

# King Saud University

# Arabian Journal of Chemistry

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# **ORIGINAL ARTICLE**

# Fabrication of a novel electrochemical sensor based on tin disulfide/multi-walled carbon nanotunbesmodified electrode for rutin determination in natural vegetation



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Received 11 April 2022; accepted 18 January 2023 Available online 26 January 2023

# KEYWORDS

Tin disulfide; Multi-walled carbon nanotubes; Electrochemical sensor; Rutin **Abstract** A tin disulfide and multi-walled carbon nanotube  $(SnS_2/CNTs)$  electrochemical sensor was constructed for the sensitive and selective determination of rutin in plants. Tin disulfide nanoflowers with various particle sizes were prepared by controlling the reaction time and composited with multi-walled CNTs. The morphology, crystal structure, and chemical composition of these  $SnS_2/CNTs$  composites were characterized using XRD, XPS, and SEM-EDS. Results illustrated that the  $SnS_2/CNTs$  had a large specific surface area, good conductivity, and remarkable electrocatalytic performance. The pH of the buffer solution, the scanning rate, and the amount of modified material were also optimized for the rapid detection of rutin. A 2-electron-2-proton mechanism, involving a few rapid and consecutive stages, was speculated to occur during rutin oxidation, based on the observed slope of -53 mV/pH. There was an appreciable linear relationship between the reductive peak current from DPV and the rutin concentration, ranging from 0.005-0.05  $\mu$ mol/L and 0.1-6  $\mu$ mol/L, with a detection limit of 0.22 nmol/L (S/N = 3). The sensor also demonstrated

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https://doi.org/10.1016/j.arabjc.2023.104613

1878-5352 © 2023 The Author(s). Published by Elsevier B.V. on behalf of King Saud University. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/). good selectivity, excellent sensitivity, and reproducibility when analyzing rutin in real plant samples, with satisfactory recovery, and was also highly consistent with results of HPLC, and thus could be used to evaluate the medicinal value of natural vegetation.

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### 1. Introduction

Rutin (vitamin P) is widely found in natural vegetation such as buckwheat, red date, and wolfberry (Al-Harbi et al., 2019; Liu et al., 2017a), and possesses several medical applications such as treating bleeding, digestive system problems, and high blood pressure (Shi et al., 2022; Sghaier et al., 2016). Therefore, quantifying the content of rutin in natural vegetation is essential to assessing and developing their medicinal value. Various analytical methods have been proposed for the qualitative and quantitative analysis of rutin, such as highperformance liquid chromatography (Kalinová et al., 2018), colorimetry (Jjgyasa, 2018), fluorescence spectrometry (Sasikumar et al., 2021), and capillary electrophoresis (Wang et al., 2016). Among these, the electrochemical method has the advantages of a fast response, low cost, simple equipment, and high sensitivity, making it suitable for the rapid detection of trace rutin in natural vegetation and evaluating the medicinal value of plants (Liang et al., 2022; Xie et al., 2022; Incebay et al., 2022; Saylakcı et al., 2021; İncebay et al., 2017; Kilic et al., 2020).

Suitable electrode materials are key to the efficient detection of rutin using electrochemical sensors, and transition metal sulfides have attracted significant attention due to their excellent physical and chemical properties (Liang et al., 2019; Diao et al., 2020; Kubendhiran et al., 2018a; Sakthivel et al., 2019). In particular, tin disulfide (SnS<sub>2</sub>) has a unique layered structure as well as favorable optical, electrical, and electrochemical properties. It has thus been widely used in sensors and lithium-ion batteries (Xu et al., 2014; Sakthivel et al., 2019; Wang et al., 2013). SnS<sub>2</sub> materials possess various morphologies, such as nanoparticles (Jiang et al., 2016), nanosheets (Mondal et al., 2021), nanoribbons (Liu et al., 2017b), and nanoflowers (Yin et al., 2019). Of these, it has been reported that application of the hierarchical nanoflower structure of SnS<sub>2</sub> as an electrode material can effectively improve electrochemical performance. The large specific surface area of SnS<sub>2</sub> promotes a fast rate of electron transfer, making it an excellent choice for the preparation of modified electrodes.

However, because  $SnS_2$  nanoparticles are prone to aggregation, which results in a reduced the electron transfer rate and affects the electrochemical performance, their use in electrode modification materials can be limited (Sakthivel et al., 2019). To overcome such issues, a combination of carbon nanotubes and other nanomaterials are often added when fabricating electrochemical sensors. The presence of carbon nanomaterials can not only retard aggregation and improve the structural stability of SnS<sub>2</sub> nanoflowers, but also improve the conductivity and sensitivity of the sensor (Pham et al., 2017; Clancy et al., 2018; Xue et al., 2016; Wang et al., 2021). As a result, integrating multi-walled carbon nanotubes and SnS<sub>2</sub> to create synergistic effects can improve selectivity and activity in electrochemical sensors.

