RAC-alpha serine/threonine-protein kinase | AKT1 |
| genistein | Androgen receptor | AR |
| genistein | Serine-protein kinase ATM | ATM |
| genistein | Apoptosis regulator BAX | BAX |
| genistein | Apoptosis regulator Bcl-2 | BCL2 |
| genistein | Baculoviral IAP repeat-containing protein 5 | BIRC5 |
| genistein | Baculoviral IAP repeat-containing protein 7 | BIRC7 |
| genistein | Mitotic checkpoint serine/threonine-protein kinase BUB1 | BUB1 |
| genistein | Caspase-3 | CASP3 |
| genistein | Caspase-9 | CASP9 |
| genistein | C-C motif chemokine 2 | CCL2 |
| genistein | Cyclin-A2 | CCNA2 |
| genistein | G2/mitotic-specific cyclin-B1 | CCNB1 |
| genistein | Cell division control protein 2 homolog | CDK1 |
| genistein | Cell division protein kinase 2 | CDK2 |
| genistein | Cyclin-dependent kinase inhibitor 1 | CDKN1A |
| genistein | Cyclin-dependent kinase inhibitor 2 | CDKN2A |
| genistein | Cystic fibrosis transmembrane conductance regulator | CFTR |
| genistein | Serine/threonine-protein kinase Chk2 | CHEK2 |
| genistein | Interleukin-8 | CXCL8 |
| genistein | Epidermal growth factor receptor | EGFR |
| genistein | Leukocyte elastase | ELANE |
| genistein | Receptor tyrosine-protein kinase erbB-2 | ERBB2 |
| genistein | Estrogen receptor | ESR1 |
| genistein | Estrogen receptor beta | ESR2 |
| genistein | Prothrombin | F2 |
| genistein | Basic fibroblast growth factor receptor 1 | FGFR1 |
| genistein | Fibronectin | FN1 |
| genistein | Glial fibrillary acidic protein | GFAP |
| genistein | 15-hydroxyprostaglandin dehydrogenase [NAD +] | HPGD |
| genistein | Intercellular adhesion molecule 1 | ICAM1 |
| genistein | Insulin-like growth factor 1 receptor | IGF1R |
| genistein | Interleukin-1 beta | IL1B |
| genistein | Insulin | INS |
| genistein | Transcription factor AP-1 | JUN |
| genistein | Prostate-specific antigen | KLK3 |
| genistein | Mitogen-activated protein kinase 1 | MAPK1 |
| genistein | Mitogen-activated protein kinase 14 | MAPK14 |
| genistein | Mitogen-activated protein kinase 3 | MAPK3 |
| genistein | E3 ubiquitin-protein ligase Mdm2 | MDM2 |
| genistein | Nephrilysin | MME |
| genistein | Matrix metalloproteinase-9 | MMP9 |
| genistein | Mesothelin | MSLN |
| genistein | Nitric oxide synthase, inducible | NOS2 |
| genistein | Nitric oxide synthase, endothelial | NOS3 |
| genistein | Progesterone receptor | PGR |
| genistein | Phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit, gamma isoform | PIK3CG |
| genistein | Urokinase-type plasminogen activator | PLAU |
| genistein | Peroxisome proliferator activated receptor gamma | PPARG |
| genistein | Peroxisome proliferator activated receptor gamma | PPARG |
| genistein | Phosphatidylinositol-3,4,5-trisphosphate 3-phosphatase and dual-specificity protein phosphatase PTEN | PTEN |
| genistein | Prostaglandin G/H synthase 2 | PTGS2 |
| genistein | Transcription factor p65 | RELA |
| genistein | Signal transducer and activator of transcription 1-alpha/beta | STAT1 |
| genistein | Signal transducer and activator of transcription 3 | STAT3 |
| genistein | Transforming growth factor beta-1 | TGFB1 |
| genistein | Transforming growth factor beta-2 | TGFB2 |
| genistein | Metalloproteinase inhibitor 1 | TIMP1 |
| genistein | Tumor necrosis factor | TNF |
| genistein | Cellular tumor antigen p53 | TP53 |
| genistein | Vascular endothelial growth factor A | VEGFA |

The top 10 enriched biological processes are shown in Fig. 4A. The enrichment results included apoptosis, programmed cell death, regulation of programmed cell death, regulation of cell death, regulation of apoptotic process,

regulation of cell population proliferation, response to oxygen-containing compound, negative regulation of programmed cell death, and negative regulation of cell death. The top 10 enriched cellular components are shown in

Table 6 GO enrichment analysis of the “guava leaf-lung cancer” interaction gene.

