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ORIGINAL ARTICLE

Controlling the diffusion of micro-volume Pb solution on hydrophobic polyurethane membrane for quantitative analysis using laser-induced breakdown spectroscopy (LIBS)



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KEYWORDS

Drop-wising; LIBS; Liquid–solid conversion; Membrane; Polyurethane **Abstract** This work reports a novel controlling mechanism of analyte diffusion in a micro volume solution (100 μ L) into a hydrophobic membrane. This study was designed to facilitate the liquid–solid conversion using membrane for laser-induced breakdown spectroscopy (LIBS) in quantitatively analyzing aqueous lead (Pb) pollutant. Herein, we used the same analyte (Pb) solution applied on one side of the membrane (*back* side) to enhance the diffusion of the analyte administered from the other side (*front* side). The membrane was confirmed hydrophobic with contact angles ranged from 104.6° ± 1.3° to 106.28° ± 1.7°, where its morphology had smooth surface and randomly distributed small pores. We found the limit of detection (LOD) to reach 184.2 mg/L derived from a

Abbreviations: AAS, Atomic absorption spectroscopy; BET, Brunauer-Emmett-Teller; BJH, Barret-Joyner-Halenda; CPUM, Castor oil-based polyurethane membrane; DDG, Digital delay generator; ICCD, Intensified charge-coupled device; LIBS, Laser-induced breakdown spectroscopy; LOD, Limit of detection; LSC, Liquid-to-solid conversion; MDI, 4,4'-diphenylmethane diisocyanate; RMSE, Root-mean-square errors; SEM, Scanning electron microscopy; TDI, Toluene diisocyanate.

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calibration curve with Pb I (405.7 nm) line intensity as the dependent variable, where the rootmean-square-errors (RMSE) and correlation (\mathbb{R}^2) were 1.08 M and 0.999, respectively. In comparison, the membrane *back* side with distilled water achieved LOD as low as 134.53 mg/L obtained from the similar calibration curve (RMSE = 5.8 M; $\mathbb{R}^2 = 0.986$). Further analysis using the LIBS spectra confirmed the role of the analyte ion on the *back* side of the membrane in enhancing the analyte diffusion.

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1. Introduction

Laser-induced breakdown spectroscopy (LIBS) has gained many scholars' attention in the field of analytical chemistry (Anabitarte et al., 2012). The technique is capable of performing multi-elemental analysis with good accuracy and precision via micro ablation sampling for metal and solid materials. Moreover, LIBS has a practical advantage that requires no chemical solvents and is often considered as an eco-friendly analytical tool (Hussain Shah et al., 2020). As of today, LIBS has been developed to carry analysis on much more complicated substrates; soft organic samples such as milk powder (Iqbal et al., 2018), coffee bean (Abdulmadjid et al., 2017; Nasution et al., 2019), raw fish (Marpaung et al., 2020), and saliva (Zahroh et al., 2019). The main challenge in LIBS technique is the presence of matrix effects which could be derived from the physical and chemical properties of the substrate (Guezenoc et al., 2019). This challenge is primarily obtained when handling liquid samples, where water evaporation and splashing are expected.

Overcoming the matrix effects of liquid samples is the key to enabling LIBS applicability in environmental monitoring for water pollution. Some have tried enhancing the water surface hardness by freezing (Cáceres et al., 2001) or forming a jet flow (Cheri and Tavassoli, 2011). Others have modified the system by employing double-pulse laser (Zhao et al., 2019). However, complex preparation and equipment settings are the major drawbacks of those foregoing alternatives. Thus, our group has developed a simpler sample pretreatment technique that relies on liquid-to-solid conversion (LSC) using adsorbent mediums (Iqhrammullah et al., 2021; Iqhrammullah et al., 2020a). Similar efforts have also been introduced by many studies (Fu et al., 2020; Lee et al., 2020; Lin et al., 2016; Nam et al., 2018; Ripoll et al., 2021; Wang et al., 2015), of which, the conversion carried out via drop-wising is the most practical requiring only small volume (micro level) of analyte solution (Rupiasih et al., 2013).

