



ORIGINAL ARTICLE

Design, characterization and quantum chemical computations of a novel series of pyrazoles derivatives with potential anti-proinflammatory response

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Abstract The synthesis and characterization of the full family of 11 pyrazoles were performed by means of UV–Vis, FTIR, ¹H NMR, ¹³C NMR, two-dimensional NMR experiments and DFT simulations. As pyrazoles are known for showing diverse biological actions, they were also tested in the NCI-60 cancer cell line panel, showing moderate to good activity against different cell lines.

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NCI-60;
DFT

Furthermore, the anti-proinflammatory activity test of a set of pyrazoles of the form (*E*)-4-((4-bromophenyl)diazanyl)-3,5-dimethyl-1-*R*-phenyl-1*H*-pyrazole was performed, this is based on the study of the blockage of the increase in intracellular $[Ca^{2+}]$ observed in response to platelet-activating factor (PAF) treatment of four pyrazoles (*i.e.* **6**, **8**, **9** and **10**), which successfully displayed $[Ca^{2+}]$ channel inhibition. Therefore, the obtained intracellular $[Ca^{2+}]$ signal results indicate that the pyrazole family characterized in this study, in particular compounds **6** and **10**, are potent blockers of the PAF-initiated Ca^{2+} signaling that mediates the hyperpermeability typically observed during the development of inflammation.

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1. Introduction

The biological activity of pyrazole derivatives, see Fig. 1a, has been a focus of interest in recent years, because they have been subject of a variety of medical investigations. Those medical investigations mainly have been oriented to determine the potential pharmacological properties of these molecules. The pyrazole nuclei is present in numerous compounds with biological activity (Elguero et al., 2001), including very successful drugs, *v. g.* Celebrex (Penning et al., 1997) and Viagra (Terrett et al., 1996). Additionally, pyrazoles are used as drugs for the treatment of anxiety (Wustrow et al., 1998), as antipyretics, analgesics and anti-inflammatories (Menozzi et al., 1997; Surendra Kumar et al., 2016) and have shown antimicrobial activity (Abunada et al., 2009; B'Bhatt and Sharma, 2017; Kumar et al., 2014) and antiparasitic activity (Escario et al., 1988). Compounds of this family have been described as potent inhibitors of PDE4B or PDE4D (Card et al., 2005), GABA antagonist receptors, insecticides (Sammelsson et al., 2004), others are potential ligands for the CB1 receptor in PET trials (Kumar et al., 2004) and have also shown growth inhibitory activity (Baraldi et al., 2003).

Although various methods have been developed to prepare pyrazoles, the regioselective synthesis remains a challenge for chemists (Elguero, 1996, 1984; Makino et al., 1998). The most commonly reported method is the reaction of hydrazines with 1,3-dicarbonyl compounds, where the reaction often results in a mixture of isomers, as the reactivity of the carbonyl groups is not sufficiently different. An alternative is the replacement of 1,3-dicarbonyl compounds with α,β -ethynyl ketones or α -ketoesters, which allows a much more rigorous control of regioselectivity (Bishop et al., 2003; Miller and Reiser, 1993;

Norris et al., 2005). Another important method is the 1,3-dipolar cycloaddition of diazoalkanes or nitriloidines with olefins or alkynes. This reaction has limited applications in the synthesis of pyrazoles as the 1,3-dipoles are difficult to prepare and are potentially explosive. However, this method has high regioselectivity (Aggarwal et al., 2003; Foti et al., 1999; Huisgen and Huisgen, 1963; Jung and Trifunovich, 1992; Nakano et al., 1989; Padwa, 2009). The reaction of hydrazones with activated alkenes (Grigg et al., 1987; Le Fevre and Hamelin, 1980; Osella et al., 1995) or alkynes (Bardakos et al., 1975) has also been reported to produce pyrazolidines or pyrazoles, respectively, and there is believed that the mechanism of reaction involves a 1,3-dipole molecule generated *in situ* (Grigg et al., 1987). In addition, the reaction requires rigorous conditions, since hydrazones have low nucleophilicity and the activation of the alkene or alkyne is often complicated. In contrast, nitroolefins are strong electrophiles widely used in organic synthesis (Barrett, 1991; Barrett and Graboski, 1986; Denmark et al., 2001) and have been found to react with diazo compounds to give pyrazoles (Mancera et al., 1988; Parham et al., 1961; Parham and Bleasdale, 1951). Therefore, it can be concluded that hydrazones and nitroolefins are ideal precursors to synthesize pyrazoles, mainly because they are available at low cost or are comfortably prepared (Deng and Mani, 2006).

In literature, the synthesis of an extensive family of 3-(2-(*R*-phenyl) hydrazono) pentane-2,4-dione (β -diketohydrazones) has been described. These compounds are prepared by a coupling reaction of one diazonium salt with a β -diketone (Yao, 1964). Those compounds retain the dicarbonyl system of β -diketone used as a precursor, contain a resonance assisted intramolecular hydrogen bond (RAHB) that

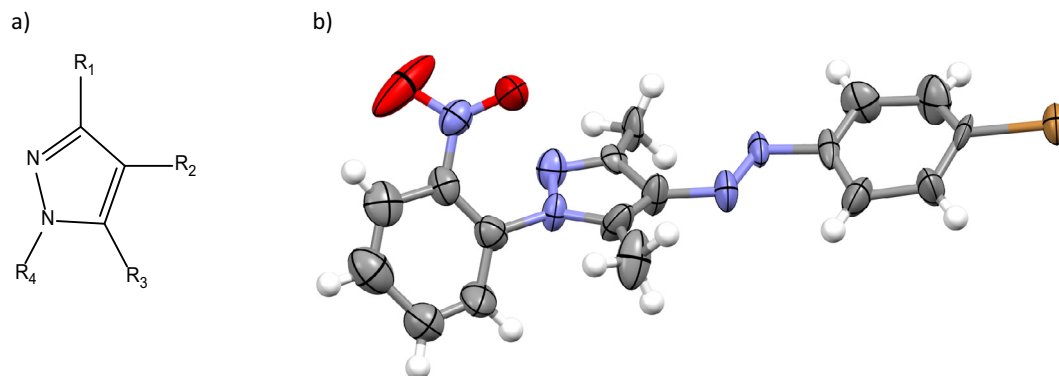


Fig. 1 a) Structure of the pyrazole ring and b) Ortep view of compound 9.

increases the electrophilic character of the carbonyl group involved, thus the compounds do not undergo tautomerization processes.

Previous reports show that in acidic media β -diketohydrazone react with substituted phenylhydrazines to generate pyrazoles of the form (E) -3,5-dimethyl-1- (R 1 -phenyl) -4- (R 2 -phenyldiazenyl) - 1H-pyrazoles (Bustos et al., 2012, 2009).

On the other hand, among the valuable biological activities that pyrazoles have shown, it has been reported that they exhibit anti-inflammatory properties. In this sense, as the development of inflammation depends on an increase in $[Ca^{2+}]_i$, several pyrazole analogues have been found to be potent blockers of store-operated Ca^{2+} channels and T-type Ca^{2+} channels ($Ca_v3.1$, $Ca_v3.2$ and $Ca_v3.3$) (Bezençon et al., 2017; Dago et al., 2018). The anti-inflammatory effect of pyrazoles may be related to the modulation of Ca^{2+} signals, however this relation has not been determined. It is very well known that inflammation is a complex process mediated by the interaction of several signals and, among them, platelet-activating factor (PAF) is an important and potent pro-inflammatory agent. Endothelial cells of post-capillary venules are one of the most relevant targets of PAF during inflammation (Jiang et al., 2008), where it triggers a strong Ca^{2+} -mediated increase in the endothelial cell barrier to macromolecules, leading to edema (Tiruppathi et al., 2002).

In the herein report, our aim is the design and synthesis of a complete family of pyrazoles derived from 3-(2-(4-bromophenyl)hydrazinylidene)pentane-2,4-dione by treatment with substituted arylhydrazines, in acid medium. These pyrazoles have a general name (E)-4-((4-bromophenyl)diazenyl)-3,5-dimethyl-1-R-phenyl-1H-pyrazole (with R = 4-OCH₃ (1), 4-H (2), 4-F (3), 4-Cl (4), 4-CN (5), 4-NO₂ (6), 3-Cl (7), 3-NO₂ (8), 2-NO₂ (9), 2,4-(NO₂)₂ (10), C₆F₅-NHNH₂ (11)). The products obtained in these reactions were characterized by analytical techniques (MP, MS-HPLC), spectroscopic (UV-visible, FTIR, ¹H NMR, ¹³C NMR), bidimensional experiments such as DEPT-135, HMBC and HMQC and theoretical calculations in the Density Functional Theory framework. Finally, the previously reported biological activity that pyrazoles have shown, encourage us to carry out antiproliferative tests over a set of 60 cell lines including brain cancer, breast cancer and prostate cancer, among a large list, as well as anti-proinflammatory studies where the inhibitory activity of these compounds on the response initiated by PAF, in primary cultures of mesenteric post-capillary venules endothelial cells was evaluated.