| GO ID | GO Term | Percentage of genes | P-Value | Corr P-Value |
|------------|--|---------------------|----------|--------------|
| GO:0006915 | apoptotic process(BP) | 112/2458 (4.56%) | 9.13E-64 | 1.18E-60 |
| GO:0012501 | programmed cell death (BP) | 114/2605(4.38%) | 1.59E-63 | 2.05E-60 |
| GO:0043067 | regulation of programmed cell death (BP) | 103/1991(5.17%) | 4.85E-62 | 6.24E-59 |
| GO:0010941 | regulation of cell death(BP) | 105/2153(4.88%) | 4.24E-61 | 5.45E-58 |
| GO:0042981 | regulation of apoptotic process(BP) | 101/1971(5.12%) | 5.21E-60 | 6.69E-57 |
| GO:0042127 | regulation of cell population proliferation(BP) | 100/2092(4.78%) | 2.59E-56 | 3.32E-53 |
| GO:1901700 | response to oxygen-containing compound(BP) | 98/2088(4.69%) | 4.63E-54 | 5.94E-51 |
| GO:0051247 | positive regulation of protein metabolic process(BP) | 97/2115(4.59%) | 2.12E-52 | 2.72E-49 |
| GO:0043069 | negative regulation of programmed cell death(BP) | 78/1182(6.60%) | 2.21E-51 | 2.83E-48 |
| GO:0060548 | negative regulation of cell death(BP) | 78/1299(6.16%) | 1.23E-50 | 1.58E-47 |
| GO:0031093 | platelet alpha granule lumen(CC) | 15/88(17.05%) | 3.46E-16 | 1.14E-14 |
| GO:0031091 | platelet alpha granule(CC) | 16/118(13.56%) | 1.51E-15 | 4.83E-14 |
| GO:0031983 | vesicle lumen(CC) | 24/470(5.11%) | 4.76E-13 | 1.48E-11 |
| GO:0060205 | cytoplasmic vesicle lumen(CC) | 23/469(4.90%) | 3.51E-12 | 1.05E-10 |
| GO:0034774 | secretory granule lumen(CC) | 22/448(4.91%) | 1.05E-11 | 3.06E-10 |
| GO:0000307 | cyclin-dependent protein kinase holoenzyme complex(CC) | 10/57(17.54%) | 2.54E-11 | 7.12E-10 |
| GO:0045121 | membrane raft(CC) | 21/441(4.76%) | 5.74E-11 | 1.55E-09 |
| GO:0098857 | membrane microdomain(CC) | 21/442(4.75%) | 5.98E-11 | 1.56E-09 |
| GO:0098589 | membrane region(CC) | 21/457(4.60%) | 1.11E-10 | 2.77E-09 |
| GO:1902911 | protein kinase complex(CC) | 11/137(8.03%) | 1.36E-08 | 3.26E-07 |
| GO:0004672 | protein kinase activity(MF) | 72/1534(4.69%) | 1.14E-36 | 2.51E-34 |
| GO:0043085 | positive regulation of catalytic activity(MF) | 76/1809(4.20%) | 9.45E-36 | 2.07E-33 |
| GO:0016773 | phosphotransferase activity, alcohol group as acceptor(MF) | 73/1661(4.39%) | 2.32E-35 | 5.06E-33 |
| GO:0016301 | kinase activity(MF) | 75/1798(4.17%) | 5.68E-35 | 1.23E-32 |
| GO:0004674 | protein serine/threonine kinase activity(MF) | 60/1094(5.48%) | 1.79E-33 | 3.87E-31 |
| GO:0051338 | regulation of transferase activity(MF) | 64/1298(4.93%) | 3.03E-33 | 6.51E-31 |
| GO:0043549 | regulation of kinase activity(MF) | 61/1153(5.29%) | 3.28E-33 | 7.01E-31 |
| GO:0071900 | regulation of protein serine/threonine kinase activity(MF) | 50/708(7.06%) | 1.36E-32 | 2.89E-30 |
| GO:0045859 | regulation of protein kinase activity(MF) | 58/1067(5.44%) | 4.95E-32 | 1.05E-29 |
| GO:0033674 | positive regulation of kinase activity(MF) | 50/7755(6.45%) | 9.88E-31 | 2.08E-28 |

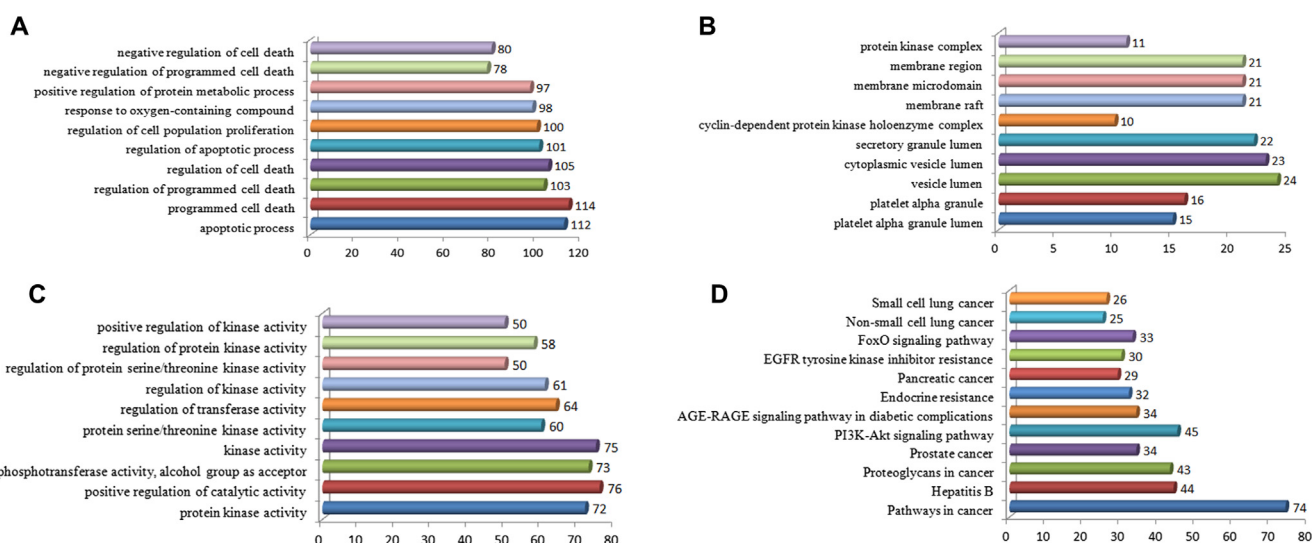


Fig. 4 GO and KEGG analysis of ‘guava leaf – lung cancer’ interactive gene with P-values from small to large. (A) GO biological process analysis of ‘guava leaf – lung cancer’ interactive gene with P-values from small to large. (B) GO cell components analysis of ‘guava leaf – lung cancer’ interactive gene with P-values from small to large. (C) GO molecular function analysis of ‘guava leaf – lung cancer’ interactive gene with P-values from small to large. (D) KEGG analysis of ‘guava leaf – lung cancer’ cross-gene.

Fig. 4B. The enrichment results included platelet alpha granule lumen, platelet alpha granules, vesicle lumen, cytoplasmic vesicle lumen, secretory granule lumen, cyclin-dependent protein kinase holoenzyme complex, membrane raft, membrane microdomain, membrane region, and protein kinase complex. The top 10 enriched molecular function terms are shown in **Fig. 4C.** The enrichment results included protein kinase activity, positive regulation of catalytic activity, phosphotransferase activity, alcohol group as acceptor, phosphokinase activity,

kinase activity, protein serine/threonine kinase activity, regulation of transferase activity, regulation of kinase activity, regulation of protein serine/threonine kinase activity, regulation of protein kinase activity, and positive regulation of kinase activity.