In this work, we used a polyurethane membrane - a scarcely used membrane for LSC in LIBS qualitative analysis. The membrane was prepared by employing castor oil (Ricinus communis L.) as the polyol source and toluene diisocyanate (TDI) as the isocyanate source (Marlina et al., 2020, 2017). To enable the attachment of analyte onto the membrane surface, a complexing agent, 1,10-fenantroline, was added as a filler (Nisah et al., 2021). The membrane produced has a hydrophobic characteristic derived from the castor oil properties (Iqhrammullah et al., 2020a; Marlina et al., 2019). The hydrophobicity contributes to the durability of the membrane along with its prominent mechanical strength and flexibility (Chakraborty and Chatterjee, 2020). More importantly, hydrophobic feature prevents the absorbance of water molecules from ambient air into the membrane. Low content of water could essentially reduce the water vapor-associated plasma quenching during LIBS analvsis. Unfortunately, this hydrophobicity also hinders the analyte diffusion that is counterproductive to our objective.

The diffusion allows the analyte mobility across the entire membrane inner surface which is important for the homogeneity. Transmembrane movement may depend on ion-ion interaction (Zhang et al., 2019). In a forward osmosis membrane, the draw solution's ionic charges have been associated with the ion trans-membrane mobility (Irvine et al., 2013; Saiful et al., 2020). Thus, we hypothesized that a certain solution on one side of the membrane allows us to control the analyte diffusion from the other side of the membrane. For this purpose, we employed Pb ion as the ionic analyte model, as it is considered the most common heavy metal pollutant found in areas with intense industrial activities (El Azhari et al., 2017; Li et al., 2020; Yulianto et al., 2020). Prior to the wastewater treatment, concentration of Pb could reach hundreds of mg/L, and it is difficult for conventional method such as atomic absorption spectroscopy (AAS) as it requires multiple dilution. Hence, our method is proposed to monitor Pb concentration produced from the industrial activities. The study has a significance of employing a small amount of sample (as low as 100 μ L), which contributes to the practicality of the proposed method.

2. Materials and methods

2.1. Materials

The main requirement in preparing polyurethane material is polyol and isocyanate sources. The polyol source used to prepare the membrane was industrial grade castor oil, extracted from *Ricinus communis* L. (purchased from PT. Rudang Jaya, Medan, Indonesia). As for the isocyanate source, this study employed toluene diisocyanate (TDI). To bind the analyte on the membrane surface 1,10-fenantroline was used as a complexing agent and acted as a filler. Other chemicals used in this study were HNO₃, KOH, acetone and Pb(NO₃)₂. Otherwise mentioned, all the used chemicals were analytical grade and purchased from Merck (Selangor, Malaysia).

2.2. Preparation of hydrophobic polyurethane membrane

The substrate used in this study for liquid-solid conversion was castor oil-based polyurethane membrane (CPUM). The preparation employed a one-shoot method (Iqhrammullah et al., 2020b; Marlina et al., 2020), where castor oil (3.5 g) and 1,10-fenantroline (5 mg) were firstly mixed and stirred for 15 min at 250 rpm. The mixture was then added with 1.75 g TDI to obtain the doping solution, heated at 50 °C and 280 rpm for 15 min. The doping solution was sonicated using Crest Genesis[™] XG-500-6 ultrasonic (40 kHz; 500 W) along with the addition of 2 g acetone. After it is completely dissolved in acetone, the solution was casted on a glass plate and heated in the oven at 40 °C for 24 h. The solid film membrane was then released from the glass plate with the help of acetone and left cold for 5 h. The CPUM was cut into a circle shape with diameter of 2 cm for further use. The preparation of CPUM has been illustrated in Scheme 1.

2.3. Characterization

Prepared CPUM was characterized for its morphology using Jeol. Jsm-6510 LA scanning electron microscopy (SEM) (Tokyo, Japan), run at 10 kV with 20,000x magnification. Measurement of contact angle was performed on Biolin Scientific Attension Goniometer (Uusimaa, Finland) in triplicate, where the solution pH was adjusted to 7. Crater from the laser ablation was observed under Stereomicroscope Primo Star Zeiss (Jena, Germany) with 40x magnification. Total surface area and total pore volume of the priorly dried CPUM were determined based on Brunauer-Emmett-Teller (BET) and Barret-Joyner-Halenda (BJH) isotherms of adsorption–desorption of N₂ at 77.3 K, performed on QuadraSorb Station 1 (ver. 5.06) (Quantachrome, FL, USA).