2. Chemistry

2.1. Experimental

All of the synthesis reagents, *p*-bromoaniline, acetylacetone, sodium nitrite, sodium hydroxide, hydrochloric acid, sodium acetate and substituted arylhydrazines, R-C₆H₄-NH-NH₂ were purchased from Sigma-Aldrich Corporation. Solvents, methanol, ethanol, CHCl₃ were obtained from common commercial sources (Merck, Fisher and T.J. Baker) and used without purification. The precursor 3-(2-(4-bromophenyl)hydrazinylidene)pentane-2,4-dione was prepared as described in the literature (Bertolasi et al., 2006; Yao, 1964). Its structure was corroborated by FTIR spectroscopy.

2.2. Physical measurements

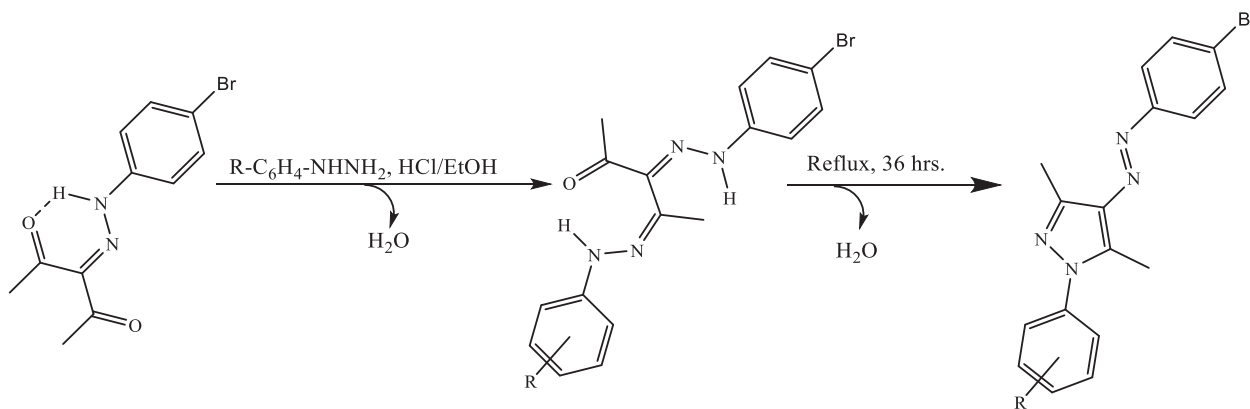
The melting points were recorded on an Electrothermal digital recorder, Model IA 9000 SERIES. Elemental analysis was performed on a PerkinElmer 2400. The molecular mass of these compounds was determined by dissolution chromatography on a Micromass equipment, model ZQ. The determination was carried out in 0.1% formic acid solution in methanol. The UV-visible spectra were obtained using concentrated solutions of CHCl₃, 5×10^{-4} mol/L and diluting to 5×10^{-5} mol/L. Recording was performed on a Perkin Elmer spectrophotometer, model Lambda 35 in quartz cells of 10 mm path length, in the range of 1100–220 nm. The infrared spectra were obtained in KBr pellets in a Jasco model FTIR-4200, the measurement was performed between 4000 and 550 cm⁻¹. ¹H, ¹³C NMR, DEPT-135, HMQC and HMBC spectra were recorded by conventional procedures in 5 mm diameter glass tubes in CDCl₃ solution on a Bruker spectrometer, Model AC 200P 400 MHz spectrophotometer, using as reference signal of the solvent CDCl₃.

2.3. Synthesis

3.5 mmol (1.0 g) of 3-(2-(4-bromophenyl)hydrazinylidene)pentane-2,4-dione are added to a distillation flask, 3.5 mmol of an arylhydrazine, R-C₆H₄-NH-NH₂ [98%, 0.63 g, 4-CH₃O (1);97%, 0.39 g, 4-H (2);97%, 0.58 g, 4-F (3);98%, 0.64 g, 4-Cl (4);97%, 0.61 g, 4-CN (5);97%, 0.55 g, 4-NO₂ (6); ≥ 97%, 0.65 g, 3-Cl (7);98%, 0.67 g, 3-NO₂ (8); 97%, 0.56 g, 2-NO₂ (9);0.70 g, 2,4-(NO₂)₂ (10);97%, 0.72 g, C₆F₅-NHNH₂ (11)], 2 mL of hydrochloric acid and 30 mL of ethanol. The mixture was refluxed with stirring for 36 h, see Fig. 2. It is then cooled to room temperature and the solid obtained is filtered with vacuum, washed with water (500 mL) and dried in a vacuum oven at 40 °C. All compounds were recrystallized from solvents such as ethanol, acetone, and/or ethanol mixture with tetrahydrofuran.

2.4. Data Collection

Single crystals X-Ray diffraction data set was collected at 294 K for 9. Compound 9 up to a max 2θ of ca. 52° on an Enraf-Nonius CAD diffractometer, using monochromatic MoKα radiation, λ = 0.71073 Å. Data processing was performed using WinGX program in the diffractometer package (Farrugia, 1999). Fig. 1 exhibits an ORTEP view of the compound. Table S1, shows selected bonds length and torsion angles. The data collection and structural refinement are given in Table S1. The structures were solved by direct methods using the SIR-2004 program (Burla et al., 2005). All calculations to solve the structures to refine the model proposed and to obtain results were carried out with the computer programs SHELXL97 (Sheldrick, 1990). Crystallographic data (excluding structure factors) for the structures reported in this paper have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication, CCDC No. 1,589,910 for 9. Copies of this information may be obtained free of charge from The Director, CCDC, 12 Union Road, Cambridge CB2 1EZ, UK. Fax: +44 1223 336 033. E-mail: data_request@ccdc.cam.ac.uk. Web page: <http://www.ccdc.cam.ac.uk>.



R: 4-OCH₃ (1), 4-H (2), 4-F (3), 4-Cl (4), 4-CN (5), 4-NO₂ (6), 3-Cl (7), 3-NO₂ (8), 2-NO₂ (9); 2,4-(NO₂)₂ (10); C₆F₅-NHNH₂ (11)

Fig. 2 General synthetic procedure and structure of the synthesized pyrazoles.

2.5. Chemical characterization

(E)-4-((4-bromophenyl)diazenyl)-1-(4-methoxyphenyl)-3,5-dimethyl-1H-pyrazole (1): Yield: 61.3%, crude. Recrystallized from EtOH. Melting point: 148.0–148.2° C. Molecular mass (g/mol), calculated for C₁₈H₁₇BrN₄O: 384.26 g/mol (⁷⁹Br) and 386.26 g/mol (⁸¹Br); found 384.00 g/mol (⁷⁹Br) and 386.00 g/mol (⁸¹Br). Elemental Analysis: Calculated: C, 56.12; H, 4.45; Br, 20.74; N, 14.54; O, 4.15; Found: C, 56.10; H, 4.43; Br, 20.54; N, 14.74; O, 4.15. UV-visible spectrum in CHCl₃ 5 × 10⁻⁵ mol/L, λ (nm) (logε): λ₁, 439 (3.32); λ₂, 347 (4.39); λ₃, 240 (4.21). Infrared spectrum in KBr $\bar{\nu}$, cm⁻¹ Tablets: $\bar{\nu}$ (C–H) Arom.: 3078w, 3060w; 3002w; $\bar{\nu}$ (C–H) Aliph.: 2985w, 2962w, 2939w, 2919w, 2837w; $\bar{\nu}$ (N=N), $\bar{\nu}$ (C=N) and/or $\bar{\nu}$ (C=C): 1590w, 1571w, 1553w, 1519 s; $\bar{\nu}$ (N–N): 1412 m. 1H NMR (400 MHz, CDCl₃) δ 7.69 (w, J = 8.0 Hz, 2H), 7.59 (w, J = 8.2 Hz, 2H), 7.39 (w, J = 8.1 Hz), 7.01 (J = 8.1 Hz, 2H), 3.87 (s, 3H), 2.59 (s, 3H), 2.56 (s, 3H). NMR (101 MHz, CDCl₃) δ 159.57, 152.56, 143.87, 139.53, 136.05, 132.23 (+), 132.20, 126.57 (+), 123.55, 123.52 (+), 114.55 (+), 55.74 (+), 14.18 (+), 11.40 (+).