Based on previous reports and to the best of our knowledge, there have been no reports on the use of tin disulfide nanomaterials and multi-walled carbon nanotubes with high selectivity and sensitivity for rutin detection in natural vegetation to evaluate the medicinal value of plants. In this work, three types of SnS<sub>2</sub> nanomaterials with different particle sizes were prepared, and their structure, crystal phase, and morphology were characterized by X-ray diffraction (XRD), X-ray photoelectron spectroscopy (XPS), and scanning electron microscopy (SEM). In addition, cyclic voltammetry (CV) and differential pulse voltammetry (DPV) were utilized to optimize other aspects of electrochemical detection, including the pH of the buffer solution, the scanning rate, and the amount of modified material by an SnS<sub>2</sub>/CNTs

electrochemical sensor, to realize the enhanced sensitivity and efficient electrochemical detection of rutin in the presence of common interferents. Subsequently, the viability of rutin detection was demonstrated using plant samples, which proved the feasibility of the  $SnS_2/CNTs$  electrochemical sensor.

#### 2. Experimental section

#### 2.1. Materials and reagents

Multi-walled CNTs were purchased from Adamas Co. ltd. Rutin, thioacetamide, anhydrous stannic chloride, ethanol, potassium ferricyanide, potassium hexacyanoferrate (II), sodium dihydrogen phosphate, dibasic sodium phosphate, potassium chloride, nitric acid, sodium hydroxide, and acetone were purchased from Sigma-Aldrich Chemical Co., China. All chemicals were analytically pure and used without further purification. The phosphate buffer solution (PBS, 0.1 M) was composed of NaH<sub>2</sub>PO<sub>4</sub> and Na<sub>2</sub>HPO<sub>4</sub> mixed with distilled water.

#### 2.2. Instruments and measurements

A CHI 660E electrochemical workstation was used to conduct the electrochemical experiment in a voltametric cell (10 mL) at room temperature. The three-electrode cell consisted of an SnS<sub>2</sub>/CNT-modified glassy carbon electrode (GCE) as the working electrode, a platinum wire as the auxiliary electrode, and a saturated calomel electrode (SCE) as the reference electrode. The above instruments were purchased from Chenhua Instruments Co., China. XRD was measured using a max-2600PC via ceramic-monochromatic Cu K, operating at 45 kV and 20 mA, with a scanning rate of 5° per min in  $2\theta$ and a scanning range from 20 to 65°. XPS was performed for elemental analysis with a 60 W monochromatic Mg Ka source. The binding energy was calibrated based on the carbon 1 s peak at 284.6 eV. SEM images were obtained using a Hitachi Regulus 8100, and the energy dispersive spectrometer (EDS) spectra were recorded using an EDAX EDS Element. The TEM images, selective area electron diffraction (SAED) patterns, and low-magnification scanning TEM (STEM) images were obtained using a Titan Cubed Themis G2 300 (FEI) instrument.

# 2.3. Synthesis of tin disulfide/multi-walled carbon nanotube composites

Anhydrous tin tetrachloride (0.05 M) and thioacetamide (0.15 M) were dissolved in anhydrous ethanol (240 mL), and ultrasonically treated for 2 min to obtain a clear solution. The solution was heated for 4 h, 6 h, and 8 h at 60 °C to obtain 4h-SnS<sub>2</sub>, 6h-SnS<sub>2</sub>, and 8 h-SnS<sub>2</sub> samples, respectively, which were cooled down, centrifuged for 10 min, and then washed

with deionized water and ethanol. Finally, all samples were dried in a vacuum furnace at 60 °C to obtain dry brownishyellow solids.  $SnS_2/CNT$  composites were prepared by dispersing carboxylated multi-walled carbon nanotubes and tin disulfide (ratio = 1:1) in ethanol.

### 2.4. Fabrication of 4h-SnS<sub>2</sub>/CNTs-modified electrode

The 0.05  $\mu$ m and 0.3  $\mu$ m alumina slurries were placed on suede and dissolved with twice-distilled water to form a suspension, and then the glassy carbon electrode was carefully polished for 3 min. The electrode surface was cleaned after immersion in ethanol, acetone, and twice-distilled water for ultrasonic oscillation cleaning for 5 min. Finally, a bright mirror was obtained. Next, 5  $\mu$ L of the 4 h-SnS<sub>2</sub>/CNTs composite with ethanol was dropped onto the electrode surface and dried under an infrared lamp to obtain the 4h-SnS<sub>2</sub>/CNTs modified electrode, which was gently rinsed with twice-distilled water.

# 2.5. Optimization of the experimental conditions

The experimental conditions of the  $SnS_2/CNT$  electrodes, including the pH of the buffer solution, the amount of modified material, and the scanning rate, were optimized to determine the rapid detection of rutin.