KEGG enrichment analysis results of common genes between guava leaf constituent target and lung cancer related genes are shown in **Fig. 4D**, **Table 7** and **Table 8**. The top 10 common enriched pathways are listed below: cancer path-

Table 7 Results of KEGG enrichment analysis of “guava leaf-lung cancer” interactive gene.

| KEGG ID | GO Term | Percentage of genes | P-Value | Corr P-Value |
|----------|--|---------------------|-----------|--------------|
| hsa05200 | Pathways in cancer | 74/397(18.64%) | 1.63E-101 | 3.53E-99 |
| hsa05161 | Hepatitis B | 44/146(30.14%) | 1.06E-66 | 1.15E-64 |
| hsa05205 | Proteoglycans in cancer | 43/205(20.98%) | 3.33E-59 | 2.41E-57 |
| hsa05215 | Prostate cancer | 34/89(38.20%) | 2.56E-54 | 1.39E-52 |
| hsa04151 | PI3K-Akt signaling pathway | 45/342(13.16%) | 9.08E-54 | 3.94E-52 |
| hsa04933 | AGE-RAGE signaling pathway in diabetic complications | 34/101(33.66%) | 9.60E-53 | 3.47E-51 |
| hsa01522 | Endocrine resistance | 32/97 (32.99%) | 2.15E-49 | 6.68E-48 |
| hsa05212 | Pancreatic cancer | 29/66(43.94%) | 8.75E-48 | 2.37E-46 |
| hsa01521 | EGFR tyrosine kinase inhibitor resistance | 30/81(37.04%) | 1.39E-47 | 3.35E-46 |
| hsa04068 | FoxO signaling pathway | 33/134(24.63%) | 2.60E-47 | 5.64E-46 |
| hsa05223 | Non-small cell lung cancer | 25/56(44.64%) | 2.17E-41 | 3.63E-40 |
| hsa05222 | Small cell lung cancer | 26/86(30.23%) | 2.45E-39 | 2.95E-38 |

Table 8 The enrichment gene list of KEGG of “guava leaf - lung cancer” interaction gene.

| KEGG ID | Enriched genes in the KEGG pathway |
|----------|--|
| hsa05200 | CCND1,BCL2,BCL2L1,PIK3CG,TGFB2,XIAP,KIT,BIRC5,MAP2K1,PTGS2,NOS2,JUN,RAD51,ABL1,PTEN,CASP3,TP53,GSTP1,CXCL8,DAPK1,IGF1,RB1,EP300,CDK4,BMP2,AKT2,CDK6,FGFR1,FGF2,TGFB1,HGF,MMP1,MMP2,RASSF1,APC,AKT1,BAX,MMP9,IGF1R,CYCS,BIRC7,BRAF,MDM2,EGFR,RAF1,EGF,MYC,E2F1,PPARG,MET,STAT3,NFKBIA,FN1,CDKN1A,MAPK3,MAPK1,CASP8,PRKCA,MAPK8,ERBB2,IL6,CDKN2A,STAT1,FASLG,KLK3,CDK2,VEGFA,AR,RARB,HRAS,RELA,PIK3R1,CASP9,CTNNA1 |
| hsa05161 | CCND1,BCL2,PIK3CG,TGFB2,BIRC5,MAP2K1,JUN,TNF,PTEN,CASP3,TP53,CXCL8,RB1,EP300,CDK4,CDK2,CCNA2,PIK3R1,AKT1,BAX,MMP9,AKT2,CYCS,RAF1,TGFB1,MYC,E2F1,CDK6,SRC,STAT3,NFKBIA,CASP9,CDKN1A,MAPK3,MAPK1,CASP8,PRKCA,MAPK8,IL6,STAT1,FASLG,HRAS,RELA,CREB1 |
| hsa05205 | CCND1,PIK3CG,MYC,MAP2K1,ESR1,TNF,CAV1,CASP3,TP53,KDR,IGF1,MMP9,AKT2,FGFR1,FGF2,HGF,MMP2,AKT1,IGF2,IGF1R,MDM2,BRAF,EGFR,RAF1,TGFB1,TGFB2,MAPK14,SRC,MET,PLAU,FN1,CDKN1A,MAPK3,MAPK1,PRKCA,STAT3,ERBB2,FASLG,VEGFA,HRAS,PIK3R1,ERBB3,ERBB4 |
| hsa05215 | CDK2,CDKN1A,CREB1,E2F1,EGF,EGFR,EP300,ERBB2,AKT1,AKT2,FGFR1,GSTP1,HRAS,IGF1,IGF1R,KLK3,INS,AR,MDM2,NFKBIA,PIK3CG,PIK3R1,MAPK1,MAPK3,MAP2K1,PTEN,RAF1,RB1,CCND1,BCL2,RELA,BRAF,TP53,CASP9 |
| hsa04151 | CDK2,CDK4,CDK6,CDKN1A,CREB1,EGF,EGFR,AKT1,AKT2,FGF2,FGFR1,FN1,HGF,HRAS,IGF1,IGF1R,IL2,FASLG,IL4,IL6,INS,JAK2,KDR,KIT,MCL1,MDM2,MET,MYC,NOS3,PIK3CG,PIK3R1,PRKCA,MAPK1,MAPK3,MAP2K1,PTEN,RAF1,CCND1,BCL2,RELA,BCL2L1,SPP1,TP53,VEGFA,CASP9 |
| hsa04933 | CDK4,MAPK14,AKT1,AKT2,FN1,HRAS,ICAM1,IL1A,IL1B,IL6,CXCL8,JAK2,JUN,MMP2,NOS3,SERPINE1,PIK3CG,PIK3R1,PRKCA,MAPK1,MAPK3,MAPK8,BAX,CCND1,BCL2,RELA,CCL2,STAT1,STAT3,TGFB1,TGFB2,TNF,VEGFA,CASP3 |
| hsa01522 | CDK4,CDKN1A,CDKN2A,MAPK14,E2F1,EGFR,ERBB2,AKT1,AKT2,ESR1,ESR2,HRAS,IGF1,IGF1R,JUN,MDM2,MMP2,MMP9,PIK3CG,PIK3R1,MAPK1,MAPK3,MAPK8,MAP2K1,BAX,RAF1,RB1,CCND1,BCL2,SRC,BRAF,TP53 |
| hsa05212 | CDK4,CDK6,CDKN2A,E2F1,EGF,EGFR,ERBB2,AKT1,AKT2,PIK3CG,PIK3R1,MAPK1,MAPK3,MAPK8,MAP2K1,RAD51,RAF1,RB1,CCND1,RELA,BCL2L1,BRAF,STAT1,STAT3,TGFB1,TGFB2,TP53,VEGFA,CASP9 |
| hsa01521 | EGF,EGFR,ERBB2,ERBB3,AKT1,AKT2,FGF2,HGF,HRAS,IGF1,IGF1R,IL6,JAK2,KDR,MET,PIK3CG,PIK3R1,PRKCA,MAPK1,MAPK3,MAP2K1,PTEN,BAX,RAF1,BCL2,BCL2L1,SRC,BRAF,STAT3,VEGFA |
| hsa04068 | CDK2,CDKN1A,MAPK14,GADD45A,EGF,EGFR,EP300,AKT1,AKT2,HRAS,IGF1,IGF1R,FASLG,IL6,IL10,INS,MDM2,ATM,PIK3CG,PIK3R1,MAPK1,MAPK3,MAPK8,MAP2K1,PTEN,RAF1,CCND1,BRAF,STAT3,TGFB1,TGFB2,CAT,CCNB1 |
| hsa05223 | CDK4,CDK6,CDKN2A,RASSF1,E2F1,EGF,EGFR,ERBB2,AKT1,AKT2,FHIT,HRAS,PIK3CG,PIK3R1,PRKCA,MAPK1,MAPK3,MAP2K1,RAF1,RARB,RB1,CCND1,BRAF,TP53,CASP9 |
| hsa05222 | CDK2,CDK4,CDK6,E2F1,AKT1,AKT2,FHIT,FN1,XIAP,MYC,NFKBIA,NOS2,PIK3CG,PIK3R1,CYCS,PTEN,PTGS2,RARB,RB1,CCND1,BCL2,RELA,BCL2L1,TP53,BIRC7,CASP9 |