(E)-4-((4-bromophenyl)diazenyl)-3,5-dimethyl-1-phenyl-1H-pyrazole (2): Yield: 77.2%, crude. Recrystallized from EtOH. Melting point: 134.6–134.9° C. Molecular mass (g/mol) calculated for C₁₇H₁₅N₄Br: 354.23 g/mol (⁷⁹Br) and 356.23 g/mol (⁸¹Br); found 354.00 g/mol (⁷⁹Br) and 356.00 g/mol (⁸¹Br). Elemental Analysis: Calculated: C, 57.48; H, 4.26; Br, 22.49; N, 15.77; Found: C, 57.40; H, 4.16; Br, 22.59; N, 15.85. UV-visible spectrum in CHCl₃ 5 × 10⁻⁵ mol/L mol/L, λ (nm) (logε): λ₁, 431 (3.32); λ₂, 344 (4.38); λ₃, 240 (4.14). Infrared spectrum in KBr pellets, $\bar{\nu}$, cm⁻¹: $\bar{\nu}$ (C–H) Arom.: 3083w, 3068w; 3048w; $\bar{\nu}$ (C–H) Aliph.: 2987w, 2961w, 2923w; $\bar{\nu}$ (N=N), $\bar{\nu}$ (C=N) and/or $\bar{\nu}$ (C=C): 1597 m, 1584w, 1570 m, 1552 m, 1509 s; $\bar{\nu}$ (N–N): 1414 s. 1H NMR (400 MHz, CDCl₃) δ 7.70 (w, J = 8.1 Hz, 2H), 7.60 (w, J = 8.3 Hz, 2H), 7.55–7.37 (m, 5H), 2.65 (s, 3H), 2.58 (s, 3H). NMR (101 MHz, CDCl₃) δ 152.53, 144.08, 139.62, 139.18, 136.34, 132.25 (+), 129.42 (+), 128.32 (+), 125.05 (+), 123.67, 123.55 (+), 14.26 (+), 11.52 (+).

(E)-4-((4-bromophenyl)diazenyl)-1-(4-fluorophenyl)-3,5-dimethyl-1H-pyrazole (3): Yield: 64.6%, crude. Recrystallized

from EtOH. Melting point: 145.8–146.6° C. Molecular mass (g/mol), calculated for C₁₇H₁₄BrFN₄: 372.22 g/mol (79 Br) and 374.22 g/mol (81 Br); found 372.00 g/mol (⁷⁹Br) and 374.00 g/mol (⁸¹Br). Elemental Analysis: Calculated: C, 54.71; H, 3.78; Br, 21.41; F, 5.09; N, 15.01; Found: C, 54.61; H, 3.18; Br, 21.36; F, 5.14; N, 15.01. UV-visible spectrum in 5 × 10⁻⁵ mol/L, λ (nm) (logε): λ₁, 430 (3.30); λ₂, 346 (4.42); λ₃, 244 (4.12). Infrared spectrum in KBr Tablets, cm⁻¹: $\bar{\nu}$ (C–H) Arom.: 3086w, 3071w, 3055w; $\bar{\nu}$ (C–H) Aliph.: 2992w, 2965w, 2928w; $\bar{\nu}$ (N=N), $\bar{\nu}$ (C=N) and/or $\bar{\nu}$ (C=C): 1582w, 1568w, 1555 m, 1514 s; $\bar{\nu}$ (N–N): 1407 m. 1H NMR (400 MHz, CDCl₃) δ 7.69 (w, J = 8.1 Hz, 2H), 7.59 (w, J = 8.2 Hz, 2H), 7.47 (dw, J = 8.2 Hz, 4H), 2.61 (s, 3H), 2.56 (s, 3H). 1H NMR (CDCl₃) δ 162.21 (w, J = 248.6 Hz), 152.41, 144.14, 139.55, 136.23, 135.26 (w, J = 3.1 Hz), 132.23 (+), 126.89 (+) J = 8.7 Hz, 123.74, 123.53 (+), 116.34 (+) (w, J = 23.0 Hz), 14.18 (+), 11.38 (+).

(E)-4-((4-bromophenyl)diazenyl)-1-(4-chlorophenyl)-3,5-dimethyl-1H-pyrazole (4): Yield: 76.8%, crude. Recrystallized from acetone. Melting point: 192.0–192.5° C. Molecular mass (g/mol), calculated for C₁₇H₁₄BrClN₄: 388.68 g/mol (⁷⁹Br) and 390.68 g/mol (⁸¹Br); found 388.00 g/mol (⁷⁹Br) and 390.00 g/mol (⁸¹Br). Elemental Analysis: Calculated: C, 52.40; H, 3.62; Br, 20.50; Cl, 9.10; N, 14.38; Found: C, 52.48; H, 3.60; Br, 20.42; Cl, 9.11; N, 14.39. UV-visible spectrum in CHCl₃ 5 × 10⁻⁵ mol/L, λ (nm) (logε): λ₁, 429 (3.39); λ₂, 347 (4.45); λ₃, 244 (4.22). Infrared spectrum in KBr $\bar{\nu}$, cm⁻¹ Tablets: $\bar{\nu}$ (C–H) Arom.: 3097w, 3085w, 3052w; $\bar{\nu}$ (C–H) Aliph.: 2990w, 2963w, 2928w; $\bar{\nu}$ (N=N), $\bar{\nu}$ (C=N) and/or $\bar{\nu}$ (C=C): 1591w, 1584w, 1567w, 1553 m; 1504 s; $\bar{\nu}$ (N–N): 1414 s. 1H NMR (400 MHz, CDCl₃) δ 7.69 (w, J = 7.6 Hz, 2H), 7.60 (w, J = 8.0 Hz, 2H), 7.47 (dw, 4H), 2.73 (s, 3H), 2.56 (s, 3H). NMR (101 MHz, CDCl₃) δ 152.43, 144.38, 139.57, 137.72, 136.47, 134.08, 132.28 (+), 129.60 (+), 126.10 (+), 123.85, 123.58 (+), 14.25 (+), 11.52 (+).

(E)-4-((4-bromophenyl)diazenyl)-3,5-dimethyl-1H-pyrazol-1-yl)benzotrile (5): Yield: 82.6%, crude. Recrystallized from EtOH: THF (10: 3). Melting point: 213.9–214.4° C. Molecular mass (g/mol), calculated for C₁₈H₁₄BrN₅: 379.24 g/mol (⁷⁹Br) and 381.24 g/mol (⁸¹Br); found 379.00 g/mol (⁷⁹Br) and 381.00 g/mol (⁸¹Br). Elemental

Analysis: Calculated: C, 56.86; H, 3.71; Br, 21.01; N, 18.42; Found: C, 56.82; H, 3.715; Br, 21.12; N, 18.30. UV-visible spectrum in CHCl_3 5×10^{-5} mol/L, λ (nm) (log ϵ): λ_1 , 431 (3.30); λ_2 , 347 (4.50); λ_3 , 248 (4.16). Infrared spectra of KBr Tablets, cm^{-1} : $\bar{\nu}$ (C—H) Arom.: 3109w, 3054w, 3047w; $\bar{\nu}$ (C—H) Aliph.: 2988w, 2963w, 2928w, 2854w; $\bar{\nu}$ (C \equiv N): 2226 s; $\bar{\nu}$ (N=N), $\bar{\nu}$ (C=N) and/or $\bar{\nu}$ (C=C): 1603 m, 1583 m, 1567w, 1556 m; 1514 s; $\bar{\nu}$ (N—N): 1405 s. ^1H NMR (400 MHz, CDCl_3) δ 7.81 (w, $J = 8.0$ Hz, 2H), 7.70 (dw, $J = 7.7$ Hz, 4H), 7.61 (w, $J = 8.1$ Hz, 3H), 2.74 (s, 3H), 2.57 (s, 3H). NMR (101 MHz, CDCl_3) δ 152.30, 145.13, 142.74, 139.93, 137.10, 133.45 (+), 132.34 (+), 124.55 (+), 124.23, 123.66 (+), 118.23, 14.41(+), 11.82 (+).