First, the 4h-SnS<sub>2</sub>/CNTs/GCE surface was immersed in 0.05 mol/L PBS at different pH values (pH 2.3, 3, 4, 5, and 6), in which the concentration of rutin was 5.0  $\mu$ mol/L. Then, the immersed electrode was applied by CV, which was performed in the range of 0.1 V to 0.8 V at a scan rate of 100 mV and sample interval of 0.001 V.

Second, the 4h-SnS<sub>2</sub>/CNTs composite with different modification amounts (1  $\mu$ L, 2  $\mu$ L, 3  $\mu$ L, 4  $\mu$ L, 5  $\mu$ L, and 6  $\mu$ L) was dropped onto the electrode surface and dried under an infrared lamp, which was gently rinsed with twice-distilled water. The modified electrode was immersed in 5.0  $\mu$ mol/L rutin solution, 0.05 mol/L PBS, and pH of 3.0. CV was recorded on the electrodes in the potential range between 0.1 and 0.8 V at a scan rate of 100 mV/s.

Thirdly, the CV of  $5.0 \,\mu$ mol/L of rutin at different scanning rates (40 mV/s, 60 mV/s, 80 mV/s, 100 mV/s, 120 mV/s, 140 mV/s, and 160 mV/s) was recorded. Then, the reproducibility test was performed using three different GC electrodes under the exact same conditions. Repeatability and stability tests were also performed multiple times using the same electrode for rutin detection.

# 2.6. Preparation of plant samples before determination

The feasibility of the sensor was ascertained using 9 different natural plant samples including red date, black fruit wolfberry, asparagus, wolfberry, grape peel, tangerine, eggplant peel, buckwheat and apples. The plant samples were prepared for analysis according to reference (Liao et al., 2015). To prepare the samples for testing, the stems and soil of the fresh vegetables were removed for later use. All samples were crushed, freeze-dried, and passed through a 40–60 mesh sieve. Then, 0.5 g of sample (accurate to 0.0001 g) was added to 70 % ethanol (12.5 mL) and then sonicated for 15 min (150 W, 20 °C, 60 ~ 62 KHz). After sonication, the samples were centrifuged

for 5 min, and finally, the supernatant was filtered with a 0.45  $\mu$ m filter membrane for later use.

## 3. Results and discussion

# 3.1. Characterization of the SnS<sub>2</sub>/CNTs composites by XRD, XPS, SEM-EDS, and TEM

SnS2 nanoparticles were prepared by the chemical bath deposition method, and their sizes were controlled by controlling reaction times. The crystal phases of the SnS<sub>2</sub>/CNTs composites and SnS<sub>2</sub> nanoparticles were characterized by XRD (Fig. 1). In comparison to the standard card (JCPDS 23-667), the peak height, peak shape, and peak angle of 4h-SnS<sub>2</sub>, 6h-SnS<sub>2</sub>, 8 h-SnS<sub>2</sub> nanoparticles were the same, and their characteristic diffraction peaks coincided. It was also found that in these materials, longer reaction times resulted in smaller half peak widths and larger grain sizes. Interestingly, compared to the XRD patterns observed in the CNTs, the diffraction peaks located at the  $2\theta$  angle of 25.8° were regenerated in the  $SnS_2/CNTs$ , which represented the (002) diffraction planes (Sakthivel et al., 2019) according to the periodicity between the graphene layers (Reznik et al., 1995). These peaks confirmed successful composite formation. In addition, in order to study the interaction between SnS<sub>2</sub> nanoparticles and CNTs, zeta potential was measured. Zeta potential is related to the surface charge and electric double layer of nanoparticles, which can be used to explain functional group performance, adsorption/desorption phenomena, chemical surface modification or stability. The potentials of  $SnS_2$  particles and CNTs are (1.29  $\pm$  0.58) mv and (-2.86  $\pm$  0.92) mV respectively. Therefore, they could combine through electrostatic attraction.

To evaluate the chemical composition, purity, integrity, and state of surface oxidation of the 4h-SnS<sub>2</sub>/CNTs composites, XPS was applied to determine the binding energy of the elements in the micro and nano materials (Kubendhiran et al., 2018a). The XPS survey scan spectra and characteristic peaks for the Sn 3d, S 2p, and C 1 s levels in the 4h-SnS<sub>2</sub>/CNT composites were as shown in Fig. 2. The peaks at 494.8 eV and



Fig. 1 XRD patterns of the  $SnS_2/CNTs$  and  $SnS_2$  nanoflowers according to the standard card (JCPDS 23–667).



Fig. 2 (A) XPS survey spectrum of the 4h-SnS<sub>2</sub>/CNTs; core level spectrum of (B) Sn 3d, (C) S 2p, (D) C 1 s.