way, hepatitis B, proteoglycans in cancer, prostate cancer, PI3K-Akt signaling pathway, AGE-RAGE signaling pathway in diabetic complications, endocrine resistance, pancreatic cancer, EGFR tyrosine kinase inhibitor resistance, and FoxO signaling pathway. The non-small cell lung cancer and small cell lung cancer pathways were also recorded.

One of these pathways is the PI3K-Akt signaling pathway, which has been shown to play an important regulatory role in tumor therapy. The PI3K-Akt signaling pathway is an important pathway for cell survival, metabolism, angiogenesis, apoptosis, proliferation and differentiation. The substrate is used to control key cellular processes, and many targets are involved in this pathway and AKT also plays an important role in the regulation of this pathway (Ebrahimi et al., 2017). In order to investigate the effectiveness of guava leaf constituents on lung

cancer via the PI3K-Akt signaling pathway, we analyzed the expression (Fig. 5A) and the pathway activities (Fig. 5) of these genes in lung squamous cell carcinoma (LUSC) and lung adenocarcinoma (LUAD) samples from TCGA database in the GSCAlite online platform. The results indicated that the PI3K-Akt signaling pathway genes that could be potentially affected by the guava leaf constituents showed differential expression with some of them being downregulated (Fig. 5A, blue color dot) or upregulated (Fig. 5A, red color dot) in both LUSC and LUAD. In addition, we found that some of the target genes of the guava leaf constituents did not show differential expression in lung cancer (Fig. 5A). Furthermore, in pathway activity analysis, we found that these target genes were involved in the inhibition or activation of important pathways such as PI3K-Akt, apoptosis, cell cycle, EMT,