2.4. Analyte loading

Pb solution was prepared by diluting the Pb(NO₃)₂ stock solution into a desired concentration, ranging from 0.1 to 0.7 M. The pH of the analyte solution was maintained at 7 using KOH and HNO₃. For the purpose of this investigation, we specify the two sides of the membrane; the *back* side and the *front* side. The *back* side was where the solution concentration remained constant, meanwhile at *front* side, the analyte concentration was varied. The Pb solution (0.1 M) and distilled water, as much as 100 μ L, was exposed to the *back* side by drop-wising for 30 min. Membrane receiving Pb solution 0.1 M was labelled as CPUM0.1 and the one with distilled water was labelled as CPUM0. Afterwards, the analyte (100 μ L) with a varied concentration was loaded onto the *front* side of the membrane by drop-wising and left for 30 min, before dried and stored for further analysis.

2.5. LIBS analysis

LIBS system used a 1064 nm Q-switched Nd:YAG laser (Quanta-Ray, LAB 130-10, USA) operating at 8 ns width and 10 Hz frequency. The laser beam was reflected toward the vertically positioned sample surface passing through a focus lens (170 mm). The sample was rotated at 5 rpm during the ablation. The signal from the plasma was recorded using an optical fibre located 5 cm from the sample and right angle to laser beam. The signal collected was then processed using a spectrometer (McPherson model 2061, focal length 1000 mm, f/8.6, Czerny Turner configuration) with an intensified charge-coupled device (ICCD) camera (Andor 117 iStar' intensified CCD 1024 \times 256 pixels, UK). Laser and ICCD camera were controlled using a digital delay generator (DDG 535, Stanford Research System, 119 USA). The system was run in air at 1 atm, where each run consists of 10 data accumulations.

3. Results and discussion

3.1. Characteristics of the polyurethane membrane

The CPUM produced from the one-shot method in this study had a thickness of around 0.017 ± 0.001 mm. Results from SEM investigation on the membrane surface from its *front* and *back* sides have been presented (Fig. 1a,b). The morphology of the membrane *front* side was similar to that of the *back* side, where both had smooth surface. The presence of ran-



Scheme 1 Preparation of CPUM from castor oil and TDI using one-shoot method.

domly distributed pores was observed at both sides of the membrane with diameters reaching around 1.7 μ m. Some of 1,10-fenantroline particles formed agglomerates which is observable at both sides of the membrane. Our current findings share a similarity with those from previously reported, such as the one synthesizing CPUM with red seaweed filler (Nurman et al., 2021). In our previous studies using 4,4'-diphenylmethane diisocyanate (MDI) as the isocyanate source, the surface morphology also appeared smooth along with a dense structure (Iqhrammullah et al., 2020; Iqhrammullah et al., 2020b).

The calculation using BET and BJH isotherm on the adsorption–desorption of N₂ revealed that the membrane had a total surface area of $32.56 \text{ m}^2/\text{g}$ and a total pore volume of 0.035 cm³/g. These values are close to those reported in a study synthesizing polyurethane membrane prepared from MDI and *Chaetomorpha Linum*-derived polyol (Marlina et al., 2020). Based on the contact angle formed between the water droplet (pH 7) and the membrane surface, both the *front* and *back* sides had similar contact angle values of more than 90° (104.6°±1.3° and 106.28°±1.7°, respectively), confirming their hydrophobicity (Fig. 1c). Previously, a polyurethane foam made of castor oil-derived polyol was reported to have similar hydrophobicity (Vieira Amorim et al., 2021). However, CPUM was also reported with hydrophilic characteristic (contact angle < 90°) (Macalino et al., 2017). The author argued

that the hydrophilicity was the results of the polymerization using aliphatic isocyanate – 1,6-hexamethylene diisocyanate (Macalino et al., 2017). Meanwhile, hydrophobic feature of castor oil-based polyurethane was obtained when cyclic isocyanate was used (Vieira Amorim et al., 2021), including that in this present study. In addition to the stated characteristics, the crater formed after laser ablation was also presented (Fig. 1e). The laser ablation was shown to have left a crater on the membrane surface with a diameter of 68.39 μ m.