486.4 eV (Fig. 2B) correspond to the Sn  $3d_{3/2}$  and Sn $3d_{5/2}$  spin–orbit spin components of Sn<sup>4+</sup> in SnS<sub>2</sub> (Wang et al., 2017; Xu et al., 2019). In addition, the high-resolution scan of S 2p unambiguously represents two major peaks due to S  $2p_{1/2}$  at 163.2 eV and S  $2p_{3/2}$  at 162.0 eV, separately shown in Fig. 2C (Huang et al., 2018). As shown in Fig. 2D, the three dominant peaks of the C 1 s at 285.0 eV, 286.1 eV, and 287.0 eV could be allocated to the non-oxygenated ring carbon (C—C/C=C,), the carbon in the C—O bonds, and the carbon in the —COOH bond, respectively (Kubendhiran et al., 2018b; Zhou et al., 2015).

The low-resolution SEM images (Fig. 3A–C) reveal that morphologically, the 4h-SnS<sub>2</sub>, 6h-SnS<sub>2</sub>, and 8 h-SnS<sub>2</sub> nanoparticles consisted of flower-like microspheres with a smooth surface. The size of these flower-like microspheres increased with the increase of the reaction time. The SEM images of 4h-SnS<sub>2</sub>/ CNTs (Fig. 3D) and 4h-SnS<sub>2</sub> (Fig. 3A) indicated that CNTs were uniformly adsorbed on the surface of these microspheres, thus resolving the aggregation problem of CNTs and increasing the specific surface area of the modified materials. In addition, the TEM images of the nanoparticles and the nanoparticle/CNTs materials obtained to verify the size distribution of the SnS<sub>2</sub>, 4h-SnS<sub>2</sub>, 6h-SnS<sub>2</sub>, and 8 h-SnS<sub>2</sub> nanoparticles were as shown in Fig S1. The size of the flower-like microspheres increased with increasing reaction time. Moreover, as shown in Fig. S1D, it was observed that carbon nanotubes covered the nanoflower-like  $SnS_2$  structures in the  $SnS_2/CNTs$  composites. The obtained TEM images corroborated the SEM images, both confirming the formation and presence of the  $SnS_2/CNTs$  composites. To further confirm that the synthesized materials were  $SnS_2$  and  $4h-SnS_2/CNTs$ , their elemental weight distributions and atomic contents were analyzed by SEM-EDS, and results were as shown in Fig. 3. The atomic ratio of tin and sulfur in these two materials was approximately 1:2, as given in Table S1, and a high proportion of carbon was found in the  $SnS_2/CNTs$ , indicating the successful synthesis of both  $SnS_2$  and  $4h-SnS_2/CNTs$ .

### 3.2. Electrochemical behavior

# 3.2.1. Electrochemical characterization of SnS<sub>2</sub> and SnS<sub>2</sub>/ CNTs-modified electrodes

To investigate the performances of the modified electrodes, bare GCE, 4h-SnS<sub>2</sub>/GCE, 6h-SnS<sub>2</sub>/GCE, 8 h-SnS<sub>2</sub>/GCE, CNTs/GCE, and 4h-SnS<sub>2</sub>/CNTs/GCE were scanned in a  $K_3$ [Fe(CN)<sub>6</sub>]/K<sub>4</sub>[Fe(CN)<sub>6</sub>] solution (5 mmol/L) using cyclic voltammetry (CV). The electrochemical performances of the modified electrodes were analyzed by comparing their redox



Fig. 3 SEM images of  $SnS_2$  and  $SnS_2/CNTs$ : (A) 4h-SnS<sub>2</sub>, (B) 6h-SnS<sub>2</sub>, (C) 8 h-SnS<sub>2</sub>, (D) 4h-SnS<sub>2</sub>/CNTs, and SEM-EDS spectra of (E) 4h-SnS<sub>2</sub> and (F) 4h-SnS<sub>2</sub>/CNTs.



Fig. 4 CV and EIS of different electrodes in 0.1 mol/L of KCl solution containing 5 mmol/L of K[Fe(CN)<sub>6</sub>]<sup>3-/4-</sup> at a scan rate of 100 mV/s: (A) GCE, 4h-SnS<sub>2</sub>/GCE, 6h-SnS<sub>2</sub>/GCE, 8h-SnS<sub>2</sub>/GCE, (B) GCE, 4h-SnS<sub>2</sub>/GCE, CNTs/GCE, 4h-SnS<sub>2</sub>/CNTs/GCE and (C) EIS of different modified electrodes recorded.