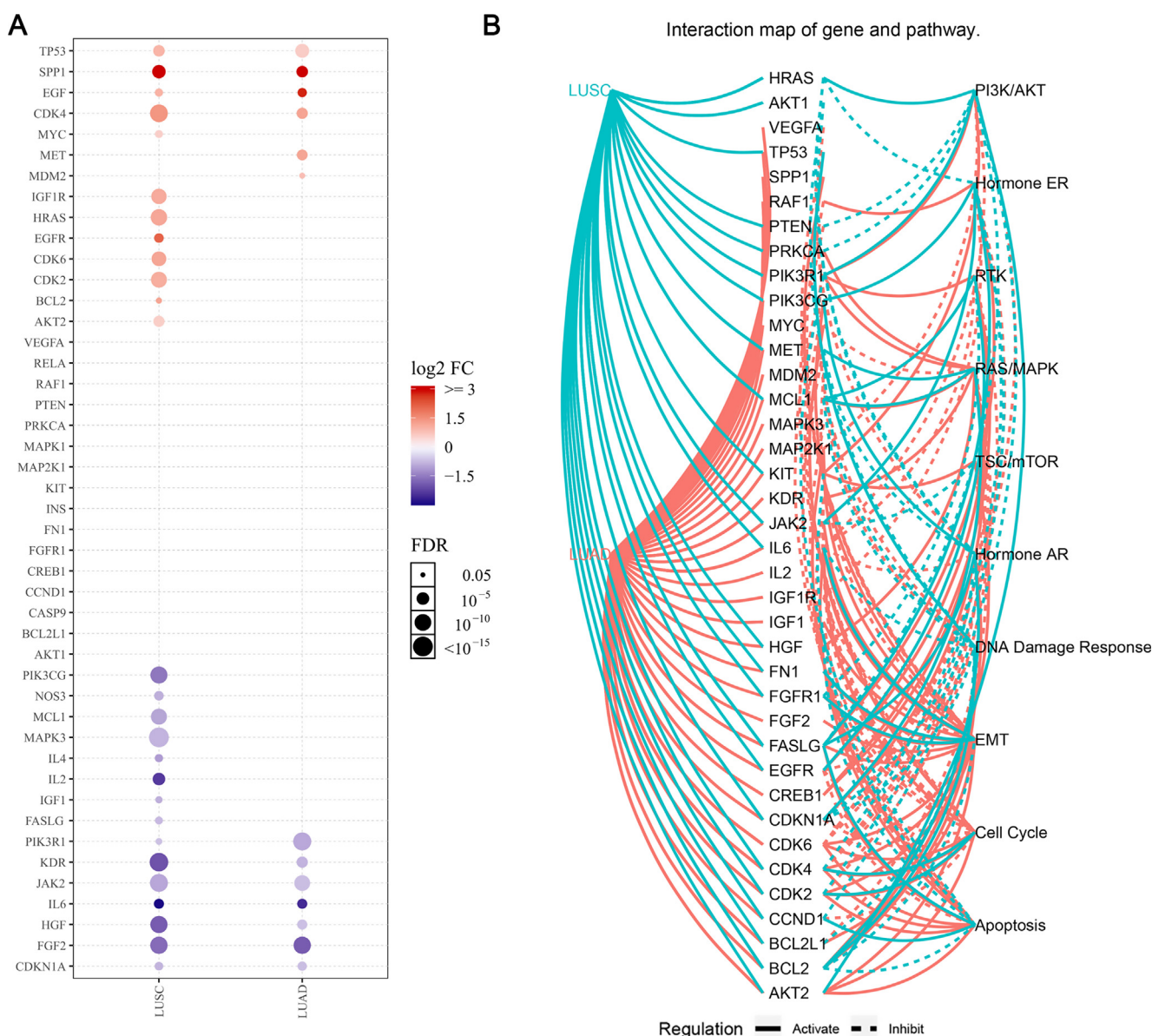


Fig. 5 The expression and pathways activity analysis of guava leaf constituent target genes in the PI3K-Akt signaling pathway. (A) The mRNA expression profile of genes. The dots represent the fold change; the blue color indicates downregulation while the red color indicates upregulation. The size of the dot is proportional to the expression foldchange (FC). (B) Pathway activity analysis.

DNA damage response, hormone AR, TSC/mTOR, RAS/MAPK, RTK and hormone ER pathways.

3.6. Molecular docking

The previous drug-disease interaction gene network topological analysis showed that the 10 genes with the strongest interaction were STAT3, TP53, AKT1, JUN, IL6, VEGFA, SRC, TNF, HRAS, and CDKN1A, which are effective in the “Guava leaf-constituent category” network. The Cytoscape

interaction network of component-genes showed that the five compounds with strong interaction between guava leaf constituents and lung cancer genes were quercetin, genistein, apigenin, ursolic acid, and daidzein. The five compounds and 10 target proteins were molecularly docked by online molecular docking on the systemsDock website. The score of the database system docking was between 0 and 10. The larger the docking score, the better the docking effect, and the greater the binding activity between the docking molecule and the target. A docking score greater than 4.25 indicates that a ligand

Table 9 Docking results of receptor proteins and ligand compounds.

| Ligand ID | compounds | Receptor proteins | PDB ID | gene | Docking results |
|-----------|--------------|--|--------|--------|-----------------|
| 5,280,343 | quercetin | Signal transducer and activator of transcription 3 | 1BG1 | STAT3 | 7.475 |
| 5,280,863 | kaempferol | Signal transducer and activator of transcription 3 | 1BG1 | STAT3 | 7.437 |
| 5,280,961 | Genistein | Signal transducer and activator of transcription 3 | 1BG1 | STAT3 | 7.809 |
| 5,281,708 | Daidzein | Signal transducer and activator of transcription 3 | 1BG1 | STAT3 | 7.811 |
| 64,945 | ursolic acid | Signal transducer and activator of transcription 3 | 1BG1 | STAT3 | 4.88 |
| 5,280,343 | quercetin | Cellular tumor antigen p53 | 5MGT | TP53 | 6.658 |
| 5,280,863 | kaempferol | Cellular tumor antigen p53 | 5MGT | TP53 | 6.689 |
| 5,280,961 | Genistein | Cellular tumor antigen p53 | 5MGT | TP53 | 6.669 |
| 5,281,708 | Daidzein | Cellular tumor antigen p53 | 5MGT | TP53 | 6.662 |
| 64,945 | ursolic acid | Cellular tumor antigen p53 | 5MGT | TP53 | 5.464 |
| 5,280,343 | quercetin | RAC-alpha serine/threonine-protein kinase | 3CQW | AKT1 | 6.847 |
| 5,280,863 | kaempferol | RAC-alpha serine/threonine-protein kinase | 3CQW | AKT1 | 6.858 |
| 5,280,961 | Genistein | RAC-alpha serine/threonine-protein kinase | 3CQW | AKT1 | 6.824 |
| 5,281,708 | Daidzein | RAC-alpha serine/threonine-protein kinase | 3CQW | AKT1 | 6.85 |
| 64,945 | ursolic acid | RAC-alpha serine/threonine-protein kinase | 3CQW | AKT1 | 8.331 |
| 5,280,343 | quercetin | Interleukin-6 | 1ALU | IL6 | 6.684 |
| 5,280,863 | kaempferol | Interleukin-6 | 1ALU | IL6 | 6.656 |
| 5,280,961 | Genistein | Interleukin-6 | 1ALU | IL6 | 6.632 |
| 5,281,708 | Daidzein | Interleukin-6 | 1ALU | IL6 | 6.663 |
| 64,945 | ursolic acid | Interleukin-6 | 1ALU | IL6 | 4.936 |
| 5,280,343 | quercetin | Transcription factor AP-1 | 5TO1 | JUN | 6.569 |
| 5,280,863 | kaempferol | Transcription factor AP-1 | 5TO1 | JUN | 6.548 |
| 5,280,961 | Genistein | Transcription factor AP-1 | 5TO1 | JUN | 6.59 |
| 5,281,708 | Daidzein | Transcription factor AP-1 | 5TO1 | JUN | 6.637 |
| 64,945 | ursolic acid | Transcription factor AP-1 | 5TO1 | JUN | 4.892 |
| 5,280,343 | quercetin | Vascular endothelial growth factor A | 6BFT | VEGFA | 5.849 |
| 5,280,863 | kaempferol | Vascular endothelial growth factor A | 6BFT | VEGFA | 5.937 |
| 5,280,961 | Genistein | Vascular endothelial growth factor A | 6BFT | VEGFA | 6.601 |
| 5,281,708 | Daidzein | Vascular endothelial growth factor A | 6BFT | VEGFA | 6.65 |
| 64,945 | ursolic acid | Vascular endothelial growth factor A | 6BFT | VEGFA | 5.799 |
| 5,280,343 | quercetin | Proto-oncogene tyrosine-protein kinase Src | 2BDJ | SRC | 6.46 |
| 5,280,863 | kaempferol | Proto-oncogene tyrosine-protein kinase Src | 2BDJ | SRC | 6.785 |
| 5,280,961 | Genistein | Proto-oncogene tyrosine-protein kinase Src | 2BDJ | SRC | 6.246 |
| 5,281,708 | Daidzein | Proto-oncogene tyrosine-protein kinase Src | 2BDJ | SRC | 6.789 |
| 64,945 | ursolic acid | Proto-oncogene tyrosine-protein kinase Src | 2BDJ | SRC | 7.927 |
| 5,280,343 | quercetin | Tumor necrosis factor | 3ALQ | TNF | 4.904 |
| 5,280,863 | kaempferol | Tumor necrosis factor | 3ALQ | TNF | 5.01 |
| 5,280,961 | Genistein | Tumor necrosis factor | 3ALQ | TNF | 5.109 |
| 5,281,708 | Daidzein | Tumor necrosis factor | 3ALQ | TNF | 5.063 |
| 64,945 | ursolic acid | Tumor necrosis factor | 3ALQ | TNF | 5.945 |
| 5,280,343 | quercetin | GTPase HRas | 6D5W | HRAS | 6.434 |
| 5,280,863 | kaempferol | GTPase HRas | 6D5W | HRAS | 6.582 |
| 5,280,961 | Genistein | GTPase HRas | 6D5W | HRAS | 6.595 |
| 5,281,708 | Daidzein | GTPase HRas | 6D5W | HRAS | 6.605 |
| 64,945 | ursolic acid | GTPase HRas | 6D5W | HRAS | 7.867 |
| 5,280,343 | quercetin | Cyclin-dependent kinase inhibitor 1 | 3TS8 | CDKN1A | 5.02 |
| 5,280,863 | kaempferol | Cyclin-dependent kinase inhibitor 1 | 3TS8 | CDKN1A | 5.023 |
| 5,280,961 | Genistein | Cyclin-dependent kinase inhibitor 1 | 3TS8 | CDKN1A | 4.663 |
| 5,281,708 | Daidzein | Cyclin-dependent kinase inhibitor 1 | 3TS8 | CDKN1A | 4.94 |
| 64,945 | ursolic acid | Cyclin-dependent kinase inhibitor 1 | 3TS8 | CDKN1A | 8.148 |