3.2. Optimization of LIBS parameters

In order to reduce the matrix effects, we studied and optimized the LIBS parameters *viz* delay time and laser energy. Delay time adjustment is used to discriminate the continuum noise resulted from hot plasma (Anabitarte et al., 2012). Longer delay time allows the signal to be captured where the continuum noise is at minimum. However, extending more delay time will allow the plasma to decay which eventually leads to lower intensity. To study this effect of delay time variation, the experiment was conducted at constant laser energy of 122 mJ. In our case (Fig. 2a), delay time of 1 μ s is sufficient to distinguish the signal from analyte with that of continuum noise. Usually, the 3 μ s was reported to be the optimum delay time for 1 atm open-air LIBS system (Marpaung et al., 2019). However, at 2 μ s, the intensity decreased indicating the decay



Fig. 1 SEM images of the (a) *front* and (b) *back* sides of the membrane observed under 5,000x magnification. Red arrows and red circles indicate pores and 1,10-fenantroline agglomerates, respectively. Contact angle measured on the surface of (c) *front* and (d) *back* sides of the membrane measured at pH 7. (e) Crater created after the laser ablation on the membrane surface with the longest diameter of 68.39 μm.

process (Fig. 2a). These different findings might be attributed to the differences in the membrane substrate properties that affect the plasma characteristics – known as substrate-based matrix effects.

Other LIBS parameter optimized in this study is laser energy, as it determines the amount of ablated substrate. There is a *give-and-take* when it comes to this parameter; lower energy is less destructive and the noise could be kept minimum, but higher energy is required to generate higher signal intensity (Abdulmadjid et al., 2015). To study the influence of laser energy, the experiment was conducted at constant delay time of 1 μ s. As shown in Fig. 2b, the highest laser energy level, 122 mJ, yielded the highest intensity with more background noise. However, the high intensity could compensate the background noise very well. The destruction level was also found to be tolerable against the membrane substrate, allowing us to perform multiple repetitions upon the analysis. Therefore, based on the explanation above, the optimum laser parameters



Fig. 2 Spectra of Pb 0.7 M-loaded CPUM0.1 analyzed under different (a) delay times and (b) laser energy levels. Delay time variation was conducted at constant laser energy of 122 mJ and laser energy levels – at constant delay time of 1 μ s. (c) Identification of elements contained in CPUM and Pb 0.7 M-loaded CPUM0 and CPUM0.1 based on LIBS spectral profile obtained at 1 μ s and 122 mJ.

taken for further investigations were 1 μ s delay time and 122 mJ laser energy.

3.3. Qualitative analysis

LIBS is commonly used for its ability in simultaneously analyzing multiple elements in a substrate. We have conducted analyses on the CPUM0.1 and CPUM0 after the analyte loading with the highest concentration (0.7 M), then compared the spectra with that of pure membrane (CPUM) to qualitatively see whether the analyte could be attached to the membrane substrate (Fig. 2c). As shown, only loaded CPUM0.1 and CPUM0 generate the Pb I (405.7 nm) spectral line suggesting that the analyte has been embedded onto the membrane substrate. Other elements, such as K, Ca, and Mg were found to be present in the substrate which can be originated from the human skin contact (Yang and Yoh, 2018), the contaminated analyte solution, or impurities from the membrane ingredient. It is also interesting to note that the membrane is an organic substrate containing O, N, C, and H which give the spectra with a unique characteristic - possessing many molecular bands (Abdulmadjid et al., 2017; Marpaung et al., 2020). CN bands are among the most observable molecular bands, especially the CN (0-0) at 388.3 nm that has the tallest vibrational intensity.

3.4. Quantitative analysis

In order to perform a quantitative analysis, the membrane substrate should be loaded with a variation of analyte concentration ranging from 0.1 to 0.7 M. Higher analyte concentration consequently generate higher spectral intensity, observed from the LIBS spectra obtained from Pb-loaded CPUM0.1 (Fig. 3a) or CPUM0 (Fig. 3b). However, to overcome the uncertainty in LIBS measurement, normalizing the value with internal standard lines could be conducted (Wei et al., 2015). Herein, we selected the spectral lines from C and CN as they might be originated from the membrane and are not varied in concentration from one substrate to another (Iqhrammullah et al., 2020a). Other than the spectral lines, noise signal intensity could also be one of the options to compensate the uncertainty in LIBS. The noise signal was selected from the nearest vicinity of the Pb I (405.7 nm) spectral line. However, the contribution of other external or internal factors, such as CO_2 from breathing and ambient N₂, may affect the quality of the calibration curve as well. This is why we conducted the investigation by constructing the calibration curve using all possible normalization.