peak currents and peak potentials. As discernible in Fig. 4A, the redox peak current of  $Fe^{2+}/Fe^{3+}$  pairs for bare GCE was the smallest, whereas the redox peak current of  $SnS_2/$ GCE was slightly higher, indicating that tin disulfide had a catalytic effect on the redox reaction. Moreover, the peak current of 4h-SnS<sub>2</sub>/GCE was found to be the largest, followed by 6h-SnS<sub>2</sub>/GCE and 8 h-SnS<sub>2</sub>/GCE. These results support that smaller particle sizes correspond to larger specific surface areas, and thus better electron transfer ability and stronger conductivity. Bare GCE was modified with CNTs to further enhance its current response and results were as illustrated in Fig. 4B. As it was apparent that the electrochemical performance of CNTs was better than that of SnS<sub>2</sub>, 4h-SnS<sub>2</sub> nanoparticles were combined with CNTs to form a 4h-SnS<sub>2</sub>/ CNTs composite. The modified electrode based on 4h-SnS<sub>2</sub>/ CNTs (4h-SnS<sub>2</sub>/CNTs/GCE) had the highest response because 4h-SnS<sub>2</sub>/CNTs had excellent electrocatalytic ability for  $Fe^{2+}$ / Fe<sup>3+</sup> pairs.

Electrochemical impedance spectroscopy (EIS) can provide useful information regarding impedance changes on the electrode surface during the fabrication process, further verifying its results (Fig. 4C). The charge-transfer resistance was related to the electron-transfer kinetics of the redox probe ([Fe  $(CN)_{6}]^{3-}/[Fe(CN)_{6}]^{4-})$  at the electrode interface and could be estimated from the diameter of the semicircular part of the EIS curve. Fig. 4C illustrates the typical Nyquist plots obtained from bare GCE, 8h-SnS<sub>2</sub>/GCE, 6h-SnS<sub>2</sub>/GCE, 4h-SnS<sub>2</sub>/GCE, CNTs/GCE, and 4h-SnS<sub>2</sub>/CNTs/GCE in aqueous KCl (0.1 mol/L) containing 5.0 mmol/L [Fe(CN)<sub>6</sub>]<sup>3-</sup>/[Fe (CN)<sub>6</sub>]<sup>4-</sup>. A large diameter of the semicircle part was observed for GCE. The diameter of the semicircle was reduced when the GCE surface was coated with SnS2 nanoparticles due to improved electrical conductivity. In addition, the smaller surface area facilitated electron transport, and thus the semicircle diameter of 4h-SnS<sub>2</sub>/GCE was much smaller. The semicircle diameter showed an even greater decrease in CNTs/GCE due to the excellent electron transfer ability of the carbon nanotubes. The electrochemical impedance spectrum of 4h-SnS<sub>2</sub>/ CNTs/GCE had the smallest semicircle diameter, which proved that the fastest electron-transfer kinetics of  $[Fe(CN)_6]^{3-1}$ <sup>-/4-</sup> occurred in 4h-SnS<sub>2</sub>/CNTs/GCE rather than in the other

modified electrodes. The EIS results were in full agreement with the CV results.

To understand the electrochemical behavior of each modified electrode, the electrochemically active surface area (A) of each electrode could be derived from Randles Sevcik Eq. (1) (Zhang et al., 2020; Rao et al., 2018; Prasad et al., 2017):

$$Ip = 268600n^{3/2}AD^{1/2}CV^{1/2}$$
(1)

where Ip is the peak current (A), n is the number of electrons exchanged during the electron transfer process, A is the active surface area (cm<sup>2</sup>), D is the diffusion coefficient (cm<sup>2</sup>/s), C is the concentration (mol/cm<sup>3</sup>), and V is the scan rate (1/Vs). Because the effective physical radius was 1.5 mm, the value of the electrochemically active surface area of 4h-SnS<sub>2</sub>/ CNTs/GCE could be estimated as 0.30 cm<sup>2</sup>, which was 4.23 times that of GCE. The high electrocatalytic properties could be attributed to the large surface area as well as the greater electron transfer capability of 4h-SnS<sub>2</sub>/CNTs/GCE compared to the other electrodes.

# 3.2.2. Electrochemical responses of various modified electrodes in rutin solutions

The CV curves of bare GCE and various modified electrodes in 5  $\mu$ mol/L of rutin in 0.05 mol/L PBS (pH 3.0) were as shown in Fig. 5. At a scanning rate of 100 mV/s, a reduction peak was observed at 0.48 V and the electrochemical responses of the SnS<sub>2</sub>-modified electrodes to rutin were each slightly different (Fig. 5A). The response current of rutin in 4h-SnS<sub>2</sub>/GCE with the smallest particle size was found to be the largest, which suggested that it was the most suitable material for electrodes. As evident in Fig. 5B, the response currents of GCE, 4h-SnS<sub>2</sub>/GCE, CNTs/GCE, and 4h-SnS<sub>2</sub>/CNTs/GCE increased significantly. The 4h-SnS<sub>2</sub>/CNTs composite had a large specific surface area and good conductivity. Hence, faster electron transfer in the electrode was due to improved catalytic efficiency.