has a certain binding activity with a receptor; a score greater than 5.0 indicates a better binding activity; a score greater than 7.0 indicates a strong binding activity (Hsin et al., 2016). The PDB ID of the 10 genes was molecularly docked with the five compounds. The docking scores of all compounds and proteins were greater than 4.25, indicating the presence of binding activity. The molecular docking results are shown in Table 9.

A heat map of the molecular docking is shown in Fig. 6A. The deeper the color, the better the docking effect. The effect of molecular docking is also shown by the heat map. Table 9 and Fig. 6A showed that the compound with the highest binding activity to the protein receptor STAT3 was genistein; that with the highest binding activity to the protein receptor TP53 was kaempferol; that with the highest binding activity to the protein receptor AKT1 was kaempferol; that with the highest binding activity to the protein receptor AKT1 was quercetin; that with the highest binding activity to the protein receptor JUN was daidzein; that with the highest binding activity to the protein receptor VEGFA was genistein; that with the highest binding activity to the protein receptor SRC was ursolic acid; that with the highest binding activity to the protein receptor TNF was ursolic acid; that with the highest binding activity

to the protein receptor HRAS was ursolic acid; and that with the highest binding activity to the protein receptor CDKN1A was ursolic Acid.

The 3D and 2D structures of their docking are shown in Table 10. In the 2D docking diagram of the third column, there is a specific case where the ligand binds to the receptor, and the surrounding red bar represents a bond-free protein residue within the ligand; the structure linked by the bat is a ligand compound and is subjected to the body protein. Purple represents a ligand compound; yellow represents a receptor protein. Red font represents the binding mode between the ligand and the receptor; the green dotted line represents a hydrogen bond. In these docking results, the docking activity of the receptor protein AKT1 and the ligand ursolic acid was 8.331, which was bound by Met 227 (A), Glu234 (A), and Thr291 (A); the receptor protein CDKN1A and the ligand compound ursolic acid, with docking activity of 8.148, and bound by a hydrogen bond with Asn247 (B) and Met246 (B), with bond lengths of 3.12 and 2.76, respectively, as well as with Cys242 (B), Met243 (A), Asn239 (B), Leu137 (B), Cys238 (B), Arg175 (B), and Arg174 (B), which directly bound.

