



ORIGINAL ARTICLE

SPIONs as a nanomagnetic catalyst for the synthesis and anti-microbial activity of 2-aminothiazoles derivatives



Samar A. Abubshait^{a,b,*}, Haya A. Abubshait^c, Sara Nabil^{a,b}, Asma M. Elsharif^a, Hamad M. Alkahtani^d, Fadilah Sfouq Aleanizy^e, M. Nasiruzzaman Shaikh^f

^a Department of Chemistry, College of Science, Imam Abdulrahman Bin Faisal University, P.O. Box 1982, Dammam 31441, Saudi Arabia

^b Basic and Applied Scientific Research Center, Imam Abdulrahman Bin Faisal University, P.O. Box 1982, Dammam 31441, Saudi Arabia

^c Basic Sciences Department, Deanship of Preparatory Year and Supporting Studies, Imam Abdulrahman Bin Faisal University, P.O. Box 1982, Dammam 31441, Saudi Arabia

^d Department of Pharmaceutical Chemistry, College of Pharmacy, King Saud University, P.O. Box 2457, Riyadh 11451, Saudi Arabia

^e Department of Pharmaceutics, College of Pharmacy, King Saud University, Riyadh, Saudi Arabia

^f Interdisciplinary Research Center for Hydrogen and Energy Storage (IRC-HES), King Fahd University of Petroleum and Minerals (KFUPM), Dhahran 31261, Saudi Arabia

Received 12 January 2022; accepted 28 March 2022

Available online 1 April 2022

KEYWORDS

Nanomagnetic catalysis;
2-aminothiazole derivatives;
Pyrazolyl derivatives;
Schiff bases;
Aromatic aldehyde;
Anti-microbial activity

Abstract A series of aminothiazole derivatives have been synthesized by using ultrasml superparamagnetic iron oxide nanoparticles (SPIONs) nanomagnetic catalysis, which were prepared by reducing the Fe(II) and Fe(III) precursors using aqueous ammonia then characterized by the XRD, FTIR, SEM, and TEM. The 2-aminothiazole derivatives were obtained by coupling 2-aminothiazole diazonium salt with active methylene compounds then cyclization with hydrazine hydrate to afford pyrazolyl derivatives. The one-pot reaction of 2-aminothiazole with an aromatic aldehydes in the presence of Fe₃O₄ NPs to give Schiff bases derivatives. An efficient protocol is developed proudest yields and reduction reaction time and easy separation. Therefore, all synthesized compounds were evaluated for anti-microbial activity.

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* Corresponding author.

Peer review under responsibility of King Saud University.



1. Introduction

Thiazole is a core structural element plays an important role in nature and has a wide range of applications in medicinal chemistry (Xue et al., 2014). Thiazole heterocycle is a main

structural motif of many natural compounds such as vitamin B1 (thiamine), penicillin, and carboxylase (Zhu et al. 2012; Das et al., 2016). 2-Aminothiazoles are one of the most important classes of heterocyclic compounds, that contain nitrogen and sulfur are present in compounds possessing scaffolds due to the medical and pharmaceutical applications (Das et al., 2016; Smith et al., 2012; Giridhar et al., 2001; Ali and Sayed, 2021; Hussein et al., 2020), such as antihypertension (Patt et al., 1992), antibacterial (Joseph et al., 2017), anti-inflammation, antiviral (Venkatachalam et al., 2001), antimycobacterial (Makam et al., 2013; Elsadek et al., 2021; Dayan et al., 2021), anticonvulsant (Siddiqui and Ahsan, 2010), antileishmanial (Bhuniya et al., 2015), HIV infections (Bell et al., 1995), anticancer (Elsadek et al., 2021; Chugunova et al., 2021; Kuzhandaivel et al., 2021; Özbek and Gürdere, 2021), antitumor (Hersi et al., 2020), antidiabetic (Iino et al., 2009) and antioxidative (Uchikawa et al., 1996). Therefore, in recent times, increasing attention has been paid to the synthesis of heterocyclic compounds the development of an environmentally benign and efficient procedure for the synthesis of 2-aminothiazoles derivatives (Ali et al., 2010) has become particularly fascinating and remains a great challenge. Recently, nanomagnetic catalysts have been used as heterogeneous green catalysts. However, this study focuses on the development synthesis of 2-aminothiazoles derivatives by using the nanomagnetic catalyst for the effective synthesis via the one-pot reaction.

As ideal supports and nanocatalysts have attracted much attention in catalytic processes, due to their intriguing nanoscale dimensions, high activity, low cost, high surface area, non-toxicity, magnetically separation from the reaction media, and easy modification with other organic or inorganic species (Sadeghi et al., 2016; Azgomi and Mokhtary, 2015; Zarnegar and Safari, 2016). In recent years, magnetic nanostructures such as Fe₃O₄ nanoparticle (Fe₃O₄ NPs) was applied in the presence of atmospheric air as a green efficient, heterogeneous, and reusable catalytic system for synthesis of the heterocyclic compounds (Zolfigol et al., 2012; Bhaskaruni et al., 2020; Eshfahani et al., 2011; Polshettiwar and Varma, 2010). Although, this efficient greener protocol and cleaner conditions, milder, shorter reaction time, an excellent yield of products higher purity and easier work-up procedure, easy separation using a simple external magnetic field, low cost and operational simplicity promoted us to developed novel 2-aminothiazoles derivatives by using Fe₃O₄ NPs.

2. Experimental

2.1. Materials and methods

Melting points utilized via Stuart SMP30 Digital Advanced MP apparatus were taken in open capillary and uncorrected. FTIR spectra were carried out on IR Affinity (FTIR spectrometer) from Shimadzu, the sample prepared on a glass plate contain solid KBr. Liquid chromatography–mass spectrometry (LC/MS) were recorded on spectroscopy A LCMS-8040 Shimadzu corporation, model CAT-30A, Serial no. L20574900241 AE, 220-240v ~ 50/60 HZ 300VA. ¹H NMR and ¹³CNMR were recorded on BRUKER spectrometer, 400 MHZ. The samples were prepared by dissolution at DMSO *d*₆. Chemical shifts (δ) are presented in part per million (ppm)