#### 3.3. Optimization of electrochemical experimental conditions

#### 3.3.1. Effect of buffer solution pH

The pH value of the buffer solution is considered a significant influential factor that will influence on the results of electro-



Fig. 5 CV of different electrodes in 5  $\mu$ mol/L of rutin in 0.05 mol/L PBS (pH 3.0) at a scan rate of 100 mV/s: (A) SnS<sub>2</sub>-modified electrodes and (B) modified electrodes.

chemical detection. Therefore, it was necessary to select the appropriate pH of PBS to optimize the sensitivity for rutin determination. As shown in Fig. 6, increase in the pH of the buffer solution moved the peak potential of rutin moved in the negative direction. A linear relationship was observed between the values of potential and pH and could be expressed as *potential* = -0.0530 *pH* + 0.6219 ( $R^2 = 0.9947$ ). The slope in this relationship (53 mV/pH) approximated the theoretical value of -59 mV/pH. This indicated that the proton number and electron transfer number involved in rutin oxidation on 4h-SnS<sub>2</sub>/CNTs/GCE electrodes were equal, implying that it

possibly followed the 2-electron-2-proton mechanism (El Jaouhari et al. 2020). According to Scheme 1, the electrochemical oxidation mechanism of rutin involved several rapid and continuous stages, which was first triggered by electron transfer to produce phenoxy radicals in the form of four mesomeric intermediates (A-D). Due to the electronic effects of the hydroxyl groups, the structure of radical B was the most stable. However, it instantly experienced electron transfer once more, producing the carbocation, and then was dehydrated rapidly, transforming into the final product 30, 40-diquinone (E), which could be reduced to the original rutin in the reverse reaction



Fig. 6 (A) CV of 5.0  $\mu$ mol/L of rutin in 0.05 mol/L pH phosphate buffer solutions (pH = 2.3, 3, 4, 5, 6) at a scan rate of 100 mV/s; (B) effects of different pH values on the peak potential, and (C) effects of different pH values on the peak current.



Scheme 1 Electrochemical oxidation mechanism of rutin.

(Zhang et al., 2015; Sun et al., 2008). Moreover, the peak current first increased and then decreased as the pH of the buffer solution increased, which indicated that the electrode reaction of rutin required the participation of protons. Rutin has four phenolic hydroxyl groups, which have difficulty reacting electrochemically due to the lack of protons under alkaline conditions. The optimal pH of buffer solution was selected to be 3 based on the pH at which the largest peak current was achieved.

# 3.3.2. Effect of 4h-SnS<sub>2</sub>/CNTs dosage

The peak current of rutin will also be affected by the extent of modification to a composite. Generally, the larger the specific surface area of the electrode material, the higher the number of active sites in the electrode. However, when a high amount of modified material results in an overly thick modified film, electron transfer in the electrode is hindered and the electrochemical response is reduced. Therefore, it is necessary to control the dosage of 4h-SnS<sub>2</sub>/CNTs composite to improve its sensitivity to rutin (Fig. S2). When 5 µL of the modified materials was used, 4h-SnS<sub>2</sub>/CNTs/GCE had the highest response to rutin. However, its electrochemical response decreased as the thickness of the modified layer increased due to the restriction to the ionic transfer rate with excessive materials. Therefore, 5.0 µL of 4h-SnS<sub>2</sub>/CNTs nanocomposites was considered the optimal amount to promote electrochemical activity with extensive active sites and edges.

# 3.3.3. Effects of scanning rate and the mechanism of electro surface reaction

An increase in scanning rate from 40 mV/s to 180 mV/s resulted in not only the substantial enhancement of the oxidation and reduction peak current of rutin (Fig. 7), but also a linear movement towards the negative potential in the oxidation and reduction peak current, demonstrating that the surface of 4h-SnS<sub>2</sub>/ CNTs/GCE had improved adsorption capacity and electron transfer kinetics. The linear relationship between the anodic peak current (*Ipa*) and cathodic peak current (*Ipc*) could be expressed as *Ipa* ( $\mu$ A) = -0.2567v (mV/s) - 1.6664 (R<sup>2</sup> = 0.9992) and *Ipc* ( $\mu$ A) = 0.2181v (mV/s) + 0.3506 (R<sup>2</sup> = 0.9998). These results indicated that the electrochemical behavior of rutin on 4h $SnS_2/CNTs/GCE$  exhibited adsorption-controlled electron transfer without any fouling effect (El Jaouhari et al. 2020; Yao et al., 2019; Chen et al. 2019). As the scanning rate decreased, the current response of rutin decreased and the sensitivity worsened. In contrast, as the scanning rate increased, the current response and sensitivity both increased, whereas reversibility of the redox reaction decreased. Therefore, the optimal scanning rate was determined to be 100 mV/s.