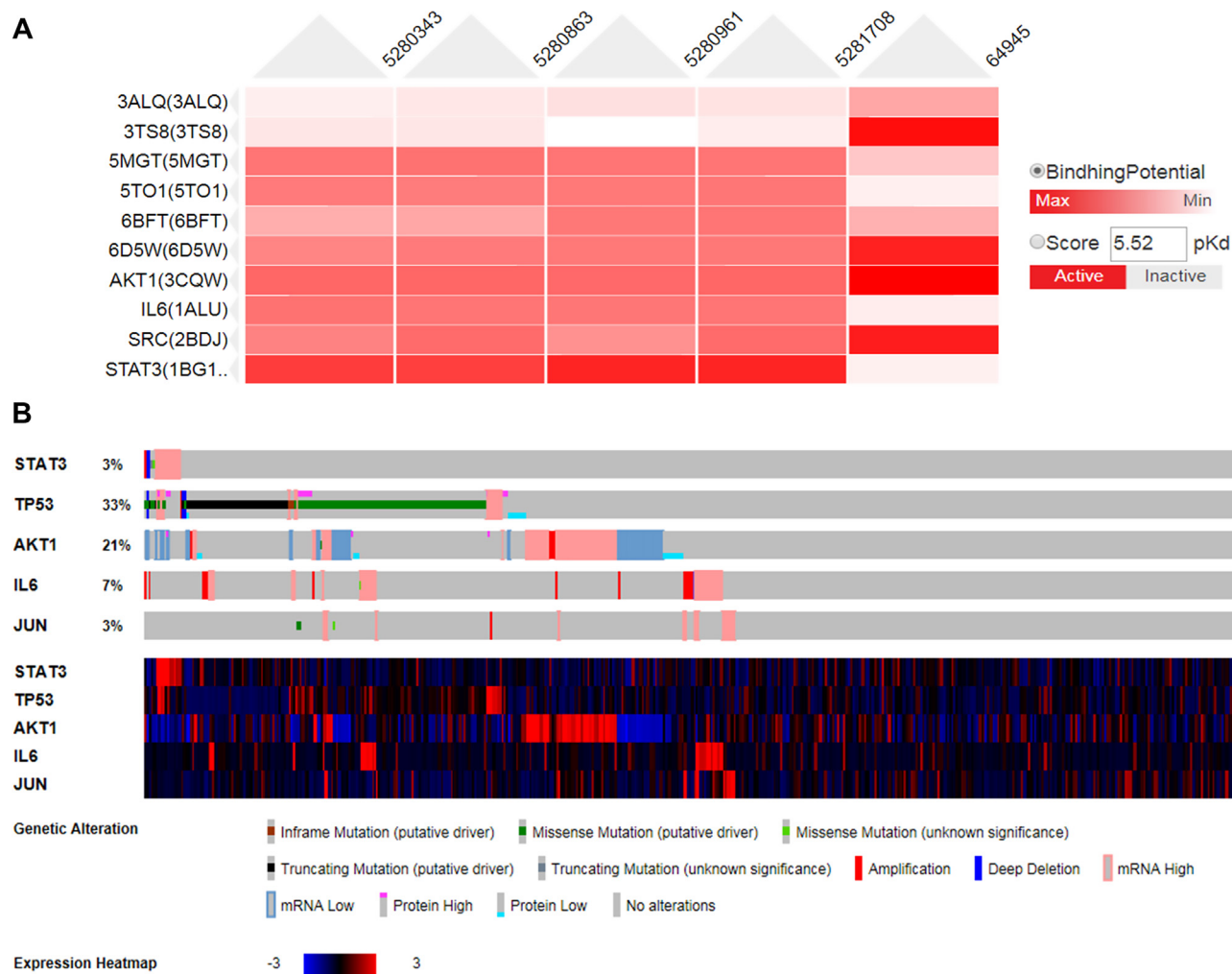
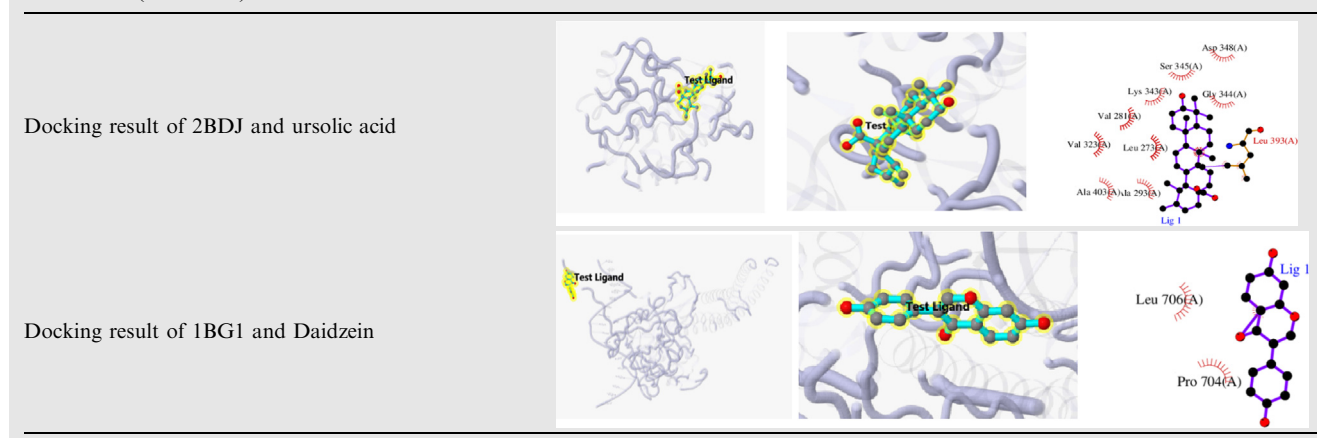


Fig. 6 Molecular docking. (A) A heat map showing the docking of receptor protein and ligand. (B) A diagram showing the genetic variation in lung squamous cell carcinoma.

Table 10 Molecular docking diagrams of the 3D and 2D structure.

| | | | |
|---|--|--|--|
| Docking result of 3ALQ and ursolic acid | | | |
| Docking result of 3TS8 and ursolic acid | | | |
| Docking result of 5GMT and kaempferol | | | |
| Docking result of 5TO1 and Daidzein | | | |
| Docking result of 6BFT and Genistein | | | |
| Docking result of 6D5W and ursolic acid | | | |
| Docking result of 3CQW and kaempferol | | | |
| Docking result of 1ALU and quercetin | | | |

(continued on next page)

Table 10 (continued)

3.7. cBioPortal analysis

cBioPortal is a database used for tumor research. The expression of the genes STAT3, TP53, AKT1, IL6, and JUN in 501 patients with lung squamous cell carcinoma was studied. The expression of these genes in the selected lung cancer samples is shown in Fig. 6B. The first part is the expression of genes, including inframe mutation, missense mutation, truncating mutation, amplification, deep deletion, mRNA level upregulation (mRNA High), mRNA level downregulation (mRNA Low), and no alterations. The second part is a heat map of the gene mRNA levels.