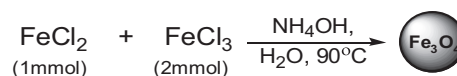
using tetramethylsilane (TMS) an internal standard. Elemental microanalysis were done on Carlo Erba analyzer model 110. X-ray diffraction (XRD) patterns were recorded on a Rigaku model Ultima-IV diffractometer employing Cu-K α radiation ($\lambda = 1.5406 \text{ \AA}$) at 40 kV and 25 mA over a 2θ range between 20 and 80°. All XRD measurement is handled in the air atmosphere. Samples for Scanning electron microscope (SEM) were prepared from ethanolic suspensions on single-sided alumina tape placed on alumina stubs. For the elemental analysis and mapping, the energy-dispersive X-ray spectra (EDS) were collected on a Lyra 3 (Tescan from the Czech Republic) attachment to the SEM. TEM micrographs were obtained from a high-resolution transmission electron microscopy (HRTEM) (JEOL JEM-2100F) equipped with an energy dispersive X-ray spectrometer (EDX) operated at 200 kV. 300 mesh copper grids coated with carbon films were used for the imaging. The transmission electron microscopy (TEM) samples were prepared by dropping it on a copper grid from an ethanolic suspension and drying at room temperature.

2.2. Synthesis of nanomagnetic catalyst

Magnetic nanoparticles with the range of 6–8 nm were prepared as the literature procedure (Nezhad and Mohammadi, 2015). Scheme 1 shows the synthetic route for the magnetic nanoparticles. In a 250 ml, round bottom flask, hydrated ferrous chloride (FeCl₂·H₂O) (5 mmol, 1 g) and ferric chloride (FeCl₃·H₂O) (10 mmol, 4.04 g) were dissolved in de-ionized water (DI-H₂O) (100 ml) under nitrogen atmosphere with a continuous stirring speed of 600 rpm. Then NH₄OH (25%) solution (25 ml) was added slowly at 90 °C to raise the pH at 9. The orange color solution was turned black in 15 min and continued stirring for another 3 h to complete the reduction. The mixture was allowed to precipitate and collected using a simple magnet. The black solid was washed several times (5 × 25 ml) to remove the unreacted metal precursors and ammonia. The powdered material was dried and used for characterization and catalytic reaction.

2.3. Synthesis of 2-(chlorodiazenyl)thiazole (2)

Dissolve (1 mmol) of 2-aminothiazole in 3 ml of HCl keep this solution in ice at 0°C diazotize this solution by using NaNO₂ (5 mmol) solution (prepared by dissolving 5 mmol of NaNO₂ in 1 ml H₂O). The reaction mixture was kept onto ice bath for 3 h to give compound (2) as pink powder; yield (89.78%), mp.: 126.7 °C. FT-IR (KBr, ν , cm⁻¹): absence of (NH₂), 3010, appearance of 2980 (CH), and 1530 (N=N); ¹H NMR: (400 MHz, DMSO *d*₆, δ , ppm), 6.90 (d, 1H, *J* = 8.0 Hz, H-2-thiazole), 7.27 (d, 1H, *J* = 8.0 Hz, H-3-thiazole); ¹³C NMR: (400 MHz, DMSO *d*₆, δ , ppm): 170.1 (S-C-N), 150.4 (C=N thiazolyl), 127.0, 108.3 (C-thiazolyl); MS (*m/z*): 146.98 (M⁺, 80.5%); Anal. calcd. for C₃H₂ClN₃S (146.96): C, 24.41; H, 1.37; Cl, 24.02; N, 28.47; S, 21.73%; found: C, 24.93; H, 1.39; Cl, 24.00; N, 28.45; S, 21.77%.



Scheme 1 Synthesis of Fe₃O₄.

2.4. Coupling with active methylene compounds (3-5)

Reflux an equivalent mixture of (2) (1 mmol) and/or (1 mmol) ethyl acetoacetate, ethyl cyanoacetate, and acetylacetone respectively in presence of 25 ml ethanol for 1 h. The solid formed was recrystallized from ethanol to give (3-5) respectively.

2.4.1. Ethyl 3-oxo-2-(thiazol-2-yl-diazanyl)butanoate (3)

Light orange powder; yield (60.78%), mp.: 135.4 °C; FT-IR (KBr, ν , cm^{-1}): 3010, 2980 (CH), 1750 (C=O ester and ketone) and 1590 (C=N); ^1H NMR: (400 MHz, DMSO d_6 , δ , ppm): 1.35, 2.08 (m, 3H, 2CH₃), 4.11 (m, 2H, -CH₂-), 4.70 (s, 1H, CH=N); 7.16 (d, 1H, J = 8.0 Hz, H-2-thiazole), 7.55 (d, 1H, J = 8.0 Hz, H-3-thiazole), ^{13}C NMR: (400 MHz, DMSO d_6 , δ , ppm): 171.0 (C=O ester), 166.9 (C=O), 150.4 (C=N thiazolyl), 125.0, 119.8 (C-thiazolyl), 62.9 (C-N), 13.6, 20.0 (CH₃); 59.2 (CH₂); MS (m/z): 240.05 ($\text{M}^+ - 1$, 11.33%); Anal. calcd. for C₉H₁₁N₃O₃S (241.05): C, 44.80; H, 4.60; N, 17.42; S, 13.29%; found: C, 43.93; H, 4.44; N, 17.45; S, 13.39%.

2.4.2. Ethyl 2-cyano-2-(thiazol-2-yl-diazanyl)acetate (4)

Orange powder; yield (65.45%), mp.: 144.3 °C; FT-IR (KBr, ν , cm^{-1}): 3010, 2980 (CH), 1749 (C=O), 1590 (C=N), 2260-2222 (CN); ^1H NMR: (400 MHz, DMSO d_6 , δ , ppm): 1.65 (s, 3H, CH₃), 2.12 (m, 3H, -CH₂-), 7.15 (d, 1H, J = 8.0 Hz, H-2-thiazole), 7.5 (d, 1H, J = 8.0 Hz, H-3-thiazole); ^{13}C NMR: (400 MHz, DMSO d_6 , δ , ppm): 171.1 (C=O), 152.4 (C=N thiazolyl), 125.0, 119.7 (CH-thiazolyl), 114.9 (CN), 50.9 (C-N), 58.7 (-CH₂), 22.9 (CH₃); MS (m/z): 224.04 (M^+ , 11.69%); Anal. calcd. for C₈H₈N₄O₂S (224.04): C, 42.85; H, 3.60; N, 24.99; S, 14.30%; found: C, 42.55; H, 3.68; N, 25.10; S, 14.40%.