(E)-4-((4-bromophenyl)diazanyl)-3,5-dimethyl-1-(4-nitrophenyl)-1H-pyrazole (6): Yield: 68.6%, crude. Recrystallized from acetone. Melting point: 201.7–202.2° C. Molecular mass (g/mol), calculated for $\text{C}_{17}\text{H}_{14}\text{BrN}_5\text{O}_2$: 399.23 g/mol (^{79}Br) and 401.23 g/mol (^{81}Br); found 399.00 g/mol (^{79}Br) and 401.00 g/mol (^{81}Br). Elemental Analysis: Calculated: C, 51.02; H, 3.53; Br, 19.96; N, 17.50; O, 7.99; Found: C, 51.22; H, 3.32; Br, 19.92; N, 17.53; O, 8.01. UV-visible spectrum in CHCl_3 5×10^{-5} mol/L, λ (nm) (log ϵ): λ_1 , 442 (3.33); λ_2 , 354 (4.52); λ_3 , 244 (4.14). Infrared spectra of KBr Tablets, cm^{-1} : $\bar{\nu}$ (C—H) Arom.: 3076w; $\bar{\nu}$ (C—H) Aliph.: 2971w, 2929w, 2852w; $\bar{\nu}$ (N=N), $\bar{\nu}$ (C=N) and/or $\bar{\nu}$ (C=C): 1606w, 1592 s, 1568 m, 1561 m; 1524 s, 1507 m; $\bar{\nu}$ (N—N): 1407 s. ^1H NMR (400 MHz, CDCl_3) δ 8.39 (w, $J = 8.6$ Hz, 2H), 7.76 (w, $J = 8.6$ Hz, 2H), 7.71 (w, $J = 8.2$ Hz, 2H) $J = 8.3$ Hz, 2H), 2.77 (s, 3H), 2.58 (s, 3H). NMR (101 MHz, CDCl_3) δ 152.29, 146.51, 145.34, 144.29, 140.13, 137.28, 132.36 (+), 125.03 (+), 124.33, 124.27 (+), 123.69 (+), 14.46 (+), 11.94 (+).

(E)-4-((4-bromophenyl)diazanyl)-1-(3-chlorophenyl)-3,5-dimethyl-1H-pyrazole (7): Yield: 74.1%, crude. Recrystallized from EtOH. Melting point: 145.7–146.2° C. Molecular mass (g/mol), calculated for $\text{C}_{17}\text{H}_{14}\text{BrClN}_4$: 388.68 g/mol (^{79}Br) and 390.68 g/mol (^{81}Br); found 388.00 g/mol (^{79}Br) and 390.00 g/mol (^{81}Br). Elemental Analysis: Calculated: C, 52.40; H, 3.62; Br, 20.50; Cl, 9.10; N, 14.38; Found: C, 51.18; H, 3.36; Br, 19.80; N, 17.59; O, 8.07. UV-visible spectrum in CHCl_3 5×10^{-5} mol/L, λ (nm) (log ϵ): λ_1 , 430 (3.31); λ_2 , 346 (4.44); λ_3 , 244 (4.16). Infrared spectra of KBr Tablets, cm^{-1} : $\bar{\nu}$ (C—H) Arom.: 3081w, 3063w; $\bar{\nu}$ (C—H) Aliph.: 2986w, 2962w, 2921w, 2853w; $\bar{\nu}$ (N=N), $\bar{\nu}$ (C=N) and/or $\bar{\nu}$ (C=C): 1594 s, 1586w, 1571 s, 1557w, 1541w, 1504 s; $\bar{\nu}$ (N—N): 1409 s. ^1H NMR (400 MHz, CDCl_3) δ 7.69 (w, $J = 7.7$ Hz, 2H), 7.59 (w, $J = 7.9$ Hz, 2H), 7.55 (s, 1H), 7.47–7.66 (m, 3H), 2.67 (s, 3H), 2.56 (s, 3H). NMR (101 MHz, CDCl_3) δ 152.39, 144.41, 140.21, 139.72, 136.52, 135.15, 132.26 (+), 130.33 (+), 128.32 (+), 125.13 (+), 123.88, 123.58 (+), 122.80 (+), 14.28 (+), 11.54 (+).

(E)-4-((4-bromophenyl)diazanyl)-3,5-dimethyl-1-(3-nitrophenyl)-1H-pyrazole (8): Yield: 85.0%, crude. Recrystallized from EtOH/THF (5:3). Melting point: 190.3–191.3° C. Molecular mass (g/mol), calculated for $\text{C}_{17}\text{H}_{14}\text{BrN}_5\text{O}_2$: 399.23 g/mol (^{79}Br) and 401.23 g/mol (^{81}Br); found 399.00 g/mol (^{79}Br) and 401.00 g/mol (^{81}Br). Elemental Analysis: Calculated: C, 51.02; H, 3.53; Br, 19.96; N, 17.50; O, 7.99; Found: C, 51.18; H, 3.34; Br, 19.94; N, 17.50; O, 8.04. UV-visible spectrum in CHCl_3

5×10^{-5} mol/L, λ (nm) (log ϵ): λ_1 , 431 (3.30); λ_2 , 342 (4.46); λ_3 , 241 (4.31). Infrared spectra of KBr Tablets, cm^{-1} : $\bar{\nu}$ (C—H) Arom.: 3103w, 3075w; $\bar{\nu}$ (C—H) Aliph.: 2965w, 2924w; $\bar{\nu}$ (N=N), $\bar{\nu}$ (C=N) and/or $\bar{\nu}$ (C=C): 1583w, 1560w, 1540 s, 1502 s; $\bar{\nu}$ (N—N): 1415 m. ^1H NMR (400 MHz, CDCl_3) δ 8.43 (s, 1H), 8.27 (w, $J = 8.2$ Hz, 1H), 7.91 (w, $J = 8.0$ Hz, 1H), 7.71 (w, $J = 8.0$ Hz, 3H), 7.61 (w, $J = 8.1$ Hz, 2H), 2.74 (s, 3H), 2.58 (s, 3H). NMR (101 MHz, CDCl_3) δ 152.30, 148.80, 145.02, 140.27, 139.86, 136.88, 132.33 (+), 130.39 (+), 130.05 (+), 124.19, 123.66 (+), 122.52 (+), 119.40 (+), 14.35 (+), 11.65 (+).

(E)-4-((4-bromophenyl)diazanyl)-3,5-dimethyl-1-(2-nitrophenyl)-1H-pyrazole (9): Yield: 83.4%, crude. Recrystallized from EtOH. Melting point: 162.3–162.7° C. Molecular mass (g/mol), calculated for $\text{C}_{17}\text{H}_{14}\text{BrN}_5\text{O}_2$: 399.23 g/mol (^{79}Br) and 401.23 g/mol (^{81}Br); found 399.00 g/mol (^{79}Br) and 401.00 g/mol (^{81}Br). Elemental Analysis: Calculated: C, 52.40; H, 3.62; Br, 20.50; Cl, 9.10; N, 14.38; Found: C, 52.38; H, 3.14; Br, 20.40; Cl, 9.18; N, 14.30. UV-visible spectrum in CHCl_3 5×10^{-5} mol/L, λ (nm) (log ϵ): λ_1 , 439 (3.25); λ_2 , 342 (4.40); λ_3 , 244 (4.20). Infrared spectrum in KBr Tablets, cm^{-1} : $\bar{\nu}$ (C—H) Arom.: 3088w; $\bar{\nu}$ (C—H) Aliph.: 2989w, 2959w, 2922w, 2866w; $\bar{\nu}$ (N=N), $\bar{\nu}$ (C=N) and/or $\bar{\nu}$ (C=C): 1611 m, 1584 m, 1571w, 1557 m; 1528 s, 1509 m; $\bar{\nu}$ (N—N): 1418 s. ^1H NMR Spectrum (400 MHz, CDCl_3) δ 8.07 (w, $J = 8.1$ Hz, 1H), 7.79–7.52 (m, 7H), 2.53 (s, 3H), 2.52 (s, 3H). NMR (101 MHz, CDCl_3) δ 152.32, 146.26, 145.47, 140.84, 136.05, 133.68 (+), 132.41, 132.23 (+), 130.21 (+), 129.63 (+), 125.59 (+), 123.93, 123.59 (+), 14.12 (+), 10.74 (+).

(E)-4-((4-bromophenyl)diazanyl)-1-(2,4-dinitrophenyl)-3,5-dimethyl-1H-pyrazole × EtOH (10): Yield: 93.5%, crude. Recrystallized from EtOH. Melting point: 139.5–140.0° C. Molecular mass (g/mol), calculated for $\text{C}_{17}\text{H}_{13}\text{BrN}_6\text{O}_4$: 444.23 g/mol (^{79}Br) and 446.23 g/mol (^{81}Br); found 444.00 g/mol (^{79}Br) and 446.00 g/mol (^{81}Br). Elemental Analysis: Calculated: C, 45.86; H, 2.94; Br, 17.95; N, 18.88; O, 14.37; Found: C, 45.80; H, 3.00; Br, 17.92; N, 18.90; O, 14.38. UV-visible spectrum in CHCl_3 5×10^{-5} mol/L, λ (nm) (log ϵ): λ_1 , 442 (3.41); λ_2 , 340 (4.38); λ_3 , 244 (4.27). Infrared spectrum in KBr Tablets, cm^{-1} : $\bar{\nu}$ (C—H) Arom.: 3125w, 3101 m, 3008w; $\bar{\nu}$ (C—H) Aliph.: 2969w, 2928w; $\bar{\nu}$ (N=N), $\bar{\nu}$ (C=N) and/or $\bar{\nu}$ (C=C): 1609 s, 1565 m, 1542 s, 1533 s, 1509 m; $\bar{\nu}$ (N—N): 1410 s. ^1H NMR (400 MHz, CDCl_3) δ 8.87 (s, 1H), 8.60 (w, $J = 8.7$ Hz, 1H), 7.79 (w, 2.60 (s, 3H), 2.53 (s, 3H), 1.23 (t, $J = 7.0$ Hz, 2H), 7.61 (w, 3H). NMR (101 MHz, CDCl_3) δ 152.16, 147.09, 147.05, 145.85, 140.52, 137.20, 136.84, 132.37 (+), 129.92 (+), 127.79 (+), 124.58, 123.73 (+), 121.40, 58.57 (-), 18.55 (+), 14.17 (+), 10.94 (+).