According to the data in Fig. 6B, TP53 and AKT1 were highly expressed in these 501 cancer patients. TP53 was mainly expressed in patients with mutations (129 cases, 25.75%), increased copy number (1 case, 0.2%), mRNA level upregulation (6 cases, 1.2%), high protein level (5 cases, 1%), low protein level (8 cases, 1.8%), and multiple alterations (17 cases, 3.39%). AKT1 was expressed mainly in patients with mutations (1 case, 0.2%), increased copy number (5 cases, 1%), mRNA level upregulation (34 cases, 6.79%), mRNA level downregulation (34 cases, 6.79%), high protein level (2 cases, 0.4%), low protein level (15 cases, 2.99%), and multiple alterations (16 cases, 3.19%).

4. Discussion

Guava leaves are medicinal herbs with various pharmacological effects and a wide range of research and development significance. Guava leaves also have certain antitumor effects. Lung cancer is a common tumor. Currently, there is no particularly effective method for the diagnosis and treatment of lung cancer. Image detection is a commonly used diagnostic method (Hong et al., 2019), followed by genetic diagnosis (Munne and Wells, 2002). Lung cancer has a very low cure rate, and recent approaches to treat lung cancer include immunotherapy, radiotherapy, and targeted therapy (Alasti et al., 2006; Petrosyan et al., 2012; Tsang et al., 2014; Zhang et al., 2019). In this study, the role of guava leaves in lung cancer was studied by network pharmacology to elucidate their correlation and provide a relevant basis for experimental research. In recent years, the pharmacology of traditional Chinese med-

icine has been continuously developed and has occupied a place in medical research.

From the research point of view, examination of the potential intersection of guava leaf potential target and lung cancer genes revealed 153 common genes, and the interaction between STAT3, TP53, AKT1, IL6, JUN, and VEGFA was the strongest. Using the 66 compounds in guava leaves and the 153 genes, we constructed a “guava leaf-constituent category-active constituent-gene” network, and our results showed that the compounds quercetin, genistein, apigenin, ursolic acid, daidzein, and lung cancer showed the strongest effect. The proteins with the strongest interaction were VDR, CDK2, MAP2K1, CDK6, MET, ABL1, PPARG, IL2, EGFR, and PGR, indicating that these genes interacted with most constituents of guava leaves, showing that these constituents have high antitumor activities.

GO annotation is an important means to examine the function of gene products (Leale et al., 2018). Through GO analysis, in biological processes, enrichment information includes apoptosis, programmed cell death, cell proliferation, etc.; in terms of cellular components, enrichment Information includes platelets, cell membranes, protein kinases, etc.; in terms of molecular function, it is mainly reflected in enzyme activity and regulation of enzyme activity.

KEGG enrichment analysis provides information on integrated metabolic pathways including metabolism, membrane trafficking, signal processing, and cell cycle. The PI3K-Akt signaling pathway is a typical tumor signaling pathway. In many primary and metastatic human cancers, PTEN activity is lost owing to mutations, deletions, or high-frequency silencing of promoter methylation. It is important for prediction in targeted therapy (Fresno Vara et al., 2004; Carnero et al., 2008; Ma and Hu, 2013). EGFR tyrosine kinases are important for the treatment of non-small cell lung cancer, including monoclonal antibodies. The current clinical representatives are cetuximab and panitumumab as well as EGFR-tyrosine kinase inhibitor. The combination inhibits the binding site of ATP and tyrosine kinase, thereby cutting the downstream signaling pathway and exerting antitumor effect (Toulabi and Ryan, 2018). EGFR is of great significance in the study of non-small cell lung cancer and has potential implications for drug therapy (Mead et al., 1980; Marchetti et al., 2005). In the small

cell lung cancer pathway, the tumor suppressor genes are p53, PTEN, RB, and FHIT. ECM activates membrane receptors, and through ITGA and ITGB conduction, activates FAK conduction to activate PI3K activity and express the PKB/AKT signaling pathway. Through continuous phosphorylation and activation of I κ B α , the free NF- κ B signal is activated into the nucleus for gene network regulation. STAT3 and VEGF may play a reverse regulatory role on the metastasis of lung cancer (Wang et al., 2011). The p53 gene also has an antitumor effect, and Ad-p53 combined with chemotherapy can reverse the chemoresistance of tumor cells and produce a synergistic antitumor effect (Matsubara et al., 2001; Meng and El-Deiry, 2002). In addition, there are other genes expressed in lung cancer that were targeted by guava leaves, representing a potential research direction as clinical anticancer targets.

Analysis of lung squamous cell carcinoma by the cBioPortal tumor database revealed that the genes targeted by Chinese medicine and disease genes are highly expressed in cancer patients, and can undergo mutations, such as addition and deletion, and expression regulation. TP53 and AKT1 were the two most strongly expressed genes in the cancer patients surveyed. Hence, different compounds in guava leaves regulated the same target, and one compound regulated multiple targets and participated in multiple pathways and biological processes, reflecting the multi-target and multi-channel characteristics of guava leaves.

5. Conclusions

Through network pharmacology research, we found that guava leaves had potential targets that interacted with various tumors, regulating the signaling pathways of cancers. The PI3K-Akt signaling pathway, an important signaling pathway in the study of tumor processes, regulates the proliferation of tumor cells and plays a role in tumor cell migration, tumor adhesion, tumor angiogenesis, and extracellular matrix degradation (Chatterjee et al., 2013; Liu et al., 2014; Wang et al., 2016a). The TP53 gene is an important tumor suppressor gene involved in many gene network regulation processes, and has a reference significance in the research and development of new drugs and targeted therapy.

Many of the pharmacological effects of guava leaves are yet to be developed. There are many compounds in guava leaves. At present, antitumor studies on guava leaves are still lacking, although some studies have shown that guava leaves exert antitumor effects. This study preliminarily verified the pharmacological basis and the related mechanism of the antitumor effect of guava leaves, providing a foundation for further research. It is also hoped that specific experiments will be carried out to verify the pharmacological effects of guava leaves against tumors. If successful, the lack of cancer research on guava leaves will be overcome. In addition, network pharmacology is a hot topic of research; we hope to conduct more research projects in the future to provide a basis for future research on new drugs.

6. Data availability

The data used to support the findings of this study are included within the article.

Author contribution

LJ has made substantial contributions to conception and design, interpretation of data, and manuscript writing. WJ has made substantial contributions to acquisition and analysis of data. YQ and JL received the funding supporting for the study. YW has made substantial contributions to acquisition of data.

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Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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