2.4.3. 3-(Thiazol-2-yl-hydrazono)-pentane-2,4-dione (5)

Yellow powder; yield (60.02%), mp.: 165.2 °C; FT-IR (KBr, ν , cm^{-1}): 3010, 2980 (CH), 1750 (C=O) and 1590 (C=N); ^1H NMR: (400 MHz, DMSO d_6 , δ , ppm): 2.09 (s, 6H, 2CH₃), 7.15 (d, 1H, J = 8.0 Hz, H-2-thiazole), 7.51 (d, 1H, J = 8.0 Hz, H-3-thiazole); ^{13}C NMR: (400 MHz, DMSO d_6 , δ , ppm): 206.0 (C=O), 85.7 (C=N), 151.4 (C=N thiazolyl), 125.0, 119.7 (C-thiazolyl), 19.3 (CH₃); MS (m/z): 211.24 (M^+ , 22.01%); Anal. calcd. for C₈H₉N₃O₂S (211.24): C, 45.49; H, 4.29; N, 19.89; O, 15.15; S, 15.18%; found: C, 46.10; H, 4.32; N, 19.69; O, 15.05; S, 15.08%.

2.5. Synthesis of pyrazol-5-one derivatives (6-8)

An equivalent mixture of (1 mmol) of (3-5) and (2 mmol) hydrazine hydrate in presence of (0.01 mmol) of Fe₃O₄ the reaction mixture was stirred for 2 h at room temperature. The solid formed was recrystallized from ethanol to give (6-8) respectively.

2.5.1. 3-methyl-4-(thiazol-2-yl-diazanyl)-1H-pyrazol-5(4H)-one (6)

Beige powder; yield (87.02%), mp.: 155.5 °C; FT-IR (KBr, ν , cm^{-1}): 3250 (NH), 3010, 2980 (CH), 1750 (C=O) and 1590 (C=N); ^1H NMR: (400 MHz, DMSO d_6 , δ , ppm): 1.75

(s, 3H, CH₃), 7.16 (d, 1H, J = 8.0 Hz, H-2-thiazole), 7.55 (d, 1H, J = 8.0 Hz, H-3-thiazole), 8.70 (s, 1H, NH-pyrazolyl); ^{13}C NMR: (400 MHz, DMSO d_6 , δ , ppm): 166.9 (C=O), 156.7 (C=N pyrazolyl), 150.4 (C=N thiazolyl), 125.0, 119.7 (C-thiazolyl), 62.9 (C-N), 22.9 (CH₃); MS (m/z): 209.06 (M^+ , 55.05%); Anal. calcd. for C₇H₇N₅OS (209.04): C, 40.18; H, 3.37; N, 33.47; S, 15.33%; found: C, 41.00; H, 3.26; N, 33.27; S, 15.13%.

2.5.2. 3-amino-4-(thiazol-2-yl-diazanyl)-1H-pyrazol-5(4H)-one (7)

Orange powder; yield (80.66%), mp. 172.0 °C; FT-IR (KBr, ν , cm^{-1}): 3330, 3250 (NH₂, NH), 3010, 2850 (CH), 1715 (C=O) and 1600 (C=N); ^1H NMR: (400 MHz, DMSO d_6 , δ , ppm): 1.81 (s, 3H, CH₃), 7.69 (d, 1H, J = 8.0 Hz, H-2-thiazole), 7.88 (d, 1H, J = 8.0 Hz, H-3-thiazole), 8.26 (s, 1H, NH-pyrazolyl), 8.54 (s, 1H, NH₂); ^{13}C NMR: (400 MHz, DMSO d_6 , δ , ppm): 163.3 (C=O), 159.2 (C=N pyrazolyl), 140.9 (C=N pyrazolyl), 140.0 (C-thiazolyl), 62.9 (C-N), 15.4 (CH₃); MS (m/z): 210.01 (M^+ , 33.2%); Anal. calcd. for C₆H₆N₆OS (210.03): C, 34.28; H, 2.88; N, 39.98; S, 15.25%; found: C, 33.20; H, 2.90; N, 40.00; S, 15.35%.

2.5.3. (3,5-Dimethyl-4H-pyrazol-4-yl)-thiazol-2-yl-diazene (8)

Yellow powder; yield (82.03%), mp.: 186.0 °C; FT-IR (KBr, ν , cm^{-1}): 3380 (NH), 3010, 2850 (CH, CH₃), and 1600 (C=N); ^1H NMR: (400 MHz, DMSO d_6 , δ , ppm): 2.01 (s, 6H, 2CH₃), 7.46 (d, 1H, J = 8.0 Hz, H-2-thiazole), 8.00 (d, 1H, J = 8.0 Hz, H-3-thiazole), 8.00 (s, 1H, NH-pyrazolyl). ^{13}C NMR: (400 MHz, DMSO d_6 , δ , ppm): 164.5 (C=N pyrazolyl), 33.4 (C=N pyrazolyl), 153.0, 143.0 (C-thiazolyl), 62.9 (C-N), 15.4 (CH₃); MS (m/z): 207.10 (M^+ , 22.11%); Anal. calcd. for C₈H₉N₅S (207.06): C, 46.36; H, 4.38; N, 33.79; S, 15.47%; found: C, 45.49; H, 4.40; N, 33.81; S, 15.45%.

2.6. General procedure for Schiff bases synthesis of compounds (9-18)

An equivalent mixture of (1 mmol) of 1 and various aromatic aldehydes (1 mmol) namely: benzaldehyde, *p*-chlorobenzaldehyde, *o*-nitrobenzaldehyde, *p*-hydroxybenzaldehyde, salicylaldehyde, *p/o*-anisaldehyde, 2,4-/2,3-dimethoxybenzaldehyde, and 2-methyl indolyl-3-carboxaldehyde in presence of (0.01 mmol) of Fe₃O₄ the reaction medium at room temperature (RT) for 1 h. The solid formed was recrystallized from ethanol to give the corresponding Schiff bases derivatives (9-18).