In the case of CPUM0.1, among the four callibration curve models, the correlation (R^2) closest to 1 was found from the one constructed based on Pb I (405.7 nm) spectral line intensity (Fig. 4a). Poorer correlation was observed when the normalization was conducted. Therefore, we concluded that the generated Pb spectral line was stable and requiring no further normalization using internal standard lines.

In contrast with CPUM0, the best correlation was obtained from the ratio of Pb I (405.7 nm) and C I (247.8 nm) spectral lines (Fig. 5c). If the uncertainty was caused by the unstable fluence or plasma temperature, the similar phenomenon should be expected in the case of CPUM0.1. This difference lead us to question the role of analyte interraction with the surface of membrane substrate. In our previously conducted investigation (Ighrammullah et al., 2021), we found that the different analyte interraction could be observed by varying the laser energy. In another previous study, we also found that Pb may form a multilayer interraction (Iqhrammullah et al., 2020b), where dissolved Pb particles are piled up in one binding site creating a distance to the surface (Chen et al., 2019; Rahmi et al., 2021). As a consequence, the particles are less attracted to the surface and become unstable, which is then sensitive to the change of plasma temperature. In the next section, we will further discuss the probable causes of the interaction attributed to the administration of Pb solution onto the back side of the membrane (CPUM0.1).

The comparison of analytical parameters obtained from each calibration curve from CPUM0.1 and CPUM0 has been presented (Table 1). A good quality calibration curve can be determined by high correlation (\mathbb{R}^2) value and low rootmean-square errors (RMSE) value. In CPUM0.1, the calibration curve with Pb I (405.7 nm) as the *y*-axis has the highest \mathbb{R}^2



Fig. 3 Pb I (405.7 nm) LIBS spectral lines after the loading of analyte with different concentrations onto (a) CPUM0.1 and (b) CPUM0.



Fig. 4 Calibration curve for aqueous Pb analysis using (a) Pb I (405.7 nm), (b) Pb I (405.7 nm)/Noise, Pb I (405.7 nm) /C I (247.8 nm), and Pb I (405.7 nm)/ CN (388.3 nm) γ -axes based on the LIBS spectra of CPUM0.1.

(0.999) and the lowest RMSE (1.08 M). Therefore, the limit of detection (LOD) was calculated based on the calibration curve with the formula of three times blank reference standard deviation divided by the slope. The LOD was obtained to be 8.9×10^{-4} M or 184.2 mg/L. As for the CPUM0, by employing the same calculation on the calibration curve with Pb I (405.7 nm) intensity as the *y*-axis (R² = 0.986; RMSE = 5.8 4 M), the LOD was obtained to be 6.5 × 10⁻⁴ M or 134.53 mg/L, which is better than that of CPUM0.1. This is due to the fact that the CPUM0 has higher Pb emission, whereas at the analyte concentration of 0.1 M, the spectral line intensity has reached 1840.92 a.u. Meanwhile, in CPUM0.1, the average intensity of Pb I (405.7 nm) spectral line was only 519.05 a.u.

Indeed, some research groups have reached LOD in parts per billion levels (Lin et al., 2016; Ripoll et al., 2021), suggesting that our system still needs improvement. Our current inves-

tigations (Iqhrammullah et al., 2021; Iqhrammullah et al., 2020a), along with some others (Lee et al., 2012; Schmidt and Goode, 2002; Zhu et al., 2012), obtained the LODs within 1 - 10 mg/L range. Other published studies reported LODs exceeding more than 20 mg/L in their systems (Horňáčková et al., 2019; Suyanto et al., 2016; Terán et al., 2019; Yang et al., 2018). Taken altogether, our system is still not a match to the previously mentioned studies in term of the sensitivity (LOD). Nonetheless, our system is potentially more practical because it only requires a small volume of analyte $(< 100 \ \mu L)$. Based on the current results, our proposed method could be useful for Pb monitoring in industrial setting. Moreover, in forensic investigation (where the analyte is typically available in a small amount) the method proposed herein could be useful, especially by considering the fact that acute toxicity of Pb could only be induced if the concentration is high (reaching hundreds of mg/L) (Besser et al., 2005). Further studies will