(E)-4-((4-bromophenyl)diazanyl)-3,5-dimethyl-1-(perfluorophenyl)-1H-pyrazole (11): Yield: 86.6%, crude. Recrystallized from EtOH. Melting point: 153.5–154.4° C. Molecular mass (g/mol), calculated for $\text{C}_{17}\text{H}_{10}\text{BrF}_5\text{N}_4$: 444.18 g/mol (^{79}Br) and 446.18 g/mol (^{81}Br); found 444.00 g/mol (^{79}Br) and 446.00 g/mol (^{81}Br). Elemental Analysis: Calculated: C, 45.87; H, 2.26; Br, 17.95; F, 21.34; N, 12.59; Found: C, 45.80; H, 2.20; Br, 17.93; F, 21.47; N, 12.61. UV-visible spectrum in CHCl_3 5×10^{-5} mol/L, λ (nm) (log ϵ): λ_1 , 431 (3.21); λ_2 , 339 (4.39); λ_3 , 244 (4.06). Infrared spectra of KBr Tablets,

cm-1: $\bar{\nu}$ (C–H) Arom.: 3080w; $\bar{\nu}$ (C–H) Aliph.: 2990w, 2967w, 2926w; $\bar{\nu}$ (N=N), $\bar{\nu}$ (C=N) and/or $\bar{\nu}$ (C=C): 1583w, 1571 m, 1536 s, 1510 m; $\bar{\nu}$ (N–N): 1409 s. ^1H NMR (400 MHz, CDCl_3) δ 7.67 (w, $J = 7.8$ Hz, 2H), 7.58 (w, $J = 8.2$ Hz, 2H), 2.54 (s, 3H), 2.49 (s, 3H). NMR (101 MHz, CDCl_3) δ 152.20, 146.64, 145.34 (dt, $J = 16.2, 4.1$ Hz), 143.92–143.11 (m), 142.80 (dt, $J = 15.5, 3.7$ Hz), 141.98, 141.83–140.84 (m), 139.65–138.96 (m), 137.12–136.59 (m), 135.97, 132.34 (+), 124.38, 123.67 (+), 114.35 (tw, $J = 14.9, 4.5$ Hz), 14.20 (+), 10.24 (+).

2.6. Computational details

The Gaussian 03 computational package (Frisch et al., n.d.) was used to perform ground-state geometry optimization calculations employing Becke's three-parameter hybrid exchange functional and the Lee Yang Parr nonlocal correlation functional CAMB3LYP (Yanai et al., 2004). The LANL2DZ basis set (Hay and Wadt, 1985) with an effective core potential for I and a 6-31G* basis set was used for C, O, N, Br, Cl, F and H atoms (Rassolov et al., 2001) were employed. The vibrational frequencies were calculated in order to corroborate the arrival to a minimum point in the potential energy surface. Time-dependent density functional theory (TDDFT) calculations were also performed using this methodology, and the first 200 singlet excited states were calculated. Calculations by the first-principles method were used to obtain accurate excitation energies and oscillator strengths. Solvent effect was considered using the polarizable continuum model (PCM) using methanol as solvent (Cossi et al., 2002).

3. Biological assays

3.1. In vitro anti-tumor assays.

The NCI's in vitro anti-tumor screening protocol consists of 60 human tumor cell lines against which compounds 4–11 were tested with 3–4,5-dimethylthiazol-2-yl-2,5-diphenyl-tetrazolium bromide (MTT) assay (Shoemaker, 2006). Cancer cells were treated with 1 μM of test compounds. Dimethyl sulfoxide (DMSO) was used as a vehicle control and six wells were prepared for each compound. After treatment, the supernatant was carefully aspirated and 150 μl of DMSO was added to each well. The absorbance was measured at 590 nm.

3.2. Primary cultures of venular endothelial cells

Microvascular endothelial cells were isolated from the mesenteric arterial beds of male Sprague-Dawley rats (200–230 g). Rats were bred and maintained in the Research Animal Facility of the Pontificia Universidad Católica de Chile and all studies were approved by the Institutional Bioethics Committee. Primary cultures of endothelial cells of post-capillary venules were prepared using a similar procedure to that described previously by Lillo et al. (Lillo et al., 2018). Briefly, rats were anaesthetized with xilazine and ketamine (10 and 90 mg/kg, i.p., respectively) and the mesenteric arterial bed was isolated as described by Gaete et al. (Gaete et al., 2012). Isolated mesenteric beds were perfused with a sterile Tyrode buffer

solution containing a mixture of antibiotics and antimycotics (Anti-Anti solution; Thermo Fisher Scientific, Waltham, MA, USA) to remove the blood from the vessels. Several post-capillary venules were isolated from the mesenteric tissue, and thus, incubated in a physiologic saline solution containing 0.2% collagenase type I and 0.1% bovine serum albumin (BSA) for 1 h at 37 °C, diluted (5-fold) with M–199 medium, and centrifuged. Pelleted cells were resuspended in M–199 medium, centrifuged, and resuspended again in M–199 medium containing 20 mg/ml endothelial cell growth supplement from bovine pituitary and 20% fetal bovine serum. Then, cells were seeded onto 35-mm culture dishes or onto sterile glass coverslips. Four hours later, nonadherent cells were removed, and the remaining adherent endothelial cells were kept at 37 °C in a 5% CO_2 –95% air atmosphere at nearly 100% relative humidity. Experiments were performed with post-capillary venule endothelial cells at 70 to 80% of confluence (~2 days of culture) in which the culture media was replaced by a MOPS-buffered Tyrode saline solution (pH 7.4).

4. Results and discussion

4.1. Synthesis

The β -diketohydrazone, 3-(2-(4-bromophenyl)hydrazinylidene)pentane-2,4-dione (Bustos et al., 2009, 2007; Yao, 1964), was used as a precursor to obtain a family of pyrazole derivatives. The general name for the obtained family of pyrazoles is (*E*)-(3,5-dimethyl-1-(4-**R**-phenyl)-(4-Br-phenyldiazanyl)-1H-pyrazoles pyrazole (with R = 4- OCH_3 (1), 4-H (2), 4-F (3), 4-Cl (4), 4-CN (5), 4- NO_2 (6), 3-Cl (7), 3- NO_2 (8), 2- NO_2 (9), 2,4-(NO_2)₂ (10), C_6F_5 - NHNH_2 (11)).

The mechanism of the pyrazoles obtention is shown in the SI as an electronic movement. The reactions were carried out under reflux for 36 h in ethanol solution, in stoichiometric ratio 1:1, using hydrochloric acid as catalyst. Compounds 1–11 were recrystallized from solvents such as ethanol, acetone, and ethanol/THF mixtures.

The characterization was performed using analytical techniques (MP, MM and EA) shown in Table S2 in the SI; where the most relevant point is related with the determination of the calculated molecular mass of pyrazole isotopic molecules, which contain isotopes of ^{79}Br and ^{81}Br . This molecular mass was obtained by mass spectrometry in solution, whose chromatograms are shown in a relation m/z near 1:1, see Fig. S1–S11 in SI, which is consistent with the percentages of abundance of each isotope of bromine in nature. The spectroscopic characterization was performed using UV–visible, IR, ^1H NMR, ^{13}C NMR and two-dimensional NMR experiments (HMBC and HMQC).

4.2. X-Ray diffraction

Suitable crystals for X-Ray diffraction were obtained for molecule 9, an ortep view is shown in Fig. S3. In the SI are provided the crystallographic and refinement data. In Table S3 are shown the selected bond distances, angles, torsion angles and DFT results of the geometrical parameters of the whole family of compounds. The discussion of the DFT data is provided in the computational simulations section (*vide infra*). As shown in

Table 1 UV-vis absorption bands, λ nm ($\log\epsilon$), of the pyrazoles (1–11), recorded in CHCl_3 solution.

