**SUPPLEMENTARY MATERIALS**

**A comprehensive assessment of phytochemicals from *Phyla nodiflora* (L.) Greene as a potential enzyme inhibitor, and their biological potential: An *in-silico*, *in-vivo*, and *in-vitro* approach**

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**Abstract:**

**Abstract:**

This work explored *Phyla nodiflora*(L.) Greene as a potential source of the bioactive medicinal agent. In this aspect, methanol (PN-M) and dichloromethane (PN-D) extracts were prepared from the whole plant and evaluated for phytochemical composition (total bioactive contents, UHPLC-MS analysis, and HPLC-PDA polyphenolic quantification), biological (antioxidant and enzyme inhibition) potential and *in-vivo* toxicity. The PN-M was found to contain higher phenolic (26.08 mg GAE/g extract) and flavonoid (50.25 mg QE/g extract) contents which might correlate to the higher radical scavenging (DPPH: 52.94 mg TE/g extract; ABTS: 72.11 mg TE/g extract) and reducing power (FRAP: 71.96 mg TE/g extract; CUPRAC: 142.65 mg TE/g extract) antioxidant potential, as well as AChE (4.33 mg GALAE/g extract), tyrosinase (125.36 mg KAE/g extract), and amylase (1.86 mmol ACAE/g extract) inhibition activity of this extract. In contrast, the PN-D extract was found to be most active for phosphomolybdenum (1.30 mg TE/ g extract) and metal chelation (54.84 mg EDTAE/g extract) assays in addition to BChE (4.70 mg GALAE/g extract) and glucosidase (0.62 mmol ACAE/g extract) enzyme inhibition activity. The PN-M extract on UHPLC-MS analysis revealed the tentative identification of 24 different secondary metabolites, most of which belonged to the flavonoid, glycoside, and terpenoid classes of phytochemicals. The polyphenolic composition of the extracts was appraised by HPLC-PDA. Seven phenolic compounds were identified in the extracts. PN-M was found to be rich in catechin (0.25 µg/extract) and 3-OH benzoic acid (0.64 µg/extract), while PN-D contained epicatechin (0.30 µg/extract), 3-OH-4-MeO benzaldehyde (0.21 µg/extract), and 2,3-Di-Meo benzoic acid (0.97 µg/extract) in higher amounts. The methanol extract was found to be non-toxic even at higher doses. Furthermore, the relationship between the phytochemicals and the tested enzymes was highlighted by molecular docking studies. In sum, this research showed that the studied extracts were effective as enzyme inhibitors and antioxidants, suggesting it would be worth investigating in more depth for further advanced studies to explore its pharmacological properties.

**Keywords:** *Phyla nodiflora*; phytochemicals; UHPLC-MS; HPLC-PDA; antioxidant; enzyme inhibition; toxicity

1. ***Phytochemical Composition***
	1. *Total phenolic and Flavonoid content*

Total phenolic and flavonoid contents were evaluated in all crude extract as described previously (Bahadori et al., 2017;Grochowski et al., 2017) by means of a well-known procedure as follow.

 For thetotal phenolic content, 0.25 mL of the sample solution (1 mg/mL) was mixed with the diluted (1:9, v/v) Folin–Ciocalteu reagent (1 mL). 0.75 mL of a Na2CO3 solution (1%) was added after 3 min, and then the sample absorbance was read at 760 nm after 2 h of incubation at room temperature.

For thetotal flavonoid content, 1 mL of the sample solution (1 mg/mL) was mixed with an equal volume of aluminium chloride (2%) solution in methanol, and the absorbance was read at 415 nm after 10 min of incubation at room temperature. The outcomes of total phenolic constituents were reported as equivalents of gallic acid (mg GAE/g extract), while the results of total flavonoid constituents were recorded as equivalents of quercetin (mg QE/g extract).

* 1. *UHPLC-MS-Instrumentation*

Secondary metabolites were evaluated by RP-UHPLC-MS. UHPLC of Agilent 1290 Infinity LC system coupled to Agilent 6520 Accurate-Mass Q-TOF mass spectrometer with dual ESI source was used. Column specifications were as: Agilent Zorbax Eclipse XDB-C18, narrow-bore 2.1 x 150 mm, 3.5 micron (P/N: 930990-902). Column and auto-sampler temperature were maintained at 25 °C and 4 °C, respectively. Flow rate was 0.5 mL/min. Mobile phases used were: A - 0.1% formic acid in water, B - 0.1% formic acid in acetonitrile. Injection volume was 1.0 µL. Run time was 25 min and post-run time was 5 min. Full scan MS analysis was done over a range of *m/z* 100-1000 using electrospray ion source in negative mode. Nitrogen was supplied as nebulizing and drying gas at flow rates of 25 and 600 L/hour, respectively. The drying gas temperature was 350°C. The fragmentation voltage was optimized to 125 V. Analysis was performed with a capillary voltage of 3500 V. Data was processed with Agilent Mass Hunter Qualitative Analysis B.05.00 (Method: Metabolomics-2017- 00004.m). Identification of compounds was done from Search Database: METLIN\_AM\_PCDL-N- 170502.cdb, with parameters as: Match tolerance: 5 ppm, Positive Ions: +H, +Na, +NH4, Negative Ions: -H (Saleem et al., 2019).