2.6.1. *N*-(benzylidene)thiazol-2-amine (9)

Yellow powder; yield (81.00%), mp.: 113-115 °C; FT-IR (KBr, ν , cm^{-1}): absence of (C=O, NH₂), 3020, 2990 (CH), 1590 (C=N); ^1H NMR: (400 MHz, DMSO d_6 , δ , ppm): 7.37-7.74 (m, 5H, CH-aromatic), 7.54 (d, 1H, J = 8.0 Hz, H-4-thiazole), 7.20 (d, 1H, J = 8.0 Hz, H-5-thiazole), 9.01 (s, 1H, CH=N); ^{13}C NMR: (400 MHz, DMSO d_6 , δ , ppm): 171.9 (N=C-S), 167.1 (HC=N), 141.6, 119.8 (C-thiazolyl), 127.8, 124.4, 123.9 (C-aromatic); MS(m/z): 188.10 (M^+ , 44.05%); Anal. calcd. for C₁₀H₈N₂S (188.04): C, 63.80; H, 4.28; N, 14.88; S, 17.03%; found: C, 63.76; H, 4.20; N, 14.91; S, 17.13%.

2.6.2. *N*-(4-chlorobenzylidene)thiazol-2-amine (10)

Brown crystal; yield (88.05%), mp.: 134.8 °C; FT-IR (KBr, ν , cm^{-1}): absence of (C=O, NH₂), 3010, 2990 (CH), 1580 (C=N); ¹H NMR: (400 MHz, DMSO *d*₆, δ , ppm): 7.66–8.02 (m, 4H, CH-aromatic), 7.60 (d, 1H, *J* = 8.0 Hz, H-4-thiazole), 6.65 (d, 1H, *J* = 8.0 Hz, H-5-thiazole), 8.03 (s, 1H, CH=N); ¹³C NMR: (400 MHz, DMSO *d*₆, δ , ppm): 171.9 (N=C–S), 166.6 (HC=N), 141.7, 119.9 (C-thiazolyl), 131.2, 129.7, 128.8 (C-aromatic); MS (*m/z*): 222.00 (M⁺, 18.52%); Anal. calcd. for C₁₀H₇ClN₂S (222.00): C, 53.93; H, 3.17; Cl, 15.92; N, 12.58; S, 14.40%; found: C, 53.83; H, 3.20; Cl, 15.99; N, 12.50; S, 14.51%.

2.6.3. *N*-(2-nitrobenzylidene)thiazol-2-amine (11)

Orange powder; yield (80.97%), mp.: 163.5 °C; FT-IR (KBr, ν , cm^{-1}): absence of (C=O, NH₂), 3020, 2990 (CH), 1610 (C=N); ¹H NMR: (400 MHz, DMSO *d*₆, δ , ppm): 6.54–7.76 (m, 4H, CH-aromatic), 7.02 (d, 1H, *J* = 8.0 Hz, H-4-thiazole), 7.01 (d, 1H, *J* = 8.0 Hz, H-5-thiazole), 8.51 (s, 1H, CH=N); ¹³C NMR: (400 MHz, DMSO *d*₆, δ , ppm): 166.8 (N=C–S), 148.6 (HC=N), 134.4, 110.7 (C-thiazolyl), 134.2, 130.5, 129.0, 124.4 (C-aromatic); MS(*m/z*): 233.02 (M⁺, 80%); Anal. calcd. for C₁₀H₇N₃O₂S (233.03): C, 51.49; H, 3.02; N, 18.02; S, 13.75%; found: C, 51.55; H, 3.32; N, 18.12; S, 13.75%.

2.6.4. 4-((thiazol-2-ylimino)methyl)phenol (12)

Yellow powder; yield (85.43%), mp.: 175.2 °C; FT-IR (KBr, ν , cm^{-1}): absence of (C=O, NH₂), 3020, 2890 (CH), 1605 (C=N); ¹H NMR: (400 MHz, DMSO *d*₆, δ , ppm): 6.38–7.33 (m, 4H, CH-aromatic), 7.07 (d, 1H, *J* = 8.0 Hz, H-4-thiazole), 7.72 (d, 1H, *J* = 8.0 Hz, H-5-thiazole), 8.27 (s, 1H, CH=N), 9.84 (s, 1H, OH); ¹³C NMR: (400 MHz, DMSO *d*₆, δ , ppm): 173.0 (N=C–S), 162.2 (HC=N), 141.3, 115.9 (C-thiazolyl), 138.6, 132.1, 116.1 (C-aromatic); MS (*m/z*): 206.05 (M⁺ + 2, 23.07%); Anal. calcd. for C₁₀H₈N₂OS (204.04): C, 58.80; H, 3.95; N, 13.72; S, 15.70%; found: C, 58.92; H, 3.98; N, 13.75; S, 15.75%.

2.6.5. 2-((thiazol-2-ylimino)methyl)phenol (13)

Brown powder; yield (90.00%), mp.: 165.8 °C; FT-IR (KBr, ν , cm^{-1}): absence of (C=O, NH₂), 3320 (OH), 3010, 2880 (CH), 1610 (C=N); ¹H NMR: (400 MHz, DMSO *d*₆, δ , ppm): 10.25 (s, 1H, OH), 6.92–8.54 (m, 4H, CH-aromatic), 7.64 (d, 1H, *J* = 8.0 Hz, H-4-thiazole), 7.01 (d, 1H, *J* = 8.0 Hz, H-5-thiazole), 8.21 (s, 1H, CH=N); ¹³C NMR: (400 MHz, DMSO *d*₆, δ , ppm): 160.8 (N=C–S), 154.3 (HC=N), 122.3, 119.5, 119.5 (C-aromatic); MS (*m/z*): 204.04 (M⁺, 55.01%); Anal. calcd. for C₁₀H₈N₂OS (204.04): C, 58.80; H, 3.95; N, 13.72; S, 15.70%; found: C, 58.78; H, 3.75; N, 13.62; S, 15.68%.

2.6.6. *N*-(4-methoxybenzylidene)thiazol-2-amine (14)

Yellow powder; yield (87.07%), mp.: 102–103 °C; FT-IR (KBr, ν , cm^{-1}): absence of (C=O, NH₂), 3020, 2990 (CH), 1610 (C=N); ¹H NMR: (400 MHz, DMSO *d*₆, δ , ppm): 6.35–7.31 (m, 4H, CH-aromatic), 7.60 (d, 1H, *J* = 8.0 Hz, H-4-thiazole), 7.21 (d, 1H, *J* = 8.0 Hz, H-5-thiazole), 8.12 (s, 1H, CH=N), 3.38 (s, 6H, OCH₃); ¹³C NMR: (400 MHz,

DMSO *d*₆, δ , ppm): 161.5 (N=C–S), 156.0 (HC=N), 135.5, 120.6 (C-thiazolyl), 129.5, 128.4, 126.1 (C-aromatic), 62.9, 55.8 (OCH₃); MS (*m/z*): 218.10 (M⁺, 60.0%); Anal. calcd. for C₁₁H₁₀N₂OS (218.05): C, 60.53; H, 4.62; N, 12.83; S, 14.69%; found: C, 60.71; H, 4.66; N, 12.88; S, 14.72%.