Comp.	$\lambda_1(\log\epsilon)$	λ_{1Th}	f				%	$\lambda_1(\log\epsilon)$	λ_{1Th}	f				%	$\lambda_1(\log\epsilon)$	λ_{1Th}	f				%
1	439 (3.32)	456	0.088	H	→	L	96	347 (4.39)	337	0.266	H-2	→	L	95	240 (4.21)	240	0.01	H-9	→	L	25
																		H-2	→	L + 1	33
																		H-2	→	L + 2	21
2	431 (3.32)	457	0.042	H	→	L	99	344 (4.38)	316	0.035	H-2	→	L	97	240 (4.14)	248	0.1	H-6	→	L	18
																		H	→	L + 2	75
3	430 (3.30)	457	0.035	H	→	L	99	346 (4.42)	377	1.335	H-1	→	L	99	244 (4.12)	259	0.24	H	→	L + 2	86
4	429 (3.39)	458	0.045	H	→	L	99	347 (4.45)	378	1.4 16	H-1	→	L	98	244 (4.22)	270	0.22	H	→	L + 1	74
																		H-4	→	L	21
5	431 (3.30)	460	0.047	H	→	L	99	347 (4.50)	381	1.567	H-1	→	L	95	248 (4.16)	268	0.36	H-2	→	L + 1	84
6	442 (3.33)	464	0.056	H	→	L	97	354 (4.52)	360	0.726	H	→	L + 1	92	244 (4.14)	246	0.23	H	→	L + 3	75
7	430 (3.31)	458	0.067	H	→	L	99	346 (4.44)	376	1.395	H-1	→	L	98	244 (4.16)	247	0.14	H	→	L + 2	79
																		H-5	→	L	16
8	431 (3.30)	418	0.030	H	→	L	99	342 (4.46)	374	1.397	H	→	L + 1	99	241 (4.31)	246	0.15	H	→	L + 3	80
																		H-4	→	L + 1	13
9	439 (3.25)	443	0.137	H	→	L	98	342 (4.40)	361	0.156	H-2	→	L	77	244 (4.20)	245	0.15	H-4	→	L + 1	14
											H	→	L + 1	19	H	→		L + 4	79		
10	442 (3.41)	420	0.133	H	→	L + 1	94	340 (4.38)	358	0.699	H-1	→	L + 2	80	244 (4.27)	259	0.11	H-10	→	L	23
											H-3	→	L	11	H-8	→		L	19		
11	431 (3.21)	456	0.078	H	→	L	99	339 (4.39)	456	1.337	H-1	→	L	98	244 (4.06)	244	0.22	H-7	→	L + 1	28
																		H-6	→	L	19
																		H-10	→	L + 1	39
																		H	→	L	29

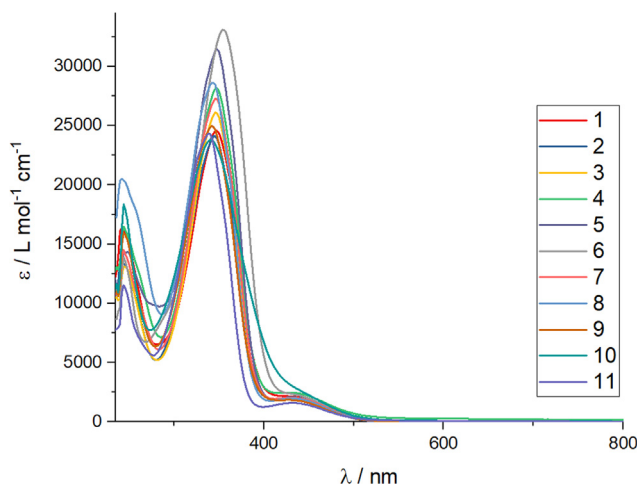


Fig. 3 UV-vis in CH₃Cl for 1–11.

Fig. S12, the azo bridge adopts *E* conformation. In case of **9**, as shown in the ortep view, due to the steric hindrance generated by the –NO₂ group, the structure does not show planarity. As it has been previously reported for these types of compounds, in the crystallographic structure of the complete supramolecular structure, there are no conventional intermolecular and intramolecular hydrogen bonds. When the substituent is located at a different position than ortho, the whole structure shows higher degree of planarity, *vide infra* in the theoretical study.

4.3. Spectroscopic study

4.3.1. UV-visible pyrazole spectroscopy

A summary of the absorption bands found in the UV-Vis spectra for **1–11** are shown in Table 1. All superimposed UV-Vis spectra are shown in Fig. 3 and the corresponding spectrum for each compound is shown in the SI (Figs. S13–S23). The theoretical UV-Vis spectra are also shown in the SI (Figs. S24–S34). Each molecule has a C=N=N–C fragment, which contains a –N=N– double bond. This double bond is conjugated on one end to the phenyl group and, on the other end to the pyrazole ring. The spectra recorded in chloroform show three absorption bands λ_1 , λ_2 and λ_3 centered on the ranges 430–442 nm, log ϵ : 3.25–3.39; 339–354 nm, log ϵ : 4.38–4.50 and 240–248 nm, log ϵ : 4.06–4.27, respectively. These bands are attributed to $\pi \rightarrow \pi^*$ transitions centered on different regions of the molecules. The first transition is centered on the fragment C=N=N–C that connects both pyrazole rings; the second is centered on the pyrazole ring and the third is due to internal transitions of the substituted aromatic rings.

4.3.2. Infrared Spectroscopy of Pyrazoles

The most important IR bands shown in the solid state KBr pellets are summarized in the SI, see Table S4 and Figs. S46–S56. The full set of spectra can be found in the SI. In general, the weak $\nu(\text{CH})$ stretchings of the aromatic protons are shown in the spectra over 3000 cm⁻¹ and, immediately below 3000 cm⁻¹, the weak aliphatic stretches $\nu(\text{C–H})$ emerge from the system 3,5-dimethylpyrazole, also involving the *p*-CH₃

group for compound **1**. Additionally, in the region of 1611–1502 cm⁻¹ are observed several bands attributed to the stretches $\nu(\text{C=N})$, $\nu(\text{N=N})$ or $\nu(\text{C=C})$. The $\nu(\text{N–N})$ stretches of the pyrazole ring are located around 1418–1405 cm⁻¹. The influence of the substituent over the IR signals is small or neglectable, thus as can be observed in Table S4, there is no clear tendency of the most important signals as the substituent is changed. Finally, in case of **5** with CN as substituent, the characteristic absorption for the $\nu(\text{C}\equiv\text{N})$ is observed at 2226 cm⁻¹.

4.3.3. Nuclear Magnetic Resonance Spectroscopy

All ¹H NMR spectra of the pyrazoles (**1–11**) were recorded in CDCl₃. The full set of spectra and a summary of the signals can be found in each ¹H NMR spectrum in the SI, see Figs. S46–S56.

According to the obtained data, the most important features of the ¹H NMR spectra of the pyrazoles are three. First, variable multiplicity resonances are observed in the range 8.87–7.19 ppm, due to the aromatic protons. Second, a singlet located in the range 2.53–2.58 ppm, due to the protons ¹C–¹H of the methyl group are observed. Third, a singlet in the range 2.49–2.77 ppm attributed to the ⁵C–⁵H protons of the second methyl group are observed. Furthermore, compound **1** shows a resonance at 3.87 ppm due to the methoxy group of the *p*-OCH₃ substituent. Although the substituent over the phenyl pyrazole ring is changed, there are small chemical shifts measured in the ¹H NMR spectra, see Table S5. This effect is attributed to the non-planarity observed in the structure; therefore, the delocalization of the ring is not over the whole structure. In this sense, the phenyl ring over the pyrazole acts as one region and the azophenylpyrazole act as a separate region in the molecule.

The signals found in the ¹³C NMR spectra are summarized in Table S6 in the SI and full set of spectra are also found in the SI, see Figs. S57–S67. Compounds **1–11** show the methyl signals of the pyrazole ring, which follow the same trend as the proton signal, showing almost static positions. Additionally, **1** and **5** show the typical resonances of the R groups located on the aromatic ring. Furthermore, **10** shows the resonances for the ethanol crystallization molecule.

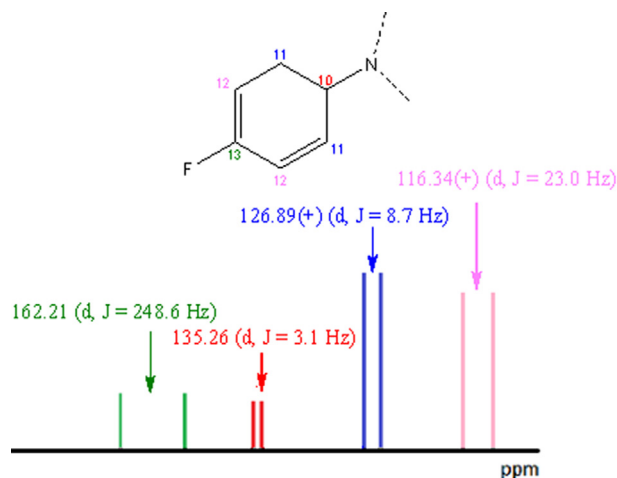


Fig. 4 C-F coupling observed in the ¹³C NMR spectrum of the 4-F-C₆H₄-N- ring of (IV).