Fig. 5 Calibration curve for aqueous Pb analysis using (a) Pb I (405.7 nm), (b) Pb I (405.7 nm)/Noise, Pb I (405.7 nm) /C I (247.8 nm), and Pb I (405.7 nm)/ CN (388.3 nm) y-axes based on the LIBS spectra of CPUM0.

be carried out to lower the LOD of our system. One of the alternatives to improve the analytical performance of our system in the future research is by constructing a non-linear calibration curve as reported previously (Zhou et al., 2021).

3.5. Analyte mobility and diffusion

Previously, we have explained that the Pb ions are stacked in multilayer on a binding site of the membrane surface. It may be due to the fact that mobility of Pb solution was restricted inside the membrane due to the hydrophobic nature of the membrane. As a result, Pb was unable to diffuse into the deeper side of the membrane and became concentrated. This condition permits the Pb analyte to have multilayer interaction with the membrane surface. On the contrary, when Pb solution was applied on the other side of the membrane, it consequently eased the analyte mobility allowing the ions to be homogeneously distributed on the membrane surface preventing the multilayer interaction.

To prove this theory, we had conducted an investigation on the *back* side of the membrane (Fig. 6). Pb emission was only detected on the CPUM0.1 (Fig. 6a), where its intensity corresponds to the analyte loading concentration. It means Pb moved from the *front* side to the *back* side, where the other Pb ions were present. As for CPUM0 (Fig. 6b), there might be a small concentration of Pb that passed through the membrane layers, but it was not sufficient to generate the signal that is three times higher than noise intensity. This investigation proves that the diffusion has been enhanced in CPUM0.1.

A clearer depiction of the changes of Pb emission intensity obtained from the ablation of the *back* side surface of the membrane, affected by the analyte loading with varied concentration, can be seen in Fig. 7. Interestingly, without the presence of Pb on the other side (distilled water), the Pb mobility

Y-axis	CPUM0.1					СРИМО		
	SC (M)	CC ± Error (M)	RMSE (M)	R ²	SC (M)	CC ± Error (M)	RMSE (M)	R ²
Pb I (405.7 nm)	0.1	0.10 ± 0.07	1.08	0.999	0.1	0.13 ± 0.22	5.84	0.986
	0.3	$0.30~\pm~0.06$			0.3	$0.26~\pm~0.20$		
	0.5	$0.48~\pm~0.06$			0.5	$0.49~\pm~0.20$		
	0.7	$0.71~\pm~0.05$			0.7	$0.72~\pm~0.18$		
Pb I (405.7 nm) /Noise	0.1	$0.17~\pm~0.57$	8.53	0.875	0.1	$0.18~\pm~0.56$	5.11	0.871
	0.3	0.24 ± 0.54			0.3	0.22 ± 0.55		
	0.5	0.41 ± 0.52			0.5	0.42 ± 0.52		
	0.7	$0.78~\pm~0.45$			0.7	$0.78~\pm~0.40$		
Pb I (405.7 nm) /C I (247.8 nm)	0.1	0.13 ± 0.63	2.15	0.867	0.1	$0.10~\pm~0.03$	0.21	0.999
	0.3	0.33 ± 0.56			0.3	0.31 ± 0.03		
	0.5	0.36 ± 0.55			0.5	$0.50~\pm~0.03$		
	0.7	$0.78 ~\pm~ 0.84$			0.7	$0.70~\pm~0.04$		
Pb I (405.7 nm) / CN (388.3 nm)	0.1	0.12 ± 0.25	1.43	0.988	0.1	$0.01 ~\pm~ 1.08$	18.33	0.735
	0.3	0.29 ± 0.22			0.3	$0.32~\pm~0.98$		
	0.5	0.45 ± 0.22			0.5	0.72 ± 1.08		
	0.7	$0.73~\pm~0.19$			0.7	0.55 ± 1.01		

Table 1 Analytical parameters obtained from the constructed calibration curves.