2.6.7. *N*-(2-methoxybenzylidene)thiazol-2-amine (15)

Orange powder; yield (90.05%), mp.: 150.9 °C; FT-IR (KBr, ν , cm^{-1}): absence of (C=O, NH₂), 3009, 2890 (CH), 1605 (C=N); ¹H NMR: (400 MHz, DMSO *d*₆, δ , ppm): 6.37–7.21 (m, 4H, CH-aromatic), 7.60 (d, 1H, *J* = 8.0 Hz, H-4-thiazole), 7.21 (d, 1H, *J* = 8.0 Hz, H-5-thiazole), 8.12 (s, 1H, CH=N), 3.33 (s, 6H, OCH₃); ¹³C NMR: (400 MHz, DMSO *d*₆, δ , ppm): 161.5 (N=C–S), 156.0 (HC=N), 135.5, 120.6 (C-thiazolyl), 129.5, 128.5, 126.1 (C-aromatic), 62.9, 55.8 (OCH₃); MS (*m/z*): 218.10 (M⁺, 66.03%); Anal. calcd. for C₁₁H₁₀N₂OS (218.05): C, 60.53; H, 4.62; N, 12.83; S, 14.69%; found: C, 60.73; H, 4.74; N, 11.93; S, 13.99%.

2.6.8. *N*-(2,4-dimethoxy-benzylidene)-thiazol-2-yl-amine (16)

Orange powder; yield (87.50%), mp.: 135.2 °C; FT-IR (KBr, ν , cm^{-1}): absence of (C=O, NH₂), 3020, 2990 (CH), 1590 (C=N); ¹H NMR: (400 MHz, DMSO *d*₆, δ , ppm): 7.37–7.67 (m, 3H, CH-aromatic), 7.60 (d, 1H, *J* = 8.0 Hz, H-4-thiazole), 7.20 (d, 1H, *J* = 8.0 Hz, H-5-thiazole), 9.01 (s, 1H, CH=N), 3.49 (s, 6H, 2CH₃); ¹³C NMR: (400 MHz, DMSO *d*₆, δ , ppm): 173.6 (N=C–S), 167.1 (HC=N), 141.7, 119.7 (C-thiazolyl), 126.9, 124.5, 123.7 (C-aromatic), 62.8, 55.8 (OCH₃); MS (*m/z*): 188.10 (M⁺, 88.01%); Anal. calcd. for C₁₂H₁₂N₂O₂S (248.06): C, 58.05; H, 4.87; N, 11.28; S, 12.91%; found: C, 59.04; H, 4.86; N, 11.29; S, 12.90%.

2.6.9. *N*-(2,3-dimethoxybenzylidene)thiazol-2-amine (17)

Yellow powder; yield (89.09%), mp.: 152.0 °C; FT-IR (KBr, ν , cm^{-1}): absence of (C=O, NH₂), 3020, 2990 (CH), 1609 (C=N); ¹H NMR: (400 MHz, DMSO *d*₆, δ , ppm): 7.37–7.64 (m, 3H, CH-aromatic), 7.64 (d, 1H, *J* = 8.0 Hz, H-4-thiazole), 7.20 (d, 1H, *J* = 8.0 Hz, H-5-thiazole), 9.29 (s, 1H, CH=N), 3.44 (s, 6H, 2CH₃); ¹³C NMR: (400 MHz, DMSO *d*₆, δ , ppm): 172.6 (N=C–S), 167.1 (HC=N), 141.6, 119.8 (C-thiazolyl), 127.9, 124.5, 123.9 (C-aromatic), 62.8, 55.8 (OCH₃); MS (*m/z*): 248.10 (M⁺, 100%); Anal. calcd. for C₁₂H₁₂N₂O₂S (248.06): C, 58.05; H, 4.87; N, 11.28; S, 12.91%; found: C, 59.05; H, 4.89; N, 11.30; S, 12.71%.

2.6.10. *N*-((2-methyl-1H-indol-3-yl)methylene)thiazol-2-amine (18)

Yellow powder; yield (86.34%), mp.: 180.2 °C; FT-IR (KBr, ν , cm^{-1}): absence of (C=O, NH₂), 3010, 2980 (CH), 1600 (C=N); ¹H NMR: (400 MHz, DMSO *d*₆, δ , ppm): 11.99 (s, 1H, NH-indolyl), 7.13–8.02 (m, 4H, CH-aromatic), 7.16 (d, 1H, *J* = 8.0 Hz, H-4-thiazole), 7.14 (d, 1H, *J* = 8.0 Hz, H-5-thiazole), 8.04 (s, 1H, CH=N), 2.67 (s, 3H, CH₃); ¹³C NMR: (400 MHz, DMSO *d*₆, δ , ppm): 184.1 (N=C–S), 148.5, 111.4 (C-thiazolyl), 150.0 (HC=N), 135.3, 125.6, 122.6, 121.8, 119.9, 113.6 (C-aromatic); MS (*m/z*): 241.10 (M⁺, 66.09%); Anal. calcd. for C₁₃H₁₁N₃S (241.07): C, 64.70; H, 4.59; N, 17.41; S, 13.29%; found: C, 64.55; H, 4.48; N, 17.38; S, 13.31%.