Those compounds that contain fluorine atoms (**3** and **11**) show typical resonances of the C-F coupling.

In general, the signals of the aromatic carbons attached to the fluorine atom of the ring 4-F-C₆H₄- in case of **3**, appear as a doublet centered at 162.21 ppm ($J_{\text{gem}} = 248.6$ Hz), 116.34 ppm ($J_{\text{o}} = 23.0$ Hz), 126.89 ppm ($J_{\text{m}} = 8.7$ Hz) and 135.26 ppm ($J_{\text{p}} = 3.1$ Hz), respectively, see Fig. 4. The intensities of these signals allow to determine the tertiary or quaternary nature of each type of carbon and, together, with the progressive decrease of the values of J ($J_{\text{gem}} > J_{\text{o}} > J_{\text{m}} > J_{\text{p}}$), which is related to the intensity of the C-F coupling. In this way, the carbons of this ring could be unequivocally assigned. A detail of what is observed in the ¹³C NMR spectra of (**3** and **11**) is shown in the SI.

On the other hand, a more complex situation is observed in case of **11**. The signals of the quaternary carbons of the C₅F₅-ring of **11** should appear as a complex multiplet due to geminal C-F coupling and long distance. In Fig. S67 of the supplementary information, an outline of the resonances of the C₅F₅-ring carbons found in the ¹³C-spectrum of **11** is shown. As observed in the spectrum, these measured signals are mixed with the resonances of the phenyldiazanyl group carbons. In fact, this spectrum shows three doublet pairs of multiplets, d_{m} , located at 144.10 ppm ($J_{\text{gem}} = 256.09$ Hz), 142.36 ppm ($J_{\text{gem}} = 259.03$ Hz) and 138.11 ppm ($J_{\text{gem}} = 259.17$ Hz). The J values are in agreement with the C-F geminal couplings and similar to the J_{gem} value found in the signal located at 162.21 ($J_{\text{gem}} = 248.6$) of **3**. Finally, carbon C₁₀, does not show a geminal C-F coupling and appear as triplet of doublets, td , at 114.35 ppm ($J_{\text{o}} = 14.9$ Hz, $J_{\text{m}} = 4.5$ Hz), this observed td symmetry is probably due to $J_{\text{m}} \sim J_{\text{p}}$.

The DEPT-135 spectrum was measured for all the compounds. In Table S7 in the SI, all the signals of the carbons are assigned as CH₃- (+), CH₂- (-) and CH- (+) are tabulated. In general, the DEPT-135 spectra of the compounds **1**–**11** show positive resonances of the carbon atoms. However,

in case of **11**, two additional signals are observed (18.35(+) ppm and 58.36(-)), which are assigned to crystallization molecules of ethanol. Even though the sample was dried for several days under high vacuum at 90°C, those signals remain appearing in the DEPT-135 spectra. The full set of spectra can be found in the SI, Fig. S69-S79.

The HMBC spectra show all the expected interactions, where the most outstanding characteristic is the appearance of an invariable pattern of interactions of the methyl groups with the immediately neighboring carbons, which reveal the presence of the pyrazole ring. This NMR behavior is found in every studied sample, as shown in the SI. A deeper analysis is performed for **1** in Fig. S80, where the mentioned interactions are emphasized, and in addition, the interaction of the substituent 4-CH₃O- with the carbon ¹³C of the aromatic ring is also observed. Furthermore, the two-dimensional HMQC spectra shows a characteristic pattern of interactions, as shown in Fig. S81 for **1**. The full set of HMQC and HMBC spectra can be found in the SI, see Figs. S80-S103.

5. In vitro anti-tumor assays

Primary in vitro one dose anticancer assay was carried out over the full NCI 60 cell panel. This panel includes 60 cell lines, which can be classified as leukemia, melanoma, non-small lung cancer, colon cancer, brain cancer, breast cancer, ovarian cancer, kidney cancer and prostate cancer cell lines. The NCI, USA, protocol was used for all the experiments. One-dose data are reported for all studied compounds as a mean graph of the percent growth of treated cells, see Fig. 5 and SI, see Figs. S104-S110. In this sense, the obtained value for the one-dose assay is expressed as growth relative to the control (cells not treated with any drug), and also relative to the number of cells present when the experiment is started. Thus, growth inhibition can be detected and related to values, which are expressed between 0 and 100. On the other hand, lethality

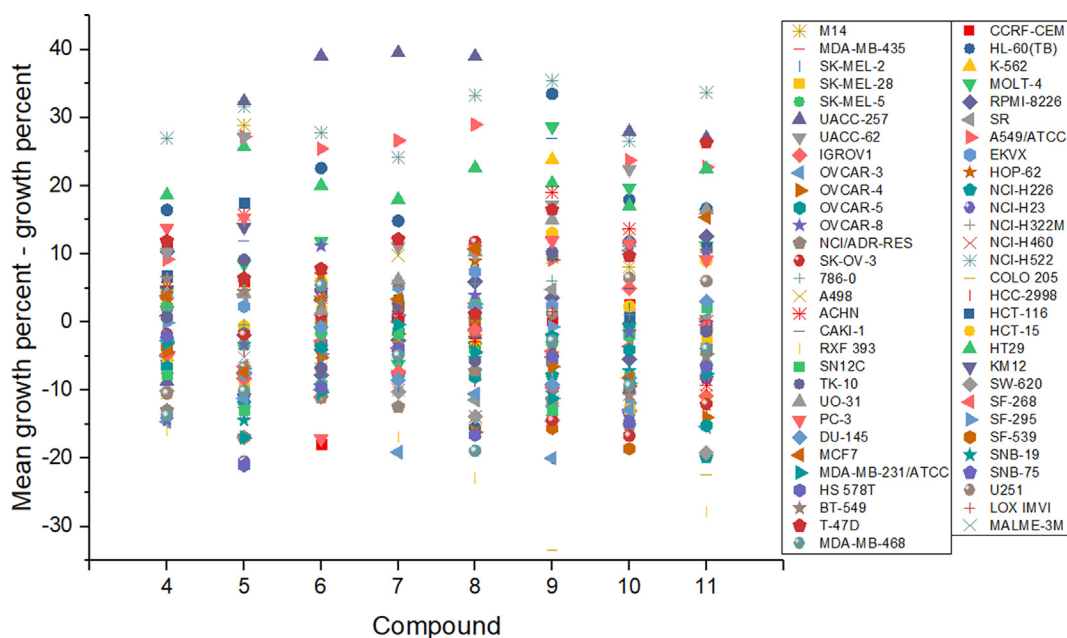


Fig. 5 Percentage of cell growth of NCI 60 cancer cell lines displayed by compounds **4**–**11**.