### 3.4. Linearity, interference, reproducibility, and stability

To explore the electrochemical performance of 4h-SnS<sub>2</sub>/CNTs/ GCE. DPV was applied to determine the concentration of rutin in optimal experimental conditions, with results as shown in Fig. 8. The optimal concentrations of rutin were 0.005-0.05 µmol/L and 0.1-6.0 µmol/L. A linear relationship was observed between the Ipa and concentration (C) of  $4h-SnS_2/$ CNTs/GCE. At low concentrations, this relationship could be expressed as the equation  $Ipa (\mu A) = -37.85C (\mu mol/L)$  -0.0162 ( $\mathbb{R}^2 = 0.9947$ ), and at high concentrations, as Ipa  $(\mu A) = -20.24C \ (\mu mol/L) - 0.9073 \ (R^2 = 0.9857).$  According to the signal measured in the blank solution, which was determined using the SnS<sub>2</sub>/CNTs electrochemical sensor measuring 6 times in parallel, the current was about 0.019  $\mu$ A (Fig. S3). Based on the equation Ipa ( $\mu A$ ) = -37.85C ( $\mu mol/L$ ) -0.0162 ( $\mathbf{R}^2 = 0.9947$ ) as well as the relationship between the current signal of lowest detection limit and the noise signal (S/N = 3), the limit of detection (LOD) was calculated to be 0.22 nmol/L.

It was also found that compared to other common methods of rutin detection, the electrochemical method used in this experiment had the lowest detection limit and the highest sensitivity (Table 1). Therefore, the proposed electrochemical method was fast, simple, and sensitive for rutin determination.

In real sample analysis, the interference of unknown substances is inevitable, thus, anti-interference is an important characteristic of modified electrodes. Ions and molecules that might interfere with the experiment were added to measure the electrochemical response of  $4h-SnS_2/CNTs/GCE$ . The interference of the  $4h-SnS_2/CNTs$  modified electrode was



Fig. 7 (A) CV of 5.0  $\mu$ mol/L of rutin in 0.05 mol/L PBS (pH 3.0) at different scanning rates (40 mV/s, 60 mV/s, 80 mV/s, 100 mV/s, 120 mV/s, 140 mV/s, and 160 mV/s); (B) linear relationship between the scanning rate and response peak current.



**Fig. 8** DPV curves in 0.05 mol/L PBS (pH 3.0) at a scan rate of 100 mV/s for rutin concentrations of (A) 0.005–0.050  $\mu$ mol/L and (B) 0.1–6.0  $\mu$ mol/L, (C) standard curve in the low concentration range of rutin (0.005–0.050  $\mu$ mol/L), and (D) standard curve in the high concentration range of rutin (0.1–6.0  $\mu$ mol/L).

Table 1 Performances of different methods for rutin detection.						
Detection method	Innovation	LOD (mol/L)	Reference			
Capillary electrophoresis	Response surface methodology	$4.97 \times 10^{-6}$	Martí et al., 2017			
	Photodiode array detector.	$5.45 \times 10^{-7}$	Memon et al., 2017			
Fluorescence spectrometry	Cysteine-stabilized copper nanoclusters	$21 \times 10^{-8}$	Wang et al., 2016			
	MoS <sub>2</sub> quantum dots	$3.50 \times 10^{-7}$	Sun et al., 2020			
High-performance liquid chromatography	UHPLC/ESI-Q-TOF-MS/MS	$1.35 \times 10^{-7}$	Ahmad et al., 2016			
	Modified QuEChERS method	$1.20 \times 10^{-7}$	Rotta et al., 2019			
Electrochemical method	Cu <sub>2</sub> O-Au/nitrogen-doped graphene nanocomposites	$3 \times 10^{-8}$	Li et al., 2017			
	rGO-InTAPc/GCE	$2 \times 10^{-9}$	Shi et al., 2022			
	SnS <sub>2</sub> /CNTs/GCE sensor	$1.67 \times 10^{-9}$	This report			

explored in solutions of 5  $\mu$ mol/L rutin, with the presence of ions (50-fold concentration) or organic substances (10-fold concentration). As shown in Fig. 9, I0 and I were the Ipa of rutin in 5  $\mu$ mol/L solutions before and after adding the interferents, respectively. The results suggested that the interferents had no significant impact on the determination of rutin by 4h-SnS<sub>2</sub>/CNTs/GCE, which indicated good selectivity for rutin in the electrodes.

The storage stability test was investigated by DPV in a solution of 5.0  $\mu$ M rutin. Notably, after 10 days of storage, the peak current was reduced by 8.2 %. Furthermore, the repeata-

bility was assessed using the same 4h-SnS<sub>2</sub>/CNTs/GCE after 6 successive tests (RSD  $\leq$  3.83 %), and reproducibility was calculated 6 times using 6 different 4h-SnS<sub>2</sub>/CNTs/GC (RSD  $\leq$  4. 18 %). The above results verified that the sensor has reliable electrochemical performance.