3. Results and discussion

Ultrasmall nanomagnetic catalysts are prepared by the coprecipitation technique using Fe(II) and Fe(III) precursors using NH_4OH as a reductant. The black powdered materials are collected by a magnet and non-magnetic materials are removed by washing with water repeatedly [Scheme 1](#). The crystallinity of the prepared nanocatalyst is confirmed by the XRD signature. The XRD patterns of Fe_3O_4 display characteristic peaks at 2θ 30.2, 35.7, 43.1, 53.4, 57.1, and 63.2 indicate the formation of crystalline cubic (Fd3m) spinel structure (JCPDS card no. 01-075-0449) ([Shaikh et al., 2018](#); [Pinna et al., 2005](#)). Scanning electron microscopic (SEM) images demonstrated highly agglomerated particles with extremely smaller-sized particles with smooth surfaces. The higher surface interaction among the bared-surface ultrasmall nanoparticle leads to aggregation of the particles. Transmission electron microscopic (TEM) images show spherically shaped, uniformly distributed, nano-sized particles with 6–8 nm diameter, [Fig. 1](#). The magnetic nature of the Fe_3O_4 NPs have been investigated and the results are shown below. The magnetization field (M–H) curve is recorded at room temperature using PMC Micromag 3900 model vibrating sample magnetometer (VSM) equipped with a 1 Tesla magnet. The magnetic measurement of the Fe_3O_4 NPs shows 62.69 emu g^{-1} , which is magnetic enough to separate the nanoparticles using a simple magnet at the bottom of the vessel after the reaction, [Fig. 2](#).

Using nanomagnetic catalysis is a modern technique for preparing organic compounds with high yield in a short time. Previously, we reported the diazotization of amino heterocyclic compounds followed by coupling with various active methylene compounds afford the corresponding coupling products by traditional method in ethanol under reflux for 2 h ([Abrams, 2021](#)). Herein, by using nanomagnetic catalysis (Fe_3O_4 NPs) in coupling the diazonium salt of 2-

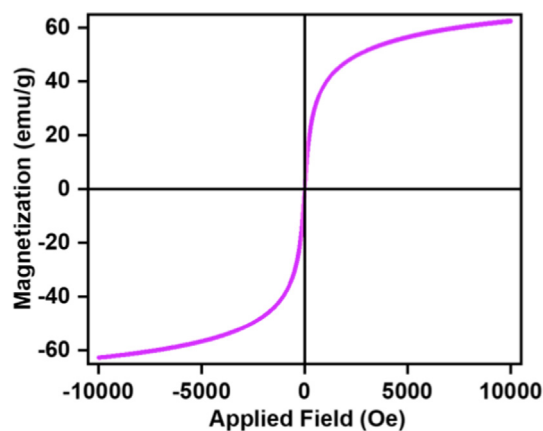


Fig. 2 Magnetic hysteresis loops of Fe_3O_4 at room temperature with 1 Tesla magnet.

aminothiazole with various active methylene compounds, it seems the reaction is completed in 1 h. In addition to the short time reaction, it affords high yields compared to the conventional method [Fig. 3](#) and [Fig. 4](#) represents the pathway of preparing the target compounds. Coupling diazonium salt of 2-aminothiazole with ethyl acetoacetate, ethyl cyanoacetate, and acetylacetone in presence of a catalytic amount of Fe_3O_4 NPs at room temperature afforded the corresponding coupling products (**3–5**) respectively in 1 h. The reaction was monitored using the TLC technique. The prepared compounds were proved via their cyclization upon nucleophilic attack within by nucleophile hydrazine hydrate afforded the corresponding pyrazolyl derivatives (**6–8**) [Scheme 2](#), these novel products were elucidated based on spectral data. The IR spectrum of (**6**) displayed absorption bands in the region 1750 cm^{-1} due to $\text{C}=\text{O}$, and at 3330, 3250 attributable for NH_2 in (**7**). ^1H NMR spectrum of (**7**) as an example, recorded new signals

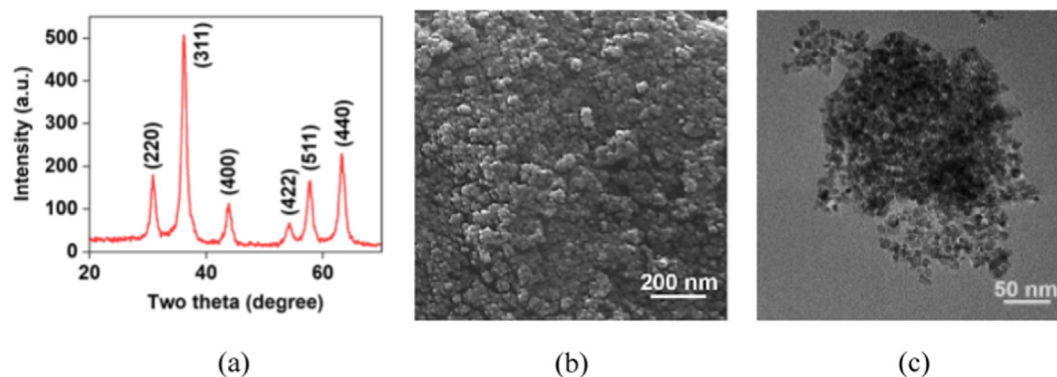


Fig. 1 X-ray diffraction(a) pattern of the Fe_3O_4 NPs, the SEM (b) and TEM (c) images show spherically shaped, uniformly distributed, nano-sized particles with 6–8 nm.

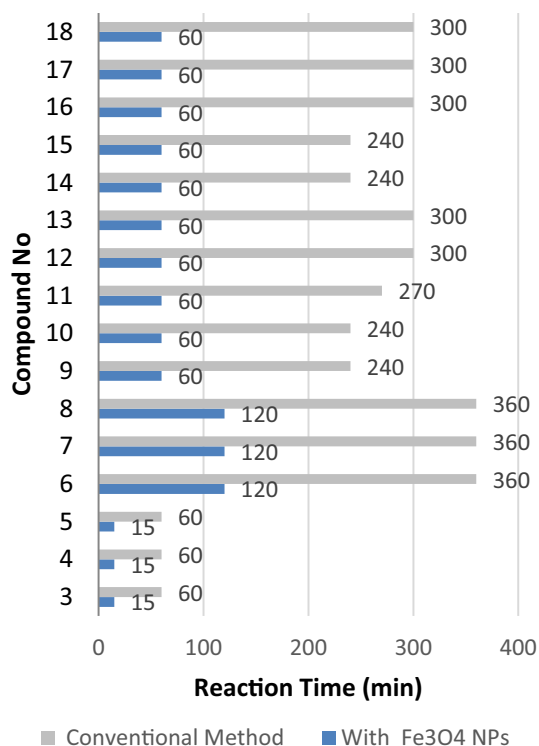


Fig. 3 Reaction time (min) of products by used conventional method and with Fe₃O₄ NPs catalysis.

at δ 8.54, 8.26 ppm assigned for NH₂ and NH, respectively. More details are required such as the absence of CN in 4 and more discussion for other products.