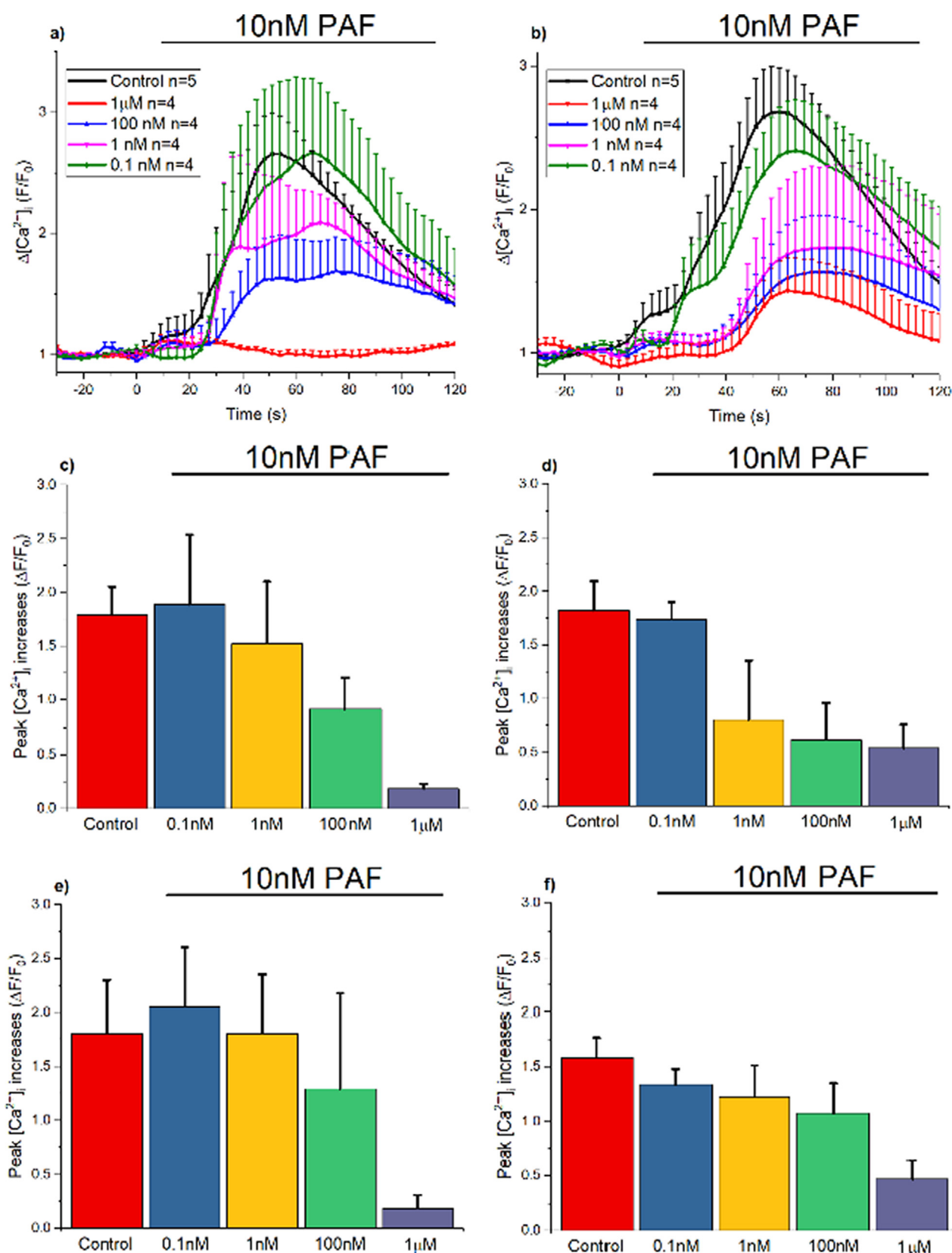


Fig. 6 Concentration-dependent inhibition of PAF-activated Ca^{2+} signaling by pyrazole analogues. The effect of different concentrations (0.1 nM to 1000 nM) of compounds **6**, **8**, **9** and **10** on the response to PAF was evaluated in primary cultures of venular endothelial cells. A and B, Time course of the increase in $[Ca^{2+}]_i$ induced by PAF in control conditions and after the treatment for 15 min with compounds **6** (A) and **8** (B). Note that compound **6** strongly inhibited the PAF-activated Ca^{2+} signal and compound **8** evoked a delay in the beginning of the response. Horizontal bars indicate the stimulation period. C to F, Analysis of the maximum increase in $[Ca^{2+}]_i$ observed in response to PAF before and after application of a concentration range from 0.1 nM to 1000 nM of compounds **6** (C), **8** (D), **10** (E) or **9** (F) for 15 min.

is related to negative values. Therefore, as an example, an obtained value of 100 is related to no growth inhibition. On the other hand, a value of 10 means 90% growth inhibition, whereas a value of 0 is related to no net growth during the experiment time. A value of -50 would mean 50% lethality and finally a value of -100 means all cells are dead. All the results related to the one-dose measurements can be found in the SI. In Figure 5, are shown the one-dose result value obtained for all compounds. As can be observed, all the studied compounds show certain degree of growth inhibition against different cell lines. Furthermore, all compounds also show certain degree of lethality against different cell lines. The median obtained value, shows that most commonly, the studied compounds show more lethality than growth inhibition. Although the studied compounds show certain degree of similarity, there is no correlation between the activity and the structure. It is important to notice that highly selective growth inhibition activity was observed against UACC-257 (melanoma cell line), NCI-H522 (lung cancer cell line), A549/ATCC (lung cancer cell line) and HT29 (colon cancer cell line). These values can be related with certain degree of tumor-tissue-type (disease-oriented) selectivity. It has been previously reported an over-expression of growth factor kinases, such as EGFR, HER2/Neu, PDGFR, VEGFR, among others (Sikorski and Trzybiński, 2013). Furthermore, lethality is observed against OVCAR-3 and RXF-393. Over those cell lines, every compound shows activity, however, it is only modest.

6. Anti-proinflammatory activity

We focused the $[Ca^{2+}]_i$ study on the analysis of four selected compounds (**6**, **8**, **9** and **10**), which hold nitro ($-NO_2$) substituent in their structures. In control conditions, stimulation of endothelial cells with PAF initiated a rapid increase in $[Ca^{2+}]_i$ that peaked at 50 to 60 s and gradually declined thereafter (see Fig. 6, a and b). Cells were treated with a concentration range from 0.1 nM to 1 μ M of each pyrazole analogues and only one concentration was tested in each experiment. Application of either compound **6**, **8**, **9** or **10** during 15 min before the stimulation, inhibited the PAF-induced Ca^{2+} signal in a concentration-dependent manner. Furthermore, compounds **6** and **10** completely abolished the response at 1 μ M (Fig. 6, c and e). On the other hand, compounds **8** and **9** only attenuated the increase in $[Ca^{2+}]_i$ at this concentration (Fig. 6, d and f). Interestingly, although the inhibition observed in the presence of compound **8** was partial, the treatment with this pyrazole analogue evoked a clear delay in the onset of the PAF-elicited Ca^{2+} signal (Fig. 6, b and d). These results are consistent with the previously described anti-inflammatory properties of pyrazoles and indicate that the pyrazole family characterized in this study, in particular compounds **6** and **10**, are potent blockers of the PAF-initiated Ca^{2+} signaling that mediates the hyperpermeability typically observed during the development of inflammation (Tiruppathi et al., 2002). Although the inhibition of store-operated Ca^{2+} channels or T-type Ca^{2+} channels may contribute to the blockade of the response to PAF, the mechanism involved in this process remain to be determined.

7. Theoretical study

Finally, DFT and TDDFT calculations were performed over the whole family of compounds to model the FTIR and the UV-Vis absorption spectra of each compound, with the aim to get deeper insights of the observed measured values. All the cartesian coordinates of the family of studied compounds are in the SI. The RMSD value for **9** (XRay structure and DFT structure) is of 3.8, showing the good agreement between the experimental and theoretical results. The bromophenyl-diazenyl fragment of the molecule is planar, whereas the phenyl fragment shows a torsion angle between 27 and 67°. In the SI can be found all the superimposed FTIR of each compound with the DFT obtained spectrum. A good agreement between the experimental and the theoretical spectra is observed. Therefore, the observed signals in the FTIR spectrum can be assigned to the corresponding vibration, as previously discussed, see Table 1. Furthermore, the UV-Vis transitions were modeled by means of TDDFT calculations. All the FMO involved in the calculated transitions can be found in the SI, see Fig. S111-S115. The first calculated excitation corresponds to a H-L transition and the second calculated transition, which is the most intense transition, involves the electron promotion from HOMO-1 or HOMO-2 to the LUMO, which can be assigned as a $n-\pi^*$ transition. Finally, the third observed band involves low laying MOs and higher unoccupied MOs and corresponds to a $\pi-\pi^*$ transition. Finally, the reactivity indexes were measured for the whole family of studied compounds, see Table S8. As shown, the most reactive compounds correspond to **6**, **8**, **9** and **10**. Specifically, it is worthy to highlight that **6** and **10** show the lowest value of chemical potential and the largest value of electrophilicity, which correlates with the fact, that these compounds are much more active in the biological trials. All the geometrical coordinates are included in the SI.

8. Conclusions

It was possible to synthesize, with good yields, a family of 11 pyrazoles, of the type of (*E*)-4-((4-bromophenyl)diazenyl)-3,5-dimethyl-1-*R*-phenyl-1H-pyrazole, by reacting 3-(2-(4-bromophenyl)hydrazinylidene)pentane-2,4-dione with substituted arylhydrazines. The compounds have geometry *E* with respect to the double bond $-N=N-$ linking the pyrazole ring with the p-bromophenyl ring, as was shown in the X-Ray diffraction and DFT calculations.

In the herein report, we provided novel pyrazol derivatives that successfully displayed a promising anti-proinflammatory activity, *i.e.* **6**, **8**, **9** and **10**, which was revealed by the blockage of the intracellular $[Ca^{2+}]_i$ increase after PAF treatment, in terms of the $[Ca^{2+}]_i$ channel inhibition. The obtained intracellular $[Ca^{2+}]_i$ signal results are consistent with the previously described anti-inflammatory properties of pyrazoles and indicate that the pyrazole family characterized in this study, in particular compounds **6** and **10**, are potent blockers of the PAF-initiated Ca^{2+} signaling that mediates the hyperpermeability typically observed during the development of inflammation (Tiruppathi et al., 2002). Although the inhibition of store-operated Ca^{2+} channels or T-type Ca^{2+} channels may con-

tribute to the blockade of the response to PAF, the mechanism involved in this process remain to be determined.

Furthermore, the pyrazoles were studied in the NCI-60 cancer cell line panel, showing moderate to good activity against different cell lines.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.arabjc.2020.05.042>.

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