Fig. 6 Pb I (405.7 nm) LIBS spectral lines obtained from the laser shot (1 µs; 122 mJ) at the *back*-sides of (a) CPUM0.1 and (b) CPUM0 with different drop-wised analyte loading concentrations.

was constrained, associated with higher spectral intensity. It is also supported by our previous findings that CPUM0 yielded higher intensity of Pb I (405.7 nm) spectral emission. Nonetheless, due to unstable interaction between the analyte and membrane surface, the errors are higher. Therefore, even though the LOD might be worse, controlling the analyte – substrate interaction is also important to obtain a stable and accurate measurement. Indeed, this problem could be overcome by normalization using internal standard line just like we did in this study. However, there are several cases where it is not possible, for instance, sample containing high organic materials may change the total carbon concentration.

The above stated results were further confirmed by investigating the spatial distribution of Pb analyte in the CPUM0.1 and CPUM0, where the data have been presented in Table 2. Spatial distribution was investigated by allowing the laser shot to target a pre-determined spot with a variation of distances from the center (0–0.6 cm). We found even distribution of Pb analyte in CPUM0.1, proven by the statistical insignificance (P = 0.2279). On contrary, CPUM0 system had a significant variation of Pb I (405.7 nm) intensities (P = 0.003). These data suggest that analysis using CPUM0.1 system had a better precision.

Depth profile of Pb contained in CPUM0.1 and CPUM0 has been presented (Table 2). The investigation was carried out on the same ablated surface for 3 runs, where each run lasted for around 2 s. Hence, we were able to calculate that each run should consist of 20 laser shots. As can be seen in Table 2, reduction of the Pb I (405.7 nm) intensity fell greatly (P < 0.001) after the second run (40 shots) and further depleted following the third run (60 shots). As in the case of CPUM0, Pb I (405.7 nm) was immediately undetectable in



Fig. 7 Spectral intensities of Pb I (405.7 nm) line obtained from the laser shot (1 μ s; 122 mJ) at the *back*-sides of CPUM0.1 with different drop-wised analyte loading concentrations. The red-dashed line is the theoretical data indicating the correlation between the analyte loads and the spectral line intensity.

Table 2	Spatial of	distribution	and dep	h profile	of the	analyte
(Pb) in C	PUM0.1	and CPUM	[0.			

Experiment	Pb I (405.7 nm) intensity (a.u.)					
	CPUM0.1	CPUM0				
Spatial distribution, distance from the center (cm)						
0	7110.67 ± 61.33	8569.33 ± 114.52				
0.2	7027.22 ± 66.1	9027.21 ± 180.42				
0.3	7039.33 ± 56.58	7413.38 ± 234.27				
0.4	7190.82 ± 66.64	8394 ± 218.1				
0.5	7113.67 ± 56.05	8133.61 ± 163.57				
0.6	7143.33 ± 61	8372.47 ± 119.6				
P-value	0.2279	0.003*				
Depth profile, number of shots						
20	7168	9048				
40	5801	-				
60	2695	-				
P-value	< 0.001**	NA				

Number of shots were roughly calculated by ablation duration (2 s) multiplied by laser frequency (10 Hz). *P*-value was obtained from one-way ANOVA. * significant at P < 0.05. ** very significant at P < 0.001. (-) not detected, the intensity could not be distinguished from the noise (3 times higher than noise). NA: not applicable.

the second run. Taken altogether, these data corroborate the Pb analyte mobility in the CPUM0.1, where on the contrary, the Pb analyte was more likely to be retained on the *front* side of CPUM0.

4. Conclusion

Diffusion of micro-volume Pb^{2+} analyte into a hydrophobic membrane surface during the LCS could be controlled by applying either distilled water or ionic solution. Preventing the analyte diffusion which could lead to the enhancement of analyte on the membrane surface via multilayer binding, where it could be achieved by applying distilled water on the other side of the membrane. On contrary, diffusion could be promoted if the *back* side of the membrane was administered with an ionic solution. It is still unclear whether the ionic solution should be the same with that of the analyte. Further investigations need to carry out to elucidate other factors which could affect the diffusion of microvolume analyte into a hydrophobic membrane.

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