Moreover, the formation of Schiff bases from 2-aminothiazole was carried out upon one-pot method by treatment with a various aromatic aldehyde in presence of Fe₃O₄ NPs catalysis at room temperature [Scheme 3](#)., The reaction vanished in less time, and furnished high yield than the conventional method ([Fig. 3](#) and [Fig. 4](#)).

The structures of these products were assigned using spectral analysis. IR for all compounds showed the absence of C=O and NH₂.

The plausible mechanism is consistent with the literature ([Sadeghi et al., 2016](#); [Zolfigol et al., 2012](#); [Safari and Sadeghi, 2017](#)). Preparation of intermediate given on proposed mechanism for the production of pyrazole derivatives in [Scheme 4](#). Fe₃O₄ NPs as the catalyst can activate the active methylene compounds (A) through coordination to the oxygen atom of carbonyl group the catalyst can participate in the conversion of (B), Afterwards, Fe₃O₄ NPs promoted cyclization with hydrazine hydrate and dehydration gives pyrazole derivatives (6) ([Polshettiwar and Varma, 2010](#)). The nanocatalyst could be magnetically recovered from the reaction mixture during the workup procedure. Furthermore, the proposed mechanism for preparation of the Schiff base derivatives were

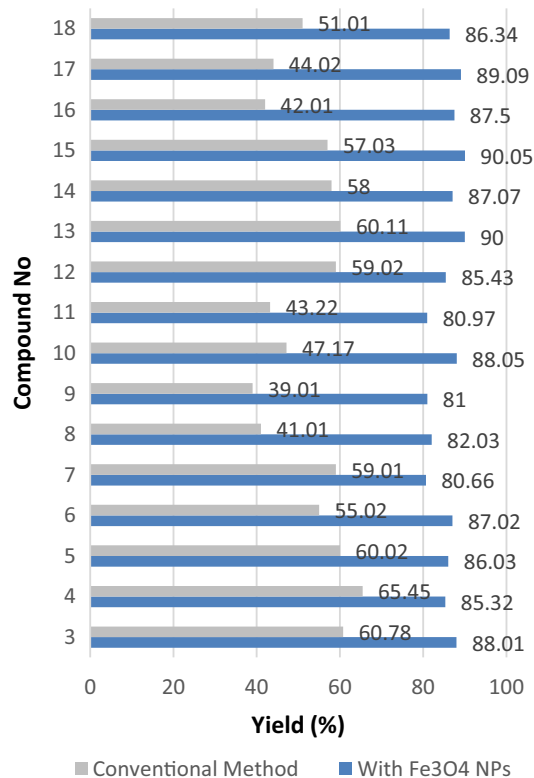
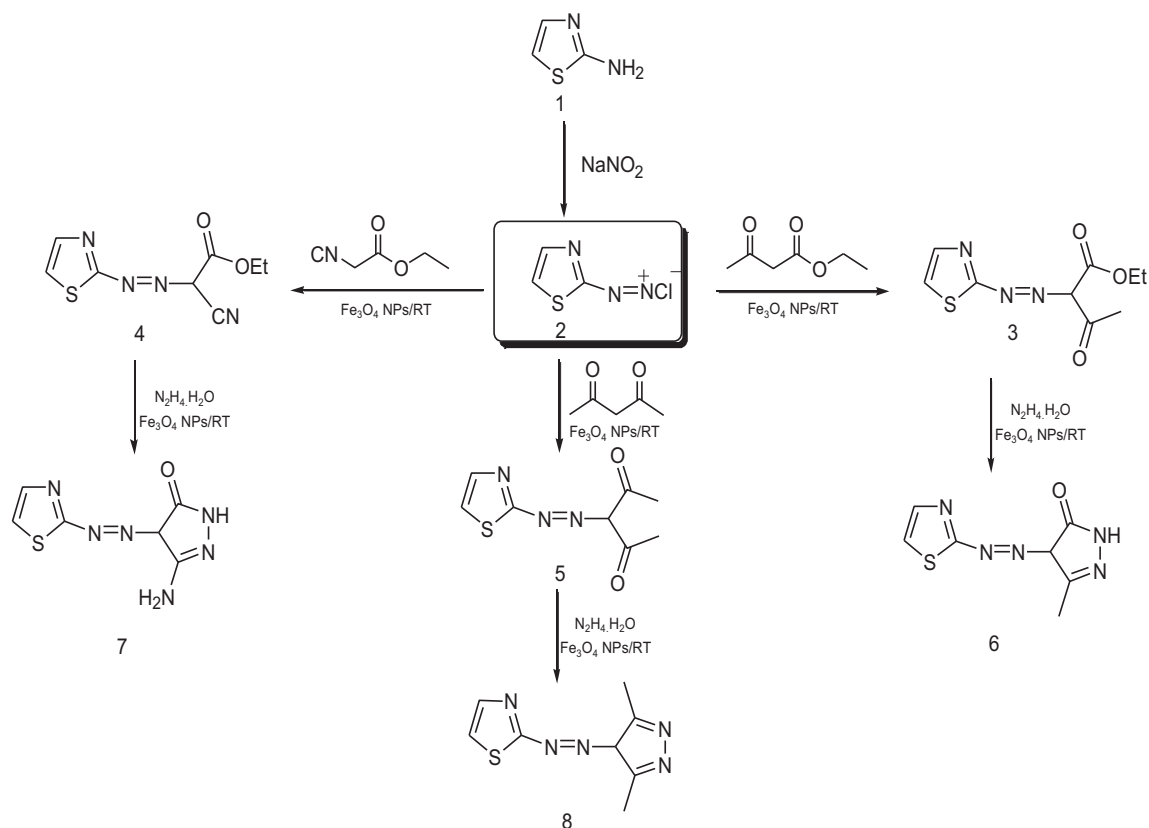


Fig. 4 The effect of the yield (%) of products by used conventional methods and with Fe₃O₄ NPs catalysis.