* 1. *HPLC-PDA polyphenolic quantification*
		1. *HPLC conditions*

HPLC analyses were performed on a Waters liquid chromatograph equipped with a model 600 solvent pump and a 2996 photodiode array detector, and Empower v.2 Software (Waters Spa, Milford, MA, USA) was used for the acquisition of data. A C18 reversed-phase packing column (Prodigy ODS(3), 4.6 × 150 mm, 5 μm; Phenomenex, Torrance, CA, USA) was used for the separation, and the column was thermostated at 301°C using a Jetstream2 Plus column oven. The UV/Vis acquisition wavelength was set at 200–500 nm. The quantitative analyses were achieved at the maximum wavelength for each compound.

The injection volume was 20 μL. The mobile phase was directly online degassed by using Biotech DEGASi, mod. Compact (LabService, Anzola dell’Emilia, Italy). Gradient elution was performed using the mobile phase water-acetonitrile (93:7, v/v, 3% acetic acid) as reported in the literature (Locatelli et al., 2017;Di Sotto et al., 2018). All the prepared sample solutions were centrifuged, and the supernatant was injected into HPLC.

* + 1. *Preparation of standard solutions and samples*

The stock solutions of phenolics were made at a concentration of 1 mg/mL in a final volume of 10 mL of methanol. Working solutions of mixed standards at the concentrations of 10, 25, 50, 75, 100, 150 and 200 μg/mL were made by dilution of stock solution in volumetric flasks with the mobile phase. Then the standards were injected into the HPLC-UV/Vis system. Working solutions of mixed standards at the concentrations of 0.25, 0.5, 1, 2.5, 5, 10, and 20 μg/mL were made by dilution of stock solution in volumetric flasks with the mobile phase. Then the standards were injected into the HPLC-UV/Vis system. Each solid sample was weighted and solubilized in mobile phase in 1:1 (*w:v*) ratio. In this case, the obtained concentrations (μg/mL) correspond to the total amount (μg/mg). After solubilization, the sample was centrifuged at 12000 x *g* before HPLC injection. Into the tables, BLD<0.1 μg/mL while BLQ<0.2 μg/mL.

1. ***Antioxidant assays***

The free radical scavenging (DPPH, ABTS), reducing power (FRAP, CUPRAC) phosphomolybdenum (total antioxidant capacity), and metal chelating (ferrous ion chelation) were evaluated following the previous methods as described by Grochowski et al. (2017) (Grochowski et al., 2017).

* 1. *Free radical scavenging activity (DPPH)*

1 mL of the sample solution was added to the DPPH solution (0.267 mM 4 mL, 0.004% methanol solution), and after 30 min of incubation, the absorbance was recorded at 517 nm. Milligrams of Trolox equivalents per gram of dry extract (TE/g extract) were the measure unit.

* 1. *ABTS radical cation scavenging activity*

ABTS+ radical cation was obtained following the reaction between 7 mM ABTS solution and 2.45 mM potassium persulfate. 1 mL of the test solution was mixed with 2 mL of ABTS solution and after 30 min the absorbance was recorded at 734 nm. The results were expressed as milligrams of Trolox equivalents per gram of dry extract (TE/g extract).

*2.3. Phosphomolybdenum method*

0.3 mL of the sample solutions were mixed with 3 mL of reagent solution (0.6 M sulfuric acid, 28 mM sodium phosphate and 4 mM ammonium molybdate), and after 90 min the absorbance was recorded at 695 nm. Millimoles of Trolox equivalents per gram of dry extract (TE/g extract) were the measurement unit.

*2.4. Cupric ion reducing (CUPRAC) method*

0.5 mL of the sample solutions were mixed with [CuCl2 (1 mL, 10 mM), neocuproine (1 mL, 7.5 mM), NH4Ac buffer (1 mL, 1 M, pH 7.0)] and after 30 min the absorbance was recorded at 450 nm. Milligrams of Trolox equivalents per gram of dry extract (TE/g extract) were the measurement unit.

*2.5. Ferric reducing antioxidant power (FRAP) method*

0.1 mL of the sample solution was added to reagent (2 mL) in acetate buffer (0.3 M, pH 3.6), 2,4,6-tris(2-pyridyl)-s-triazine (TPTZ) (10 mM) in 40 mM HCl and ferric chloride (20 mM) in a final ratio of 10:1:1 (v/v/v) and the absorbance was recorded at 593 nm 30 min. Milligrams of Trolox equivalents per gram of dry extract (TE/g extract) were the measurement unit.

*2.6. Metal chelating activity on ferrous ions*

2.0 mL of the sample solution were added to 0.05 mL of a solution of FeCl2 (2 mM), then the reaction was initiated using 0.2 mL of ferrozine (5 mM). After 10 min the absorbance was recorded at 562 nm and the results expressed as milligrams of EDTA equivalents per gram of dry extract (EDTAE/g extract).

1. ***Enzyme inhibition assays***

 The possible enzyme inhibitory capacity of all the concentrates against acetylcholinesterase (AChE), butyrylcholinesterase (BChE), tyrosinase, and α-amylase were explored utilizing earlier standard *in-vitro* bio-assays (Grochowski et al., 2017).

* 1. *Cholinesterase*

The reaction mixture composed by the sample solution (50 μL), DTNB (3 mM 125 μL) and enzyme solution (0.265 u/mL AChE or 0.026 u/mL BChE) solution (25 μL) in Tris-HCl buffer (pH 8.0) was added to the substrates [acetylthiocholine iodide (15 mM ATCI) or butyrylthiocholine chloride (1.5 mM BTCl, 25 μL)]. After 15 min of incubation, the absorbance was recorded at 405 nm and the results expressed as milligrams of galantamine equivalents per gram of dry extract (GALAEs/g extract).

* 1. *α-Amylase*

The reaction mixture composed by 25 L of the sample solution and 50 μL of the α-amylase solution (10 u/mL) in phosphate buffer (pH 6.9 with 6 mM sodium chloride) was added to 50 μL of the starch solution (0.05%) and the reaction was stopped with the addition of 25 μL of HCl (1 M). Then 100 μL of the iodine-potassium iodide solution was added. After 10 min of incubation, the absorbance was recorded at 630 nm and the results expressed as millimoles of acarbose equivalents per gram of dry extract (ACAEs/g extract).

* 1. *Tyrosinase*

25 μL of the sample solution were added to a 40 μL of tyrosinase solution (200 u/mL) and phosphate buffer (40 mM, 100 μL, pH 6.8) in a 96-well microplate and incubated for 15 min at 25 °C. Then the reaction was initiated by adding l-DOPA (10 mM, 40 μL). After 10 min of incubation at room temperature, the absorbance was recorded at 492 nm and the results expressed as milligrams of kojic acid equivalents per gram of dry extract (KAE/g extract)

**Table S1. Gradient elution program used for HPLC analyses**

|  |  |  |  |
| --- | --- | --- | --- |
| **TIME (min)** | **FLOW (mL min-1)** | **%A** | **%B** |
| 0 | 1 | 93 | 7 |
| 0.1 | 93 | 7 |
| 30 | 72 | 28 |
| 38 | 75 | 25 |
| 45 | 2 | 98 |
| 47 | 2 | 98 |
| 48 | 93 | 7 |
| 58 | 93 | 7 |

**Table S2: Analytes, retention times, and maximum wavelengths used for quantitative analyses.**

|  |  |  |
| --- | --- | --- |
| **Analytes** | **Retention Times (min)** | **λ max** |
| Gallic acid | 4.99 | 271 nm |
| Catechin | 13.36 | 278 nm |
| Chlorogenic acid | 14.29 | 324 nm |
| 4-hydroxybenzoic acid | 14.71 | 256 nm |
| Vanillic acid | 17.31 | 260 nm |
| Epicatechin | 18.30 | 278 nm |
| Syringic acid | 18.50 | 274 nm |
| 3-hydroxybenzoic acid | 19.41 | 275 nm |
| 3-hydroxy-4-methoxybenzaldehyde | 22.08 | 278 nm |
| *p-*coumaric acid | 22.65 | 310 nm |
| Rutin | 25.38 | 256 nm |
| Sinapinic acid | 26.18 | 324 nm |
| *t*-ferulic acid | 27.75 | 315 nm |
| Naringin | 29.78 | 285 nm |
| 2,3-dimethoxybenzoic acid | 30.36 | 299 nm |
| Benzoic acid | 31.20 | 275 nm |
| *o*-coumaric acid | 34.81 | 276 nm |
| Quercetin  | 40.57 | 367 nm |
| Harpagoside | 45.49 | 280 nm |
| *t*-cinnamic acid | 45.87 | 276 nm |
| Naringenin | 46.74 | 290 nm |
| Carvacrol | 49.95 | 275 nm |

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**Figure 1. Chemical standards chromatogram for the 22 analytes.**

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