### 3.5. Analysis of real samples

After the synthesis and optimization of 4h-SnS<sub>2</sub>/CNTs, a 4h-SnS<sub>2</sub>/CNTs/GCE sensor was fabricated to detect rutin in medicine samples. This sensor was applied to detect trace levels of



Fig. 9 Selectivity of 4h-SnS<sub>2</sub>/CNTs/GCE toward rutin.

rutin in real samples. Twenty tablets of rutin were weighed to be 1.8481 g in total, and the average weight of each was 92.41 mg. The content of rutin in 1 g tablets was 0.2164 g, and 1.099 µmol/L of rutin solution was prepared from a small amount of powder. The peak current response for 4h-SnS<sub>2</sub>/ CNTs/GCE was calculated, and the average rutin concentration was 1.06 µmol/L (RSD  $\leq$  3.58 %). When the tablets were spiked with three levels of rutin (5.00 µM, 10.00 µM, and 15.00 µM), the recoveries ranged between 96.6 % and 102.3 % (RSD  $\leq$  3.5 %, n = 5) (Table 2). Therefore, the 4h-SnS<sub>2</sub>/CNTs/GCE delivered good analytical performance in terms of sensitivity, precision, and accuracy for rutin detection in medicine samples.

Additionally, the practicability of the 4h-SnS<sub>2</sub>/CNTs/GCE sensor was explored by analysis of nine different natural plants prepared as described in section 2.6. The DPV and HPLC results under optimal conditions were as presented in Table 3. Noticeably, results from DPV and HPLC analyses were in strong agreement. This validated that the 4h-SnS<sub>2</sub>/CNTs/GCE sensor had excellent analytical performance in terms of precision and could be applied the quantification of rutin in plant samples.

Samples	Average concentration	Average concentration (µM)		
	DPV	HPLC		
Red date	98.71 ± 4.73	101.23		
Black fruit wolfberry	$272.82 \pm 18.63$	280.09		
Asparagus	$676.29 \pm 19.15$	673.85		
Wolfberry	$430.63 \pm 22.91$	422.26		
Grape peel	$291.96 \pm 6.05$	293.69		
Tangerine	$217.50 \pm 5.87$	215.23		
Eggplant peel	$268.94 \pm 13.38$	280.09		
Buckwheat	$53.13 \pm 1.48$	55.69		
Apple	$55.42~\pm~3.47$	57.98		

Table 2Recovery rat	es of rutin in tablets
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Samples	Rutin spiked (µM)	Rutin found (µM)	Recovery (%)	RSD (n = 5, %)
Rutin	0.00	1.06	-	3.58
tablets	5.00	6.83	96.6	3.29
	10.00	11.23	102.3	3.41
	15.00	16.18	101.2	3.19

### 4. Conclusion

In summary, a rapid and selective electrochemical sensor was proposed and synthesized in this study for the determination of rutin concentration. Nanoflower-like structured SnS2 was successfully synthesized and composited with multi-walled carbon nanotubes via simple sonochemical treatments. The prepared SnS2/CNTs modified electrode was then verified to possess higher effectiveness in the electrochemical determination of rutin, exhibiting a good linear range from 0.005 -0.05 µmol/L and 0.1-6.0 µmol/L with an LOD value of 0.22 nmol/L (S/N = 3). In contrast to previous methods, the proposed sensor is characterized by good anti-interference properties, superior reproducibility and repeatability, and long-term stability. Moreover, the fabricated sensor was successfully applied to determine rutin not only in commercially available rutin tablets, but also in natural vegetation to evaluate the medicinal value of plants. Therefore, this strategy could be further applied to the development of sensing devices for analyze different substances.

# CRediT authorship contribution statement

Ying Liang: Conceptualization, Methodology, Investigation, Formal analysis, Validation, Visualization, Writing – original draft, Funding acquisition. Lingyu Zhang: Formal analysis, Investigation, Validation. Hongmei Wang: Formal analysis, Investigation, Validation. Xinru Cai: Formal analysis, Investigation, Validation. Li Zhang: Formal analysis, Investigation, Validation. Li Zhang: Formal analysis, Investigation, Validation. Li Zhang: Formal analysis, Investigation, Validation. Yixin Xu: Formal analysis, Investigation, Formal analysis, Validation, Methodology, Investigation, Formal analysis, Validation, Visualization, Funding acquisition. Wenshuai Si: Conceptualization, Resources, Supervision, Methodology, Project administration. Zhipeng Huang: Conceptualization, Resources, Supervision, Methodology, Project administration. Guoyue Shi: Conceptualization, Resources, Supervision, Methodology, Project administration.

### **Declaration of Competing Interest**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

### Funding

This work was supported by the Foundation of Shanghai Municipal Commission of Science & Technology Commission (20392002200) and the Fundamental Research Funds of Shanghai University of Medicine & Health Sciences (HXXM-20-07-004, SSF-21-07-001).

# Appendix A. Supplementary material

Supplementary data to this article can be found online at https://doi.org/10.1016/j.arabjc.2023.104613.

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