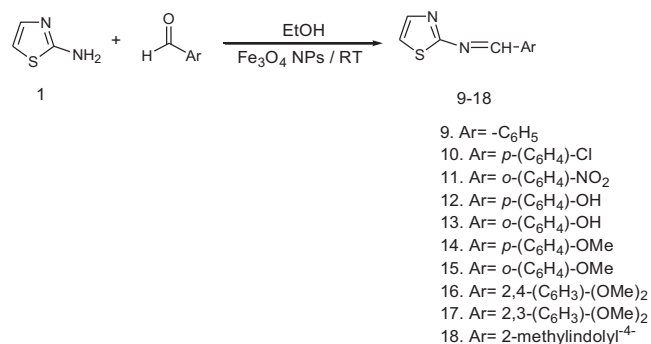
carried out by activated aromatic aldehyde by Fe(III) of Fe₃O₄ NPs. Then nucleophilic attack of the amino group to carbonyl group of the activated aldehyde via the elimination of H₂O to give Schiff base derivatives.

4. Biological activity

All of the synthesized compounds were tested for their antimicrobial activities against five types of microorganisms including gram-negative and positive bacteria as well as fungus. The gram-negative bacteria include *E. coli* and *P. aeruginosa* while *B. subtilis* and *S. aureus* represented the gram-positive bacteria. *C. albicans* was the fungus used in the screening. Generally, the tested compounds exhibited better antibacterial activities with *E.coli* being the most sensitive bacteria. However, all of the tested compounds, except compound (7), showed no antifungal activity against *C. albicans*. Azo derivatives (1, 2) and (6–8) were slightly more potent than the Schiff bases with average zones of inhibition of 15.71 and 12.87 mm, respectively. Compound (6) showed the most potent antibacterial activity against *E. coli* whereas compound (9) was the least potent among the tested series of compounds. Unsubstituted benzene in Schiff base (9) was less potent than the substituted analogues. This may suggest that benzene substitution is beneficial for the activity. In addition, the antibacterial activity of

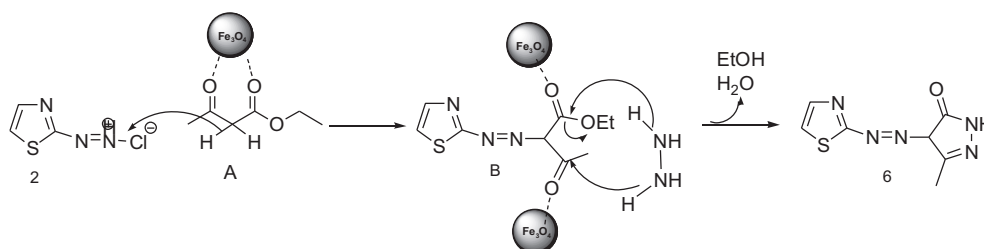


Scheme 2 Synthesis pathway for pyrazole derivatives.



Scheme 3 Synthesis pathway for Schiff bases derivatives.

the Schiff bases seems to be insensitive to the position and the nature of the substituents. The majority of the synthesized compounds did not exhibit antipseudomonal activity. Only compounds (**6**, **9**, **11**, **13**) and (**18**) showed growth inhibition against *P. aeruginosa* with the most potent activity observed with compound (**6**). However, none of the compounds was superior to piperacillin. For the gram-positive bacteria, the antibacterial activity was more significant in Schiff base derivatives when compared to azo derivatives. None of the tested compounds showed superior activity to the positive control. Compound (**7**) was the only pyrazole derivative that showed activity against both *B. subtilis* and *S. aureus* whereas, compounds (**11**, **13**) and (**16**) were the only Schiff bases that exhibited activities against both bacteria [Table 1](#).



Scheme 4 Plausible mechanism for synthesis pyrazole derivative by using Fe₃O₄ NPs.

Table 1 Anti-microbial activity of 2-aminothiazoles derivatives (6–18).

Compd. No	Inhibition Zone (mm \pm SD)				
	<i>E. coli</i>	<i>P. aeruginosa</i>	<i>Bacillus</i>	<i>S. aureus</i>	<i>Candida</i>
(1)	9.8 \pm 0.2	Nil	Nil	Nil	Nil
(2)	10.15 \pm 0.4	Nil	Nil	Nil	Nil
(6)	22 \pm 0.8	16	Nil	Nil	Nil
(7)	16 \pm 1.4	Nil	8.6 \pm 0.4	12.3 \pm 0.9	12 \pm 0
(8)	20.6 \pm 0.9	Nil	10.6 \pm 0.4	Nil	Nil
(9)	8.8 \pm 0.4	8 \pm 0	Nil	Nil	Nil
(10)	14.3 \pm 0.4	Nil	Nil	Nil	Nil
(11)	12.3 \pm 0.9	11.8 \pm 0.6	8.6 \pm 0.9	10.6 \pm 0.4	Nil
(12)	13.3 \pm 1.2	Nil	Nil	10.3 \pm 0.7	Nil
(13)	14.95 \pm 0.5	9 \pm 0.8	11.3 \pm 0.4	9.6 \pm 0.3	Nil
(14)	12.3 \pm 0.4	Nil	10.3 \pm 0.4	Nil	Nil
(15)	12.6 \pm 0.5	Nil	Nil	Nil	Nil
(16)	13 \pm 0.4	Nil	12.3 \pm 0.4	13 \pm 0.1	Nil
(17)	10.95 \pm 0.3	Nil	Nil	9.3 \pm 0.4	Nil
(18)	16.2 \pm 0.9	14.6 \pm 0.7	10 \pm 0.2	11.3 \pm 0.4	Nil

Concentration of Piperacillin = 4 μ g/ml; Cefotaxime = 1 μ g/ml; Oxacillin = 2 μ g/ml; Fluconazole = 0.5 mg/ml.

5. Conclusion

In summary, we have demonstrated an efficient, simple, efficient, and stable catalyst-Fe for the synthesis pyrazolyl derivatives and 2-aminothiazole Schiff bases derivatives of 2-aminothiazole by comparing showed best results of Fe₃O₄ NPs nanomagnetic catalysis and convention method. This method is quick, and avoids the use of toxic or heavy metals, high temperature, improved product yields, and easy workup procedure.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

We would like to thank Deanship of Scientific Research at Imam Abdulrahman Bin Faisal University for financial support under projects no: (2019-132-sci), thanks for the co-workers and trenching at Basic and Applied Scientific Research Center, Imam Abdulrahman Bin Faisal University, for making the spectral analyses, and for department of pharmaceutical chemistry and chemistry department at King Saud University to make the biological activity and nmr analysis for this research.

Appendix A. Supplementary material

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.arabjc.2022.103